



Dr. BGR
Publications

Aquaculture: Breeding, Disease management and Sustainable practices

Editors

Dr. M.Sini Margret
Dr. I.Vasudhevan
Dr. M.Santhiya
Dr. S.Rinna Hamlin

2026




Aquaculture: Breeding, Disease management and Sustainable practices

Editors

Dr. M.Sini Margret
Dr. I.Vasudhevan
Dr. M.Santhiya
Dr. S.Rinna Hamlin

2026

Verso Page

Publisher	Dr. BGR Publications India Tamil Nadu Tuticorin ✉ drbgrpublications@gmail.com 📖 https://drbgrpublications.in/books/ 📱 https://www.instagram.com/drbgrpublications/
Book Type	Edited Volume (Collection of 10 Articles)
Title	Aquaculture: Breeding, Disease Management and Sustainable Practices
ISBN	978-81-69360-86-9
Page Size	A4
Language	English
Product Form	Digital Download and Online
Date of Publication	26-06-2026
Editor	Dr. M.Sini Margret
Co-Editors	Dr. I.Vasudhevan Dr. M.Santhiya Dr. S.Rinna Hamlin
Design & Typesetting	Dr. BGR Publications
Cover design credit	Dr. B.Govindarajan
Digital Production Line	This book is published in digital format and made available globally through open access platforms.
Disclaimer	The author is fully responsible for the content of this book. The publisher disclaims all liability for errors, omissions, inaccuracies, plagiarism, or interpretations. Unintentional errors may be reported to the author or publisher for correction in future editions.
Copyright Notice	© 2026 The Editors and Individual Chapter Authors This book is an Open Access publication. All chapters are distributed under the terms of the Creative Commons Attribution 4.0 International License (CC BY 4.0), which permits use, sharing, adaptation, distribution, and reproduction in any medium, provided appropriate credit is given to the author(s) and the source. The editors retain copyright over the editorial content and compilation of this book. Individual authors retain copyright of their respective chapters.
Jurisdiction Clause	Any disputes arising from this publication shall be the sole responsibility of the author(s). The publisher shall not be held liable for any legal claims, disputes, or consequences related to the content of this book.
Barcode	 ISBN 978-81-69360-86-9 9 788169 360869

An imprint of Dr. BGR Publications



Mr. C. Rajan
Secretary
Vivekananda Educational Society
Agasteeswaram, Kanyakumari
Joint Action Council for Aided and
Self-Financing Colleges,
Manonmaniam Sundaranar University,
Thirunelveli



FOREWORD MESSAGE

I am glad to note **Dr. I. Vasudhevan** from the Department of Zoology, Vivekananda College, Agasteeswaram, Kanyakumari District has written the book entitled “**Aquaculture: Breeding, Disease management and Sustainable practices**”. Which is very interesting and needed to the Students, Researchers and Fish farmers. The book contains different chapters including Fish breeding, Live feed culture and Disease management

Ornamental fishes are called “Living jewels” due to their color, shape, behavior and origin. They are kept as pets in the confined spaces of an aquarium for the purpose of enjoying their beauty or as a matter of fancy. They have immense commercial value because of their aesthetic value. Ornamental fish keeping is one of the most popular hobbies in the world today next to photography. Ornamental fish can be grouped in different ways. They are “egg-laying” species and “live-bearing” species. Live-bearers are the fishes that do not lay eggs but give birth to young ones. Guppies, Platies, Mollies and Swordtail are the commonest examples of ovo-viviparous. Egg laying (oviparous) ornamental fishes show extreme parental care in nursing their young ones for a few days. Gold fish, Gourami, Bettas and Angels are the commonest examples of oviparous.

In the trade, there are 2% coldwater and 98% tropical fish; in addition, 90% are of fresh water origin, 10% marine and 0.1% brackish water. Singapore is one of the largest exporters of tropical aquarium fishes, valued as U.S. \$ 40-50 million dollars, contributing to an extent of 23.3% to the world market. The other exporting countries include, Hong Kong 7.6%, Thailand 7%, Indonesia 6.9%, Philippines 5.1%, Malaysia 3.3%, Japan 2.5% and other countries 42.9%. It exports to over 50 countries worldwide and its export value is over U.S. \$ 20 million annually. Villages in Amtolla area and Howrah near Calcutta and Kolathur near Madras are worth mentioning. In these villages a person owning about 30 small tanks earn about Rs. 12,000/- a month.

C. Rajan

Mr. C. Rajan
Secretary

List of Contributors

S. No.	Paper ID	Title	Page No.
1)	<i>Aqua-02</i>	Role of Probiotics in Aquaculture <i>Dr. S. J. Sreeja</i>	1
2)	<i>Aqua-03</i>	Recent Advances in Aquaculture: Technological Innovations for Sustainable Production <i>Subha T</i>	9
3)	<i>Aqua-04</i>	Climate Change and Aquaculture in the Present Scenario: Drivers, Impacts, Adaptation, and Climate-Smart Strategies <i>Sreeya G. Nair</i>	20
4)	<i>Aqua-05</i>	Cichlids Breeding <i>Dr. I. Vasudhevan</i>	34
5)	<i>Aqua-06</i>	Live Jewels – Live Bearears <i>Subhitsha. S Roshini. M and Anisha Devi. B</i>	43
6)	<i>Aqua-07</i>	Management of Ornamental Fish Diseases <i>Jesi Nayaki. M, Sahaya Nithisha. C and Sheebha. S</i>	54
7)	<i>Aqua-08</i>	Marine Ornamental Fishes in India: Status, Problems and Management Strategies <i>Dr. A. Pushparaj and Dr. P. Ambika</i>	62
8)	<i>Aqua-09</i>	A review on use of Spirulina for ornamental fishes <i>Dr. S. Rinna Hamlin</i>	72
9)	<i>Aqua-10</i>	Pigment studies in ornamental fishes <i>Greeshma Thomas</i>	79
10)	<i>Aqua-11</i>	A Path to Smart Farming: Exploring Opportunities and Challenges of Artificial Intelligence (AI) in Aquaculture <i>V. Pattukumar</i>	92

Role of Probiotics in Aquaculture

S. J. Sreeja

Assistant Professor, PG & Research Department of Zoology, S. T. Hindu College, Nagercoil

Corresponding Author Email: sreejazoology@gmail.com

Abstract

Aquaculture has become a major contributor to global food production, but its rapid expansion has led to challenges such as disease outbreaks, environmental degradation, and overuse of antibiotics. Probiotics have emerged as an effective and eco-friendly solution to these issues. They are live microorganisms that provide health benefits to aquatic organisms by improving gut microbial balance, enhancing immunity, promoting growth, and maintaining water quality. Probiotics also help in reducing the dependence on antibiotics, thereby minimizing environmental and health risks. This chapter provides a comprehensive overview of probiotics in aquaculture, including their types, mechanisms of action, applications, advantages, and future prospects, highlighting their importance in sustainable aquaculture development.

Keywords: *Probiotics, ecofriendly, gut microbial balance, environmental health*

Introduction

Aquaculture is one of the fastest-growing sectors in food production, playing a crucial role in meeting the increasing global demand for fish and seafood. However, the intensification of aquaculture practices has resulted in several problems such as poor water quality, stress conditions, and frequent disease outbreaks. Traditionally, antibiotics and chemicals were used to control diseases, but their excessive use has led to antibiotic resistance, accumulation of harmful residues, and environmental pollution. These issues have necessitated the development of alternative approaches that are safe, sustainable, and environmentally friendly. Probiotics have gained importance in this context as they improve the health of cultured organisms and maintain ecological balance. Their application has become a key strategy in modern aquaculture systems.

Definition and Concept of Probiotics

Probiotics are defined as live microorganisms which, when administered in adequate amounts, confer health benefits to the host. In aquaculture, this concept is broader because aquatic organisms live in constant interaction with their surrounding water environment.

Therefore, probiotics not only improve the intestinal microbial balance but also influence the microbial composition of the water. They help in maintaining a balance between beneficial and harmful microorganisms, thereby improving both host health and environmental conditions. This dual role makes probiotics highly effective in aquaculture systems.

Types of probiotics used in Aquaculture

A wide range of microorganisms are used as probiotics in aquaculture. These include beneficial bacteria such as *Lactobacillus*, *Bifidobacterium*, *Streptococcus*, and *Enterococcus*. Spore-forming bacteria like *Bacillus subtilis* and *Bacillus licheniformis* are widely used due to their resistance to harsh environmental conditions. Yeasts such as *Saccharomyces cerevisiae* are also commonly used because they enhance digestion and immune responses. These microorganisms are carefully selected to ensure they are non-pathogenic and beneficial to the host.

Characteristics of ideal probiotics

An ideal probiotic should be non-pathogenic, safe, and capable of surviving the digestive conditions of the host. It should adhere to the intestinal lining and effectively colonize the gut. Additionally, it should produce antimicrobial substances and digestive enzymes that benefit the host. Stability during storage and ease of application are also important characteristics. These properties ensure that probiotics can function effectively in aquaculture systems.

Mechanisms of Action of Probiotics

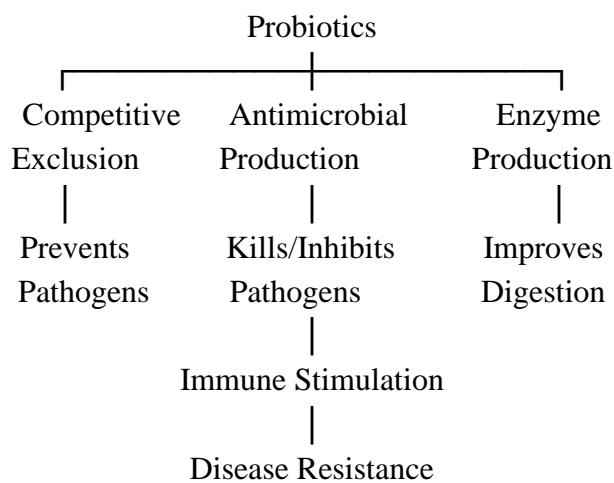


Figure 1: Mechanism of action of probiotics

Probiotics act through multiple mechanisms to benefit aquatic organisms. One of the most important mechanisms is competitive exclusion, where beneficial microorganisms compete with harmful pathogens for nutrients and attachment sites, thereby preventing their growth. Probiotics also produce antimicrobial substances such as bacteriocins, organic acids, and hydrogen peroxide that inhibit pathogenic microbes. Additionally, they produce digestive enzymes like proteases, amylases, and lipases, which enhance nutrient digestion and absorption. Probiotics also stimulate the immune system, improving the ability of aquatic organisms to resist diseases.

Role in nutrition and growth

Probiotics play an important role in improving growth and nutrition in aquaculture species. They enhance feed efficiency by breaking down complex nutrients into simpler forms, which can be easily absorbed. This results in better feed utilization and reduced wastage. Probiotics also synthesize essential nutrients such as vitamins and amino acids, contributing to improved growth performance and higher productivity.

Role in disease control

Disease outbreaks are a major concern in aquaculture. Probiotics help control diseases by inhibiting pathogenic microorganisms and boosting the immune system of the host. They reduce the occurrence of infections and improve survival rates. Unlike antibiotics, probiotics do not lead to resistance, making them a safer and more sustainable option.

Role in Water Quality Management

Water quality is a critical factor in aquaculture, as it directly affects the health and survival of aquatic organisms. Probiotics contribute to water quality management by decomposing organic matter such as uneaten feed and waste products. They reduce the accumulation of toxic substances such as ammonia, nitrite, and hydrogen sulfide. By maintaining a balanced microbial ecosystem, probiotics create a healthy environment that reduces stress and improves survival rates.

Applications and Methods of Use

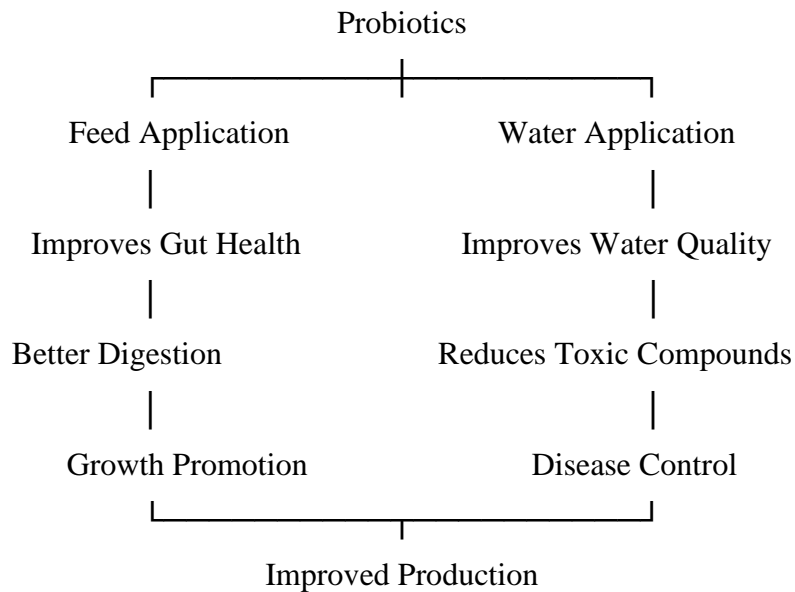


Figure 2: Application of probiotics in Aquaculture

Probiotics are widely used in fish farming, shrimp culture, and larval rearing systems. They can be applied through feed supplementation or directly into the water. Feed-based probiotics improve gut health, digestion, and growth performance, while water-based probiotics enhance environmental conditions and control harmful microorganisms. Their flexibility makes them suitable for various aquaculture practices.

Advanced Concepts: Gut Microbiota and Synbiotics

The gut microbiota of aquatic organisms plays a vital role in digestion, metabolism, and immunity. Probiotics help maintain a balanced microbial population by promoting beneficial bacteria and suppressing harmful ones. This leads to improved health and disease resistance. The concept of synbiotics, which combines probiotics and prebiotics, provides enhanced benefits by improving the survival and activity of beneficial microorganisms. These advanced approaches are increasingly being used in aquaculture.

Synbiotics and Their Importance

Synbiotics are a combination of probiotics and prebiotics that work together to provide enhanced benefits. Prebiotics are non-digestible food ingredients that promote the growth of beneficial microorganisms in the gut. When used together, synbiotics improve the survival and activity of probiotics, making them more effective. In aquaculture, synbiotics have been shown to enhance growth performance, improve immune responses, and increase

disease resistance. They also contribute to better feed utilization and overall health of aquatic organisms. The use of synbiotics is gaining popularity as an advanced strategy for improving aquaculture productivity.

Probiotics in Larval Rearing

Larval stages of aquatic organisms are highly sensitive and vulnerable to environmental stress and diseases. High mortality rates are common during this stage due to underdeveloped immune systems and unstable environmental conditions. Probiotics play a crucial role in larval rearing by improving survival rates and promoting healthy development. They help in establishing a beneficial microbial community in the rearing environment and prevent the growth of pathogenic bacteria. Probiotics also enhance the digestion of live feeds such as algae and rotifers, ensuring proper nutrition for larvae. Their use in hatcheries has shown significant improvements in larval quality and survival.

Additional Insights and Extended Concepts

Probiotics influence aquatic organisms at the molecular level by regulating gene expression related to immunity, metabolism, and stress response. They improve intestinal health by strengthening the gut lining and producing beneficial metabolites. In larval rearing systems, probiotics enhance survival and development by stabilizing microbial communities. They also play an important role in stress management by improving physiological stability and resistance to environmental changes.

In different aquaculture systems, probiotics contribute to improved productivity by maintaining ecological balance and enhancing nutrient utilization. They interact with feed to improve digestibility and reduce anti-nutritional factors. Probiotics are also used in biofloc systems, where they help recycle nutrients and maintain water quality. Their role in integrated aquaculture systems further enhances sustainability and efficiency.

Advantages of Probiotics

Probiotics offer several advantages, including improved growth rate, better feed conversion efficiency, enhanced immunity, and increased disease resistance. They reduce the need for antibiotics and chemicals, thereby minimizing environmental pollution. Probiotics also improve water quality and overall productivity, making them an essential component of sustainable aquaculture.

Challenges and Limitations

Despite their benefits, probiotics have certain limitations. Their effectiveness can vary depending on environmental conditions such as temperature, pH, and salinity. There is a lack of standardization in probiotic strains and application methods, leading to inconsistent results. Storage and shelf-life issues can affect their viability. Further research is needed to optimize their use and understand their interactions within aquaculture systems.

Factors Affecting Probiotic Efficiency

The effectiveness of probiotics in aquaculture depends on several factors. Environmental conditions such as temperature, pH, salinity, and dissolved oxygen levels can influence the activity and survival of probiotic microorganisms. The species of the host organism also plays a significant role, as different species respond differently to probiotics. The dosage and method of application are equally important, as insufficient or excessive use may not produce the desired results. Additionally, the quality and viability of probiotic strains during storage and handling can affect their performance. Therefore, careful management is required to achieve maximum benefits.

Probiotics and Stress Management

Stress is a major factor affecting the health and productivity of aquatic organisms. It can be caused by factors such as poor water quality, overcrowding, handling, and sudden environmental changes. Stress weakens the immune system and makes organisms more susceptible to diseases. Probiotics help in reducing stress by improving gut health, enhancing immune responses, and maintaining a stable internal environment. They also help in regulating physiological functions, which contributes to better stress tolerance and overall well-being of aquatic species.

Probiotics in Biofloc Technology

Biofloc technology is an advanced aquaculture system that relies on microbial communities to recycle nutrients and maintain water quality. Probiotics play a significant role in biofloc systems by promoting the growth of beneficial microorganisms. These microorganisms convert waste materials into useful microbial protein, which can be consumed by aquatic organisms as an additional food source. Probiotics help in maintaining the balance of microbial populations in biofloc systems, improving water quality, and enhancing production efficiency. Their integration into biofloc technology has shown promising results in sustainable aquaculture.

Safety and Regulatory Aspects

The use of probiotics in aquaculture requires proper safety evaluation to ensure that they do not pose any risk to the host, environment, or consumers. Probiotic strains should be non-pathogenic, non-toxic, and free from antibiotic resistance genes. Regulatory guidelines have been established in many countries to control the use of probiotics in aquaculture. These guidelines ensure the quality, safety, and effectiveness of probiotic products. Proper labeling, dosage instructions, and quality control measures are essential for the safe use of probiotics.

Economic Importance of Probiotics

Probiotics have significant economic benefits in aquaculture. By improving growth rates, feed efficiency, and survival rates, they help increase production and profitability. The reduction in disease outbreaks and the decreased need for antibiotics lower the cost of treatment and management. Improved water quality also reduces maintenance costs. Overall, the use of probiotics leads to more efficient and sustainable aquaculture operations, making them economically advantageous for farmers.

Research and Innovations

Ongoing research in probiotics is focused on discovering new strains with enhanced beneficial properties. Advanced techniques such as molecular biology and genomics are being used to study microbial communities and identify potential probiotic candidates. Innovations include the development of multi-strain probiotics, encapsulated probiotics for better delivery, and genetically improved strains. These advancements are expected to improve the effectiveness and reliability of probiotics in aquaculture systems.

Future Prospects

The future of probiotics in aquaculture is promising, with ongoing research focused on developing improved strains and formulations. Advances in biotechnology are helping identify new microorganisms with enhanced properties. The use of synbiotics, encapsulated probiotics, and multi-strain formulations is expected to improve efficiency. Probiotics will continue to play a key role in sustainable aquaculture development.

Conclusion

Probiotics have become an indispensable component of modern aquaculture due to their multiple benefits. They improve growth performance, enhance immunity, maintain water quality, and reduce the reliance on antibiotics. Their role in sustainable aquaculture is increasingly recognized, and their application is expanding with technological advancements.

Despite certain challenges, continued research and proper management practices can maximize their potential. Probiotics are expected to play a key role in the future of aquaculture by ensuring healthy production and environmental sustainability.

References

- 1) Akhter, N., Wu, B., Memon, A. M., & Mohsin, M. (2015). *Probiotics and prebiotics associated with aquaculture. Fish & Shellfish Immunology*, 45, 733–741.
- 2) Balcázar, J. L., de Blas, I., Ruiz-Zarzuela, I., Cunningham, D., Vendrell, D., & Múzquiz, J. L. (2006). *The role of probiotics in aquaculture. Veterinary Microbiology*, 114, 173–186.
- 3) Fuller, R. (1989). *Probiotics in man and animals. Journal of Applied Bacteriology*, 66, 365–378.
- 4) Gatesoupe, F. J. (1999). *The use of probiotics in aquaculture. Aquaculture*, 180, 147–165.
- 5) Hasan, K. N., & Banerjee, G. (2020). *Probiotics as beneficial mediators in aquaculture. Journal of Basic and Applied Zoology*, 81, 53.
- 6) Hoseinifar, S. H., Sun, Y. Z., Wang, A., & Zhou, Z. (2018). *Probiotics as means of disease control in aquaculture. Frontiers in Microbiology*, 9, 2429.
- 7) Irianto, A., & Austin, B. (2002). *Probiotics in aquaculture. Journal of Fish Diseases*, 25, 633–642.
- 8) Kesarcodi-Watson, A., Kaspar, H., Lategan, M. J., & Gibson, L. (2008). *Probiotics in aquaculture: The need, principles and mechanisms of action. Aquaculture*, 274, 1–14.
- 9) Moriarty, D. J. W. (1998). *Control of luminous Vibrio species in penaeid aquaculture ponds. Aquaculture*, 164, 351–358.
- 10) Nayak, S. K. (2010). *Probiotics and immunity: A fish perspective. Fish & Shellfish Immunology*, 29, 2–14.
- 11) Ninawe, A. S., & Selvin, J. (2009). *Probiotics in shrimp aquaculture: Avenues and challenges. Critical Reviews in Microbiology*, 35, 43–66.
- 12) Pandiyan, P., Balaraman, D., Thirunavukkarasu, R., et al. (2013). *Probiotics in aquaculture. Drug Invention Today*, 5, 55–59.
- 13) Verschuere, L., Rombaut, G., Sorgeloos, P., & Verstraete, W. (2000). *Probiotic bacteria as biological control agents in aquaculture. Microbiology and Molecular Biology Reviews*, 64, 655–671.
- 14) Wang, Y. B., Li, J. R., & Lin, J. (2008). *Probiotics in aquaculture: Challenges and outlook. Aquaculture*, 281, 1–4.
- 15) Zorriehzahra, M. J. et al. (2016). *Probiotics as beneficial microbes in aquaculture: Modes of action. Veterinary Quarterly*, 36, 228–241.

Recent Advances in Aquaculture: Technological Innovations for Sustainable Production

Subha T

Assistant Professor in Biotechnology, Department of Allied Health Sciences, NICHE, Kumarakoil, Kanyakumari District, Tamil Nadu.

Corresponding Author Email: subhabio86@gmail.com

Abstract

Aquaculture has emerged as a rapidly expanding sector contributing significantly to global food security, nutritional supply, and economic development. However, conventional aquaculture practices are often associated with environmental degradation, disease outbreaks, and inefficient resource utilization. To overcome these challenges, recent technological advancements have been introduced, transforming aquaculture into a more sustainable and efficient system. Innovations such as biotechnology, genetic engineering, nanotechnology, and smart aquaculture systems incorporating Internet of Things (IoT), sensors, and artificial intelligence (AI) have revolutionized the industry. These technologies not only enhance production and growth rates but also improve disease management, optimize feed utilization, and reduce environmental impact. This chapter discusses the major advancements in aquaculture technologies, their applications, benefits, and future prospects in achieving sustainable aquaculture development.

Keywords: *Aquaculture, Biotechnology, Genetic engineering, Nanotechnology, Smart aquaculture, IoT, Artificial Intelligence, Sustainability*

Introduction

Aquaculture refers to the controlled cultivation of aquatic organisms such as fish, shellfish, and aquatic plants in freshwater, brackish water, or marine environments. It plays a crucial role in meeting the increasing global demand for protein-rich food, especially as capture fisheries are declining due to overexploitation. Aquaculture has expanded significantly due to technological advancements and improved farming practices. Despite its rapid growth, traditional aquaculture faces several challenges, including poor water quality, disease outbreaks, high feed costs, and environmental pollution. These issues have highlighted the need for innovative approaches to improve productivity and sustainability.

Modern aquaculture systems are now integrating advanced scientific and technological tools to address these problems effectively.

In recent years, rapid population growth and changing dietary preferences have increased the demand for aquatic products. This has led to the intensification of aquaculture practices, where higher stocking densities and improved feeding strategies are employed to maximize production. However, intensification has also resulted in several challenges such as water pollution, disease outbreaks, environmental degradation, and resource depletion. These issues have highlighted the need for sustainable and innovative approaches in aquaculture development. Technological advancements have transformed aquaculture from a traditional practice into a modern, science-based industry. The integration of biotechnology, genetic engineering, nanotechnology, and digital technologies has significantly improved production efficiency and environmental sustainability. These innovations enable better monitoring, control, and management of aquaculture systems, ensuring optimal growth conditions and reducing risks.

Recent advances in aquaculture focus on improving water quality management, enhancing feed efficiency, and developing disease-resistant species. Techniques such as biofloc technology, recirculating aquaculture systems, and integrated multi-trophic aquaculture have been introduced to promote sustainable production. These systems aim to minimize waste, recycle nutrients, and reduce environmental impact while maintaining high productivity. The adoption of smart aquaculture technologies has further revolutionized the industry by enabling real-time monitoring and automation. The use of sensors, Internet of Things (IoT), and artificial intelligence allows farmers to track environmental parameters, predict potential problems, and make informed decisions. This has led to increased efficiency, reduced operational costs, and improved fish health and welfare.

In addition to technological innovations, there is growing emphasis on the use of alternative and sustainable feed resources to reduce dependence on fishmeal and fish oil. The development of eco-friendly feed ingredients such as plant-based proteins, insect meal, and microalgae is gaining importance in modern aquaculture practices. Furthermore, climate change has become a major concern affecting aquaculture systems through temperature fluctuations, changes in water availability, and increased frequency of extreme weather events. As a result, climate-resilient aquaculture practices are being developed to ensure long-

term sustainability and productivity. Overall, recent advances in aquaculture are focused on achieving a balance between increased production and environmental conservation. By integrating innovative technologies and sustainable practices, aquaculture is evolving into a highly efficient and environmentally responsible sector capable of meeting future global food demands. The following are the recent advances in aquaculture.

1. Biotechnology in Aquaculture

Biotechnology has become an essential component in modern aquaculture, offering innovative solutions for improving fish health, growth, and overall productivity. It involves the application of biological processes, molecular tools, and microbial technologies to enhance aquaculture practices. One of the major contributions of biotechnology is in disease diagnosis and prevention.

i) **Probiotics and prebiotics** are increasingly used in aquaculture to improve gut health, enhance immune response, and promote growth in cultured species. These beneficial microorganisms help maintain a balanced microbial environment in the gut and reduce the occurrence of diseases. Enzyme technology is also applied to improve feed digestion and nutrient absorption, leading to better feed conversion efficiency. Biotechnological advancements also support the production of functional feeds enriched with vitamins, probiotics, and immunostimulants. These feeds enhance growth performance, improve immune response, and increase resistance to diseases.

ii) **Molecular markers**, are used to identify genetic variations and select superior breeding stock. These markers help in accelerating breeding programs by enabling precise selection of desirable traits such as rapid growth, disease resistance, and adaptability to environmental stress.

iii) **Molecular techniques** such as PCR and DNA-based assays allow early detection of pathogens, enabling timely intervention and reducing mortality rates. Another significant application is in the field of genomic and transcriptomic studies, where researchers analyze the genetic makeup and gene expression patterns of aquatic organisms. These studies provide valuable insights into growth mechanisms, immune responses, and stress tolerance, thereby aiding in the development of improved aquaculture species.

iv) **Vaccination** strategies have also been developed to protect fish against bacterial and viral diseases, thereby minimizing economic losses. Biotechnology also plays a vital role in immunological advancements, such as the development of DNA vaccines and recombinant vaccines. These vaccines provide long-lasting protection against infectious diseases and reduce the need for antibiotics, thereby minimizing environmental contamination and the risk of antibiotic resistance.

iv) **Cell culture techniques** are used for the propagation of aquatic cells and tissues for research purposes, including disease studies and vaccine production. These techniques enable controlled experimentation and help in understanding host-pathogen interactions at a cellular level.

vi) **Microbial biotechnology** has gained importance in aquaculture through the use of bioaugmentation and bioremediation. Beneficial microorganisms are introduced into aquaculture systems to degrade organic waste, improve water quality, and maintain a stable microbial balance. This reduces the accumulation of harmful substances such as ammonia and nitrites. Furthermore, biotechnology contributes to the development of biofertilizers and live feeds, such as microalgae and zooplankton, which are essential for larval rearing in aquaculture. These live feeds provide essential nutrients and improve survival rates during early developmental stages.

vi) **Metagenomics** an emerging area which involves the study of microbial communities present in aquaculture environments. Understanding these microbial populations helps in optimizing culture conditions and preventing disease outbreaks.

Overall, biotechnology provides a scientific foundation for improving aquaculture practices by enhancing productivity, ensuring environmental sustainability, and promoting the health and welfare of aquatic organisms. Continuous research and innovation in this field are expected to further strengthen the aquaculture sector in the future.

2. Genetic Engineering and Selective Breeding

Genetic improvement is a powerful approach used in aquaculture to enhance desirable traits in cultured species. It includes both traditional selective breeding and advanced genetic engineering techniques.

i) **Selective breeding** involves choosing individuals with superior traits such as rapid growth, disease resistance, and high reproductive efficiency. Over successive generations, these traits are enhanced, resulting in improved stock performance and higher yield. This method is widely used in aquaculture due to its effectiveness and relatively low cost. Genetic engineering, on the other hand, involves direct manipulation of the genetic material of organisms.

ii) **Advanced techniques** such as transgenesis and gene editing enable scientists to introduce or modify specific genes to achieve desired traits. For example, fish can be genetically modified to grow faster, tolerate environmental stress, or resist diseases. The use of gene-editing tools like CRISPR has revolutionized genetic research in aquaculture by allowing precise and targeted modifications.

However, the application of genetic engineering raises ethical concerns related to environmental safety, biodiversity conservation, and public acceptance. Regulatory frameworks are also required to ensure safe and responsible use of these technologies. Despite these challenges, genetic improvement techniques significantly contribute to increased productivity, improved efficiency, and enhanced sustainability in aquaculture systems.

3. Nanotechnology Applications in Aquaculture

Nanotechnology is an emerging field that deals with materials at the nanoscale and offers innovative solutions for aquaculture development. Its applications are diverse and contribute to improved efficiency, health management, and environmental sustainability. In aquaculture, nanotechnology is used in feed formulation to enhance nutrient delivery and absorption.

i) **Nano-sized feed additives** in aquaculture are gaining attention for their ability to improve nutrient absorption and feed efficiency in cultured species. These additives enhance growth performance by delivering nutrients at the cellular level with higher bioavailability. They can also strengthen immune responses, reducing the incidence of diseases in fish and shrimp. Furthermore, nano-additives help minimize feed wastage and environmental pollution by improving digestion and nutrient utilization. The use of nanotechnology in aquaculture thus represents a promising strategy for sustainable and cost-effective fish farming.

ii) **Nanomaterials** are widely used in water treatment to remove harmful substances such as heavy metals, toxins, and pathogens. This improves water quality and creates a healthier environment for aquatic organisms. Nanotechnology also enables the development of advanced filtration systems for efficient water purification. They help improve growth performance, feed conversion efficiency, and overall health of aquatic organisms. However, careful evaluation of their toxicity and environmental impact is essential before large-scale application.

iii) In **disease management**, Nanotechnology plays a crucial role in disease management in aquaculture by enabling early detection, prevention, and targeted treatment of infections. Nano-based biosensors are used for rapid and sensitive identification of pathogens, helping in timely intervention before disease outbreaks spread. Nanoparticles with antimicrobial properties, such as silver and zinc oxide nanoparticles, are effective against a wide range of aquatic pathogens including bacteria, viruses, and fungi. These nanomaterials can be incorporated into feed or water to control infections more efficiently than conventional drugs. In addition, nano-encapsulation techniques allow controlled and targeted delivery of vaccines, antibiotics, and immunostimulants, reducing dosage frequency and minimizing side effects. This improves the immune response of fish and shrimp while lowering the risk of drug resistance.

iv) **Nano-based biosensors** are advanced analytical devices that combine nanomaterials with biological recognition elements to detect specific substances with high sensitivity and accuracy. These biosensors utilize nanoparticles, nanotubes, or nanowires to enhance signal detection due to their large surface area and unique electrical properties. In aquaculture, nano-based biosensors are used for the rapid detection of pathogens such as bacteria, viruses, and parasites in water and aquatic organisms. They enable early diagnosis of diseases, helping farmers take timely preventive measures and reduce mortality rates. Additionally, nano-based biosensors are portable, cost-effective, and require minimal sample preparation, making them suitable for field-level applications. Thus, they play a vital role in improving disease management and sustainability in aquaculture.

Overall, nanotechnology enhances aquaculture productivity while reducing environmental impact, making it a promising tool for sustainable aquaculture development.

4. Smart Aquaculture

Smart aquaculture represents a modern, technology-driven approach that integrates digital tools to improve the efficiency, productivity, and sustainability of aquaculture systems. It is often referred to as precision aquaculture, as it allows accurate monitoring and control of environmental and biological parameters. By using advanced technologies, farmers can manage aquaculture operations more effectively and reduce uncertainties associated with traditional practices.

i) The **Internet of Things (IoT)** plays a central role in smart aquaculture by connecting various devices, sensors, and systems through a network. IoT-enabled platforms continuously collect data from aquaculture environments, including parameters such as water temperature, pH, dissolved oxygen, salinity, turbidity, and ammonia levels. This real-time data is transmitted to centralized systems or cloud-based platforms, allowing farmers to monitor conditions remotely using smartphones or computers.

ii) **Sensors** are essential components of smart aquaculture systems, as they provide accurate and continuous measurements of environmental parameters. Different types of sensors are used depending on the requirements of the culture system. For example, dissolved oxygen sensors help maintain adequate oxygen levels, while temperature sensors ensure optimal thermal conditions for fish growth. Advanced sensors can also detect toxic substances and changes in water chemistry, enabling early intervention to prevent losses.

iii) **Artificial intelligence (AI)** enhances the efficiency of aquaculture systems by analyzing large volumes of data collected through IoT devices. AI algorithms and machine learning models can identify patterns and predict future conditions, helping farmers make informed decisions. For instance, AI can predict disease outbreaks based on environmental changes and fish behavior, allowing preventive measures to be taken in advance.

iv) **AI-driven systems** are also used to optimize feeding strategies by determining the appropriate quantity and timing of feed. Automated feeding systems, guided by AI, ensure that fish receive adequate nutrition without overfeeding, thereby reducing feed wastage and water pollution. In addition, computer vision technologies are used to monitor fish behavior, growth, and health status through image and video analysis.

v) **Smart aquaculture** systems also include automated control mechanisms that adjust environmental conditions based on sensor data. For example, aeration systems can be automatically activated when dissolved oxygen levels fall below a certain threshold. Similarly, water circulation and filtration systems can be regulated to maintain optimal water quality.

vi) **Cloud computing and data analytics** play a significant role in storing and processing large datasets generated by smart aquaculture systems. These technologies enable long-term analysis, performance evaluation, and predictive modeling. Farmers can use these insights to improve management practices and increase productivity.

Despite its advantages, smart aquaculture faces challenges such as high initial investment, technical complexity, and the need for reliable internet connectivity. However, ongoing advancements in technology and decreasing costs are expected to make these systems more accessible in the future. Overall, smart aquaculture represents a significant advancement in the aquaculture industry, combining digital technologies and biological knowledge to create efficient, sustainable, and intelligent farming systems. It plays a crucial role in meeting the increasing demand for aquatic products while ensuring environmental protection and resource conservation.

5. Advantages of Recent Technologies in Aquaculture

The adoption of modern technologies in aquaculture offers several benefits. These include increased production efficiency, improved growth rates, and enhanced disease resistance. Advanced monitoring systems ensure optimal environmental conditions, reducing stress and mortality in aquatic organisms. Technological innovations also promote efficient use of resources such as water and feed, minimizing wastage and environmental impact. Automation and digital monitoring reduce labor requirements and improve management efficiency. Overall, these advancements contribute to sustainable and profitable aquaculture practices.

6. Challenges and Limitations

Despite the numerous advantages, the implementation of advanced technologies in aquaculture faces several challenges. High initial investment costs can be a major barrier, especially for small-scale farmers. Technical complexity and the need for skilled personnel

also limit widespread adoption. Additionally, issues related to data reliability, infrastructure, and maintenance of advanced systems can affect performance. Ethical concerns associated with genetic engineering and environmental risks must also be carefully addressed. Regulatory frameworks and proper guidelines are necessary to ensure safe and responsible use of these technologies.

7.Future Prospects

The future of aquaculture lies in the integration of advanced technologies to create sustainable and efficient farming systems. Emerging concepts such as precision aquaculture, biofloc systems, and recirculating aquaculture systems are expected to play a significant role in the industry. Continuous research and development will lead to further innovations in biotechnology, nanotechnology, and digital technologies. The integration of renewable energy sources and eco-friendly practices will also enhance sustainability. With the growing demand for aquatic products, these advancements will help meet global food requirements while ensuring environmental protection and economic viability.

8.Conclusion

Recent advances in aquaculture have transformed traditional farming practices into modern, technology-driven systems. Innovations in biotechnology, genetic engineering, nanotechnology, and smart aquaculture have significantly improved productivity, efficiency, and sustainability.

These technologies address major challenges such as disease management, resource utilization, and environmental impact. As research continues, aquaculture is expected to become more advanced, sustainable, and capable of meeting future global food demands.

References

- 1) Ahmad, A., Abdullah, S. R. S., Hasan, H. A., Othman, A. R., & Ismail, N. I. (2021). *Aquaculture industry: Supply and demand, best practices, effluent and its current issues and treatment technology. Journal of Environmental Management, 287, 112271.*
- 2) Ahmed, M. S., Aurpa, T. T., & Azad, M. A. K. (2022). *Fish disease detection using image-based machine learning technique in aquaculture. Journal of King Saud University - Computer and Information Sciences, 34, 5170–5182.*

- 3) Alghamdi, M., & Haraz, Y. G. (2025). *Smart biofloc systems: leveraging artificial intelligence (AI) and Internet of Things (IoT) for sustainable aquaculture practices. Processes, 13(7), 2204.*
- 4) Badiola, M., Mendiola, D., & Bostock, J. (2012). *Recirculating aquaculture systems (RAS) analysis: main issues on management and future challenges. Aquacultural Engineering, 51, 26–35.*
- 5) Boyd, C. E., D'Abramo, L. R., Glencross, B. D., Huyben, D. C., Juarez, L. M., Lockwood, G. S., McNevin, A. A., Tacon, A. G. J., Teletchea, F., Tomasso Jr, J. R., Tucker, C. S., & Valenti, W. C. (2020). *Achieving sustainable aquaculture: Historical and current perspectives and future needs and challenges. Journal of the World Aquaculture Society.* Zlaugotne, B., et al. (2022). *Alternative feed ingredients in aquaculture: sustainability and challenges. Sustainability, 14(3), 1234.*
- 6) Boyd, C. E., McNevin, A. A., & Tucker, C. S. (2019). *Resource use and the environment. In J. S. Lucas, P. C. Southgate, & C. S. Tucker (Eds.), Aquaculture: Farming aquatic animals and plants (3rd ed., pp. 93–112). Wiley.*
- 7) Chandran, P. J. I., Khalil, H. A., Hashir, P. K., & Veerasingam, S. (2025). *Smart technologies in aquaculture: An integrated IoT, AI, and blockchain framework for sustainable growth. Aquacultural Engineering, 111, 102584.*
- 8) Crab, R., Defoirdt, T., Bossier, P., & Verstraete, W. (2012). *Biofloc technology in aquaculture: Beneficial effects and future challenges. Aquaculture, 356–357, 351–356.*
- 9) De Schryver, P., Crab, R., Defoirdt, T., Boon, N., & Verstraete, W. (2008). *The basics of biofloc technology: the added value for aquaculture. Aquaculture, 277(3–4), 125–137.*
- 10) Fajardo, C., Martinez-Rodriguez, G., Blasco, J., Mancera, J. M., Thomas, B., & De Donato, M. (2022). *Nanotechnology in aquaculture: Applications, perspectives and regulatory challenges. Aquaculture and Fisheries, 7(2), 185–200.*
- 11) Gutási, A., Hammer, S. E., El-Matbouli, M., & Saleh, M. (2023). *Recent applications of gene editing in fish species and aquatic medicine. Animals, 13(7), 1250.*
- 12) Lovejan, M., Jayachandran, P. R., & Rahiman, K. M. (2025). *Enhancing biofloc system performance using fermented plant biomass in tilapia culture. Aquaculture International.*

- 13) M. Ashraf, M. Aklakur, R. Sharma, A. Ahmad, M. Khan (2011). *Nanotechnology as a novel tool in fisheries and aquaculture development: A review* *Iranica Journal of Energy and Environment*, 2 (3) pp. 258-261.
- 14) Malone, R. F., & Pfeiffer, T. J. (2006). *Rating fixed film nitrifying biofilters used in RAS*. *Aquacultural Engineering*, 34(3), 389–402.
- 15) Ranjan, D., Singh, S., Misra, V. K., Singh, C. P., & Kumar, D. (2026). *Recent advancements in sustainable aquaculture: innovative techniques and future prospects*. *Frontiers in Aquaculture*, 5, Article 1770106.

Climate Change and Aquaculture in the Present Scenario: Drivers, Impacts, Adaptation, and Climate-Smart Strategies

Sreeya G. Nair

Assistant Professor and Head, Department of Zoology, Sree Ayyappa College for Women, Chunkankadai, Affiliated to Manonmaniam Sundaranar University, Tirunelveli.

Corresponding Author Email: sreeyaanair@gmail.com

Abstract

Climate change is rapidly reshaping aquatic ecosystems and aquaculture systems across the globe. Variations in temperature, ocean chemistry, hydrological cycles, and the increasing frequency of extreme weather events are significantly influencing aquaculture productivity, stability, and sustainability. Aquaculture plays a crucial role in global food security, contributing a substantial share of aquatic food production and supporting millions of livelihoods, particularly in developing regions. However, its heavy dependence on environmental conditions makes it highly vulnerable to climate variability and long-term climatic shifts. This review provides a comprehensive analysis of the major climate change drivers affecting aquaculture, including global warming, ocean acidification, salinity fluctuations, and altered precipitation patterns. It further examines the biological, ecological, and socio-economic impacts of these changes on aquaculture systems. The study also explores key adaptation strategies such as species diversification, technological innovations, and improved farm management practices. In addition, mitigation approaches aimed at reducing greenhouse gas emissions and enhancing sustainability are discussed in detail. Special emphasis is placed on the concept of climate-smart aquaculture, which integrates adaptation and mitigation strategies to improve resilience while ensuring sustainable production. The paper highlights the need for policy support, technological advancement, and collaborative efforts to address climate-related challenges. Overall, this review underscores the urgency of adopting holistic and sustainable approaches to safeguard aquaculture in the present and future climate scenarios.

Keywords: *Climate Change; Aquaculture; Adaptation; Mitigation; Climate-Smart Aquaculture; Sustainability; Food Security*

1. Introduction

Aquaculture has emerged as a cornerstone of global food systems, supplying a substantial proportion of animal protein and supporting millions of livelihoods worldwide (FAO, 2022; Golden *et al.*, 2021). The rapid expansion of aquaculture has been driven by increasing demand for seafood, population growth, and the stagnation of capture fisheries (Costello *et al.*, 2021; Naylor *et al.*, 2021). Despite its growth, aquaculture remains highly dependent on environmental conditions, making it particularly sensitive to climate change (IPCC, 2023; Barange *et al.*, 2022). Anthropogenic greenhouse gas emissions have resulted in global warming, altered precipitation patterns, and increased frequency of extreme weather events (Smale *et al.*, 2021; Bindoff *et al.*, 2022).

In the present scenario, climate variability is already influencing aquaculture productivity through changes in water temperature, salinity, dissolved oxygen, and nutrient dynamics (Boyd, 2021; Yadav *et al.*, 2024). Moreover, climate-induced stressors are exacerbating disease outbreaks and affecting species distribution, thereby impacting economic returns and food security (Revereret *et al.*, 2021; Pernet *et al.*, 2022). Understanding the complex interactions between climate change and aquaculture is essential for developing adaptive strategies and ensuring sustainable development (Ahmed & Thompson, 2022; Belton *et al.*, 2022).

2. Climate Change Drivers Affecting Aquaculture

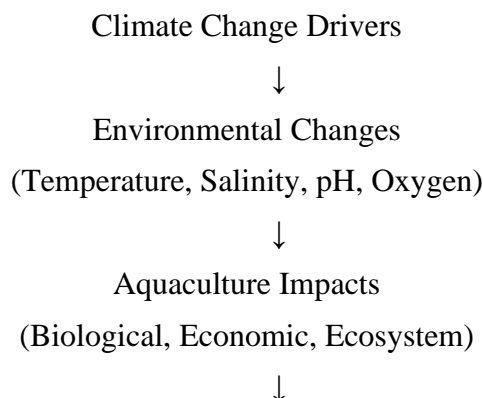
The accumulation of greenhouse gases such as CO₂, CH₄, and N₂O is the primary driver of global climate change (IPCC, 2023; Bindoff *et al.*, 2022). These gases trap heat, leading to rising temperatures that directly influence aquatic ecosystems (Barange *et al.*, 2022; Moss *et al.*, 2025). In aquaculture, warming trends alter metabolic processes, reduce dissolved oxygen availability, and increase physiological stress in aquatic organisms (Boyd, 2021; Kumar *et al.*, 2021). Temperature is a key environmental parameter regulating growth, reproduction, and survival of aquatic species (Boyd, 2021; Yadav *et al.*, 2024). Elevated temperatures increase metabolic rates and oxygen demand, reducing feed efficiency and growth performance (Froehlich *et al.*, 2022; Tacon *et al.*, 2022). Prolonged thermal stress can disrupt reproductive cycles and increase mortality rates (IPCC, 2023; Moss *et al.*, 2025). Ocean acidification results from increased CO₂ absorption, leading to decreased pH and carbonate ion availability (IPCC, 2023; Yacout *et al.*, 2025). This negatively affects calcifying organisms such as mollusks and crustaceans, impairing shell formation and growth (FAO, 2020; Barange *et al.*, 2022).

Sea-level rise threatens coastal aquaculture through flooding, salinity intrusion, and habitat degradation (IPCC, 2023; Barange *et al.*, 2022). Coastal ecosystems such as mangroves, which support aquaculture, are particularly vulnerable (Veenhof *et al.*, 2024). The increasing frequency of cyclones, floods, and droughts disrupt aquaculture production systems and damage infrastructure (IPCC, 2023; Smale *et al.*, 2021). These events lead to stock losses and economic instability (Hossain *et al.*, 2025; Troell *et al.*, 2023). Changes in precipitation patterns affect freshwater availability and water quality, impacting inland aquaculture systems (IPCC, 2023; De Silva & Soto, 2009). Water scarcity and flooding events reduce production efficiency (FAO, 2020; Yadav *et al.*, 2024).

Climate-induced salinity fluctuations stress aquatic organisms and affect survival and growth (Barange *et al.*, 2022; Kumar *et al.*, 2021). Climate change influences nutrient cycling and primary productivity, affecting food availability and ecosystem dynamics (Troell *et al.*, 2023; FAO, 2020).

Table 1. Major Climate Change Drivers and Their Effects on Aquaculture

Driver	Key Changes	Impact on Aquaculture
Global warming	Rising temperatures	Reduced oxygen, stress, mortality
Ocean acidification	Lower pH	Shell damage, reduced growth
Sea-level rise	Coastal flooding	Habitat loss, salinity intrusion
Extreme events	Cyclone, floods	Infrastructure damage, stock loss
Hydrological changes	Rainfall variability	Water scarcity/flooding
Salinity changes	Fluctuations	Osmotic stress
Productivity shifts	Nutrient imbalance	Food web disruption



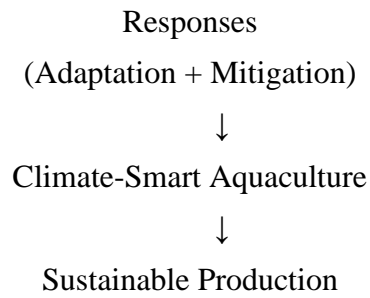


Figure 1. Conceptual Framework: Climate Change and Aquaculture

3. Impacts of Climate Change on Aquaculture

Climate change alters metabolic rates, growth, and reproductive cycles in aquatic species (Boyd, 2021; IPCC, 2023). Temperature stress reduces productivity and increases mortality (Froehlich *et al.*, 2022). Warmer temperatures and environmental stress promote pathogen proliferation, increasing disease outbreaks (Reverteret *et al.*, 2021; Pernet *et al.*, 2022). Emerging diseases pose significant threats to aquaculture sustainability (Burge *et al.*, 2021). Climate change leads to habitat degradation, biodiversity loss, and disruption of aquatic ecosystems (IPCC, 2023; Barange *et al.*, 2022). Climate-induced production losses affect income and livelihoods, particularly in developing countries (FAO, 2020; Belton *et al.*, 2022). Aquaculture is critical for global food security, and climate change threatens its ability to meet growing demand (FAO, 2022; Golden *et al.*, 2021).

Table 2. Impacts of Climate Change on Aquaculture

Impact Category	Effects
Disease	Increased outbreaks
Biological	Reduced Growth, Mortality
Ecosystem	Habitat loss
Economic	Income reduction
Food security	Supply instability

4. Vulnerability of Aquaculture Systems

Different aquaculture systems exhibit varying levels of vulnerability depending on environmental and socio-economic factors (De Silva & Soto, 2009; FAO, 2020). Small-scale farmers face higher risks due to limited adaptive capacity (Troell *et al.*, 2023; Belton *et al.*, 2022).

5. Adaptation Strategies

Adaptation strategies are essential to enhance resilience:

- Species diversification and selective breeding (Barangeet *al.*, 2022; Hossain *et al.*, 2025)
- Improved farm management practices (Boyd, 2021; Yadav *et al.*, 2024)
- Adoption of advanced technologies such as RAS and biofloc systems (D'Agaroet *al.*, 2022; Emerencianoet *al.*, 2022)
- Integrated aquaculture systems (Troell *et al.*, 2023)
- Climate forecasting and early warning systems (IPCC, 2023; Ahmed & Thompson, 2022)

6. Mitigation Strategies

Mitigation strategies in aquaculture aim to **reduce greenhouse gas (GHG) emissions**, minimize environmental impacts, and enhance the sustainability of production systems. While aquaculture is often considered less carbon-intensive than terrestrial livestock production, certain practices—such as feed production, energy use, and land conversion—contribute to climate change (Naylor *et al.*, 2021; Tacon *et al.*, 2022). Therefore, adopting mitigation strategies is essential for transitioning toward low-carbon aquaculture systems. Feed production represents one of the largest sources of emissions in aquaculture, particularly due to the use of fishmeal and fish oil derived from capture fisheries (Tacon *et al.*, 2022). Transitioning to **plant-based, microbial, or insect-based feed alternatives** can significantly reduce environmental footprints (Naylor *et al.*, 2021). Additionally, improving **feed conversion efficiency (FCR)** through precision feeding technologies reduces waste and nutrient discharge, thereby lowering emissions (Boyd, 2021).

Energy consumption in aquaculture operations—especially in intensive systems such as aeration, pumping, and recirculating aquaculture systems (RAS)—contributes to carbon emissions (D'Agaroet *al.*, 2022).

Mitigation measures include:

- Adoption of **solar and wind energy systems**
- Use of **energy-efficient aerators and pumps**
- Optimization of farm design to reduce energy demand.

The transition to renewable energy significantly lowers the carbon footprint of aquaculture systems (Naylor *et al.*, 2021). Shifting toward **low-carbon aquaculture practices** is a key mitigation strategy.

These include:

- Extensive and semi-intensive farming systems with lower external inputs
- Integrated multi-trophic aquaculture (IMTA), which utilizes waste from one species as input for another
- Biofloc systems that recycle nutrients within the culture system (Emerenciano *et al.*, 2022)

Such systems enhance resource efficiency while minimizing environmental impacts. Certain aquaculture practices, particularly **seaweed and shellfish farming**, contribute to carbon sequestration by absorbing CO₂ from the environment (Veenhof *et al.*, 2024).

Seaweed aquaculture, in particular:

- Removes dissolved carbon dioxide
- Improves water quality
- Supports ecosystem restoration

This makes it a promising nature-based solution for climate change mitigation. Efficient waste management reduces environmental pollution and GHG emissions. Strategies include:

- Recycling organic waste through biofloc systems
- Integrated farming (fish–paddy, fish–livestock systems)
- Use of sedimentation ponds and filtration systems

These approaches help minimize nutrient discharge and improve environmental sustainability (FAO, 2020; Boyd, 2021). Aquaculture expansion, particularly shrimp farming, has historically led to **deforestation of mangroves**, releasing large amounts of stored carbon (Barange *et al.*, 2022). Climate-smart intensification focuses on increasing productivity while minimizing environmental impact. This includes:

- Precision aquaculture using sensors and AI
- Optimized feeding and stocking densities
- Improved water quality monitoring systems

Such approaches reduce resource use and emissions while maintaining high productivity (Ahmed & Thompson, 2022). Effective mitigation requires strong policy support, including:

- Incentives for low-carbon technologies
- Regulations on sustainable aquaculture practices
- Carbon footprint assessment frameworks

International organizations emphasize integrating aquaculture into **national climate action plans** to achieve sustainability goals (FAO, 2020; IPCC, 2023). Mitigation in aquaculture can be broadly categorized into:

- **Technological solutions** (RAS, biofloc, renewable energy)
- **Ecological approaches** (IMTA, seaweed farming, mangrove restoration)
- **Management strategies** (efficient feeding, waste recycling)
- **Policy interventions** (regulation, incentives, climate planning)

Mitigation strategies are essential for reducing the environmental footprint of aquaculture while ensuring sustainable production. The integration of technological innovation, ecosystem-based approaches, and supportive policy frameworks can significantly contribute to climate change mitigation. In the present scenario, transitioning toward low-carbon and resource-efficient aquaculture systems is critical for achieving long-term sustainability and resilience.

7. Climate-Smart Aquaculture

Climate-smart aquaculture (CSAq) represents an integrated approach that combines adaptation and mitigation strategies to enhance resilience, productivity, and environmental sustainability in aquaculture systems (FAO, 2020; Ahmed & Thompson, 2022). It aligns with broader climate-smart agriculture principles by focusing on three core objectives: increasing productivity, enhancing adaptive capacity, and reducing greenhouse gas emissions.

A key component of CSAq is **resource-use efficiency**, which involves optimizing feed, water, and energy inputs to minimize waste and environmental impacts (Belton *et al.*, 2022; Boyd, 2021). Improved feed formulations, precision feeding techniques, and real-time monitoring systems help reduce feed conversion ratios and nutrient discharge into surrounding ecosystems. Efficient water management practices, including recirculating aquaculture systems (RAS) and biofloc technology, further enhance sustainability by conserving water and recycling nutrients (D'Agaroet *et al.*, 2022; Emerencianoet *et al.*, 2022).

CSAq also emphasizes **climate-resilient species and system diversification**. The selection of species with higher tolerance to temperature fluctuations, salinity changes, and disease conditions is crucial for maintaining productivity under changing climatic conditions (Barange *et al.*, 2022; Hossain *et al.*, 2025). Diversified farming systems, including polyculture and integrated multi-trophic aquaculture (IMTA), improve ecological balance and reduce risks associated with monoculture practices (Troell *et al.*, 2023). Another important aspect is the **integration of advanced technologies** such as artificial intelligence (AI), Internet of Things (IoT), and remote sensing for real-time monitoring of water quality, disease outbreaks, and environmental parameters (Naylor *et al.*, 2021; D'Agaro *et al.*, 2022). These technologies enable data-driven decision-making, improving farm management efficiency and reducing climate-related risks.

CSAq further promotes **ecosystem-based approaches**, including mangrove restoration, seaweed farming, and conservation of aquatic biodiversity. These approaches not only enhance resilience but also contribute to carbon sequestration and ecosystem services (Veenhof *et al.*, 2024). Socio-economic dimensions are equally important in CSAq. Capacity building, knowledge transfer, and stakeholder participation are essential to ensure the successful adoption of climate-smart practices, particularly among small-scale farmers (Ahmed & Thompson, 2022; Belton *et al.*, 2022). Overall, climate-smart aquaculture provides a holistic framework for addressing climate challenges while ensuring sustainable growth of the sector.

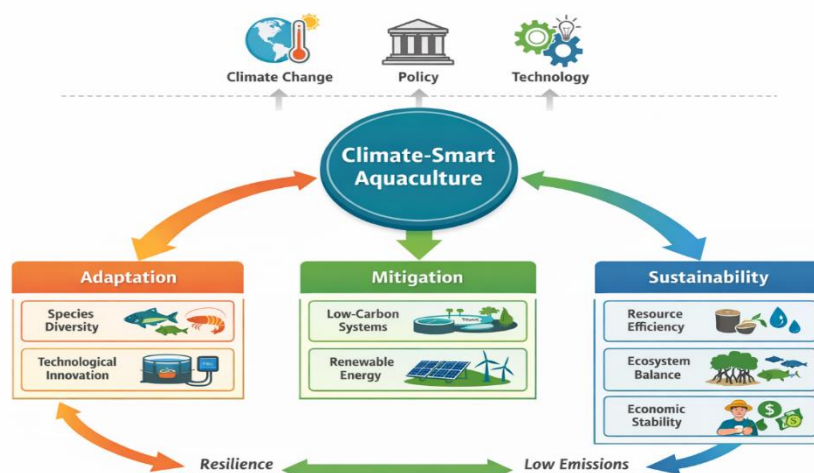


Figure 2. Climate-Smart Aquaculture Model

8. Policy and Governance

Effective policy and governance frameworks are critical for promoting climate-resilient and sustainable aquaculture systems. Governments, international organizations, and stakeholders play a central role in shaping policies that facilitate adaptation and mitigation efforts (FAO, 2020; Barange *et al.*, 2022). One of the primary policy priorities is the **integration of aquaculture into national climate strategies**, including Nationally Determined Contributions (NDCs) and climate action plans (IPCC, 2023). This ensures that aquaculture is recognized as both a vulnerable sector and a potential contributor to climate mitigation.

Regulatory frameworks are essential for ensuring sustainable aquaculture practices. These include guidelines on site selection, water use, waste management, and environmental impact assessments. Strict enforcement of regulations helps prevent ecosystem degradation and promotes responsible aquaculture development (Barange *et al.*, 2022). Financial mechanisms such as **subsidies, insurance schemes, and climate financing** are crucial to support farmers in adopting climate-resilient technologies and practices (Ahmed & Thompson, 2022). Access to credit and investment in infrastructure can significantly enhance adaptive capacity, particularly in developing countries.

Research and innovation policies play a vital role in advancing climate-smart aquaculture. Governments and institutions should invest in research on climate-resilient species, disease management, and sustainable production systems (FAO, 2020). Collaboration between academia, industry, and policymakers is essential to translate research findings into practical solutions. Capacity building and **extension services** are equally important. Training programs, awareness campaigns, and knowledge dissemination help farmers understand climate risks and adopt appropriate adaptation strategies (Belton *et al.*, 2022).

International cooperation is also critical, as climate change is a global issue. Collaborative efforts involving data sharing, technology transfer, and joint research initiatives can enhance global resilience in aquaculture (Troell *et al.*, 2023). In summary, strong governance frameworks supported by effective policies, financial investments, and institutional collaboration are essential to ensure the sustainable development of aquaculture under changing climatic conditions.

9.Future Perspectives

The future of aquaculture in the context of climate change will be shaped by technological innovation, sustainable practices, and global collaboration. Emerging technologies are expected to play a transformative role in enhancing resilience and productivity. **Precision aquaculture**, driven by AI, machine learning, and IoT, will enable real-time monitoring and predictive analytics for water quality, feeding, and disease management (D'Agaroet *et al.*, 2022; Naylor *et al.*, 2021). These technologies will improve efficiency, reduce resource use, and minimize environmental impacts.

Advancements in **biotechnology and genetic improvement** will facilitate the development of climate-resilient species with enhanced tolerance to temperature stress, salinity changes, and disease resistance (Hossain *et al.*, 2025). Selective breeding and genomic tools will play a crucial role in improving productivity under changing environmental conditions. The adoption of **sustainable intensification practices** will be essential to meet increasing global demand for aquatic food while minimizing environmental impacts (Troell *et al.*, 2023). Integrated systems such as IMTA and aquaponics will enhance resource efficiency and ecological balance.

Future aquaculture development will also focus on **low-carbon and circular economy approaches**, including waste recycling, energy efficiency, and carbon-neutral production systems (Naylor *et al.*, 2021). Seaweed farming and ecosystem restoration initiatives will contribute to carbon sequestration and environmental sustainability (Veenhof *et al.*, 2024). Climate risk management tools, including **early warning systems and climate forecasting**, will become increasingly important in reducing production risks and improving resilience (IPCC, 2023). However, challenges such as limited financial resources, technological gaps, and policy constraints must be addressed to ensure equitable development. Strengthening international collaboration, promoting knowledge exchange, and supporting small-scale farmers will be key to achieving sustainable aquaculture growth.

10.Conclusion

Climate change poses significant and multifaceted challenges to aquaculture, affecting biological processes, ecosystem dynamics, and socio-economic systems (IPCC, 2023; Barange *et al.*, 2022). Rising temperatures, ocean acidification, altered hydrological cycles, and extreme weather events are already impacting aquaculture productivity, health of aquatic

organisms, and overall system stability (Smale *et al.*, 2021; Yacout *et al.*, 2025). Despite these challenges, aquaculture also presents substantial opportunities for adaptation and mitigation. The adoption of climate-smart practices, technological innovations, and sustainable management strategies can enhance resilience and reduce environmental impacts (Ahmed & Thompson, 2022; Belton *et al.*, 2022). Advanced systems such as biofloc technology, recirculating aquaculture systems, and precision aquaculture offer promising solutions for improving efficiency and sustainability (D'Agaro *et al.*, 2022; Emerenciano *et al.*, 2022).

Policy support, research investment, and stakeholder collaboration are critical to addressing climate challenges and ensuring the long-term sustainability of the sector (FAO, 2020; Barange *et al.*, 2022). Inclusive approaches that consider the needs of small-scale farmers and vulnerable communities are particularly important. A holistic and interdisciplinary approach is essential to integrate environmental, economic, and social dimensions of sustainability. Strengthening governance frameworks, promoting innovation, and enhancing global cooperation will be key to building resilient aquaculture systems (Troell *et al.*, 2023; IPCC, 2023).

In conclusion, while climate change presents significant risks, it also provides an opportunity to transform aquaculture into a more sustainable, resilient, and climate-smart sector capable of supporting global food security in the future.

11. References

- 1) Ahmed, Nesar, & Thompson, Susanna. (2022). *Climate change adaptation in aquaculture*. *Marine Policy*, 129, 104536.
- 2) Barange, Manuel, Bahri, Terence, Beveridge, Malcolm C. M., Cochrane, Keven L., Funge-Smith, Simon, & Poulain, Florence. (2018). *Impacts of climate change on fisheries and aquaculture*. *Food and Agriculture Organization*.
- 3) Barange, Manuel, Bahri, Terence, Beveridge, Malcolm C. M., Cochrane, Keven L., Funge-Smith, Simon, & Poulain, Florence. (2022). *Climate impacts on fisheries and aquaculture*. *ICES Journal of Marine Science*, 79(5), 1234–1245.
- 4) Belton, Ben, Bush, Simon R., Little, David C., & Phuong, Nguyen Thi Kim. (2022). *Aquaculture resilience and adaptation*. *Global Environmental Change*, 73, 102452.
- 5) Boyd, Claude E. (2015). *Water quality: An introduction*. *Springer*.

- 6) Bindoff, Nathaniel L., Cheung, William W. L., Kairo, James G., Arístegui, Javier, Guinder, Verónica A., Hallberg, Robert, Hilmi, Nathalie J. M., Jiao, Nianzhi, Karim, Muhammad S., Levin, Lisa A., O'Donoghue, Sarah, PurcaCuicapusa, Susan R., Rinkevich, Baruch, Suga, Toshio, Tagliabue, Alessandro, & Williamson, Philip. (2022). *Ocean warming and marine heatwaves*. In IPCC Working Group II Report.
- 7) Boyd, Claude E. (2021). *Water quality challenges in aquaculture*. *Aquaculture Environment Interactions*, 13, 1–15.
- 8) Burge, Colleen A., Eakin, Christopher S., Friedman, Carolyn S., Froelich, Brett, Hershberger, Paul K., Hofmann, Gretchen E., Petes, Laura E., Prager, Kathleen C., Weil, Ernesto, & Willis, Bette L. (2021). *Climate change influences on marine infectious diseases*. *Annual Review of Marine Science*, 13, 397–418.
- 9) Can, Mehmet Fatih, & Mazlum, Yalçın. (2025). *Global warming and sustainable aquaculture*. *Aquaculture International*.
- 10) Costello, Christopher, Cao, Ling, Gelcich, Stefan, Cisneros-Montemayor, Andrés M., Free, Christopher M., Froehlich, Halley E., Golden, Christopher D., Ishimura, Gen, Maier, Johanna, Macadam-Somer, Ian, Mangin, Thibaut, Melnychuk, Michael C., Miyahara, Masaki, de Moor, Carryn L., Naylor, Rosamond L., Nøstbakken, Linda, Ojea, Elena, O'Reilly, Elizabeth, Parma, Ana M., Plantinga, Andrew J., Thilsted, Shakuntala H., & Lubchenco, Jane. (2021). *The future of food from the sea*. *Nature*, 588, 95–100.
- 11) D'Agaro, Enrico, Gallo, Antonio, & Sicuro, Benedetto. (2022). *Recirculating aquaculture systems under climate stress*. *Aquacultural Engineering*, 96, 102207.
- 12) De Silva, Sena S., & Soto, Doris. (2009). *Climate change and aquaculture: Potential impacts*. *Aquaculture Research*, 40(5), 551–560.
- 13) Emerenciano, Mauricio, Gaxiola, Gabriela, & Cuzon, Gerard. (2022). *Biofloc technology and sustainability*. *Aquaculture*, 548, 737676.
- 14) FAO. (2020). *Climate-smart aquaculture guidelines*. *Food and Agriculture Organization*.
- 15) FAO. (2022). *The state of world fisheries and aquaculture 2022*. *Food and Agriculture Organization*.
- 16) FAO. (2023). *Climate risk management in fisheries and aquaculture*. *Food and Agriculture Organization*.

- 17) Froehlich, Halley E., Jacobsen, Nis Sand, Essington, Timothy E., Clavelle, Tyler, & Halpern, Benjamin S. (2022). *Climate-driven shifts in aquaculture productivity*. Nature Communications
- 18) Gephart, Jessica A., Henriksson, Peter J. G., Parker, Rosamond W. R., Shepon, Alon, Gorospe, Karina D., Bergman, Kelsey, Eshel, Gidon, Golden, Christopher D., & Halpern, Benjamin S. (2021). *Environmental performance of aquaculture*. Nature Sustainability, 4, 21–30.
- 19) Golden, Christopher D., Allison, Edward H., Cheung, William W. L., Dey, Madan M., Halpern, Benjamin S., McCauley, Douglas J., Smith, Matthew, Vaitla, Bapu, Zeller, Dirk, & Myers, Samuel S. (2021). *Aquatic foods to nourish nations*. Nature, 598, 315–320.
- 20) Hossain, Md. S., Rahman, Md. M., Islam, Md. S., & Ahmed, Nesar. (2025). *Impact of climate change and adaptation strategy in aquaculture: A systematic review*. Aqua
- 21) IPCC. (2023). *Climate change 2023: Synthesis report*.
- 22) Kumar, Gopal, Engle, Carole R., Tucker, Craig S., & Kumar, Vinay. (2021). *Climate change and aquaculture sustainability*. Aquaculture Reports, 19, 100659.
- 23) Moss, Andrew, Smith, John, & Lee, David. (2025). *Aquaculture and climate change: A data-driven analysis*. Annals of Animal Science, 25(2), 547–568.
- 24) Naylor, Rosamond L., Hardy, Ronald W., Buschmann, Alejandro H., Bush, Simon R., Cao, Ling, Klingler, Dane H., Little, David C., Lubchenco, Jane, Shumway, Sandra E., & Troell, Max. (2021). *Blue food systems for sustainability*. Nature Food, 2, 437–445.
- 25) Pernet, Fabrice, Barret, Jacques, Le Gall, Patrice, Corporeau, Charlotte, & Lagarde, Franck. (2022). *Climate change and pathogen dynamics in aquaculture*. Annual Review of Marine Science, 14, 123–145.
- 26) Phillips, Michael J., Henriksson, Peter J. G., Tran, Nguyen, Chan, Chi-Kit, Mohan, Chandrika V., Rodriguez, U. P., Suri, Swati, Hall, Spencer, Koeshendrajana, Sonny, Pham, Suriya, & Subasinghe, Rohana. (2021). *Blue economy and aquaculture sustainability*. Aquaculture Economics & Management, 25(1), 1–14.
- 27) Reverter, Marc, Sarter, Sébastien, Caruso, Daniela, Avarre, Jean-Claude, Combe, Marianne, Pepey, Emmanuel, Pouyaud, Laurent, & de Verdal, Hélène. (2021). *Aquaculture at the crossroads of climate change and disease*. Fish and Shellfish Immunology, 108, 21–34.

- 28) Sharma, Saurabh, Singh, R. P., & Kumar, Ajay. (2024). *Microplastics in aquaculture systems*. Environmental Sciences Europe, 36, 181.
- 29) Smale, Dan A., Wernberg, Thomas, Oliver, Eric C. J., Thomsen, Mads S., Harvey, Ben P., Straub, Sandra C., Burrows, Michael T., Alexander, Lisa V., Benthuyesen, Jessica A., Donat, Markus G., Feng, Ming, Hobday, Alistair J., Holbrook, Neil J., Perkins-Kirkpatrick, Sarah E., Scannell, Hillary A., Sen Gupta, Alex, & Payne, Bonnie L. (2021). *Marine heatwaves threaten ecosystems*. Nature Climate Change, 11, 338–344.
- 30) Soto, Doris, Aguilar-Manjarrez, Juan, & Hishamunda, N. (2021). *Climate-smart aquaculture systems*. FAO Fisheries Reports.
- 31) Subasinghe, Rohana, Soto, Doris, & Jia, Jie. (2009). *Global aquaculture and its role*. FAO Technical Paper.
- 32) Tacon, Albert G. J., Metian, Marc, McNevin, Aaron A., & Bostock, John. (2022). *Aquaculture sustainability under climate stress*. Reviews in Aquaculture, 14(1), 158–176.
- 33) Tigchelaar, Michelle, Battisti, David S., Naylor, Rosamond L., & Ray, Deepak K. (2021). *Climate impacts on food systems*. Science Advances, 7(24).
- 34) Troell, Max, Naylor, Rosamond L., Metian, Marc, Beveridge, Malcolm C. M., Tyedmers, Peter H., Folke, Carl, Arrow, Kenneth J., Barrett, Scott, Crépin, Anne-Sophie, Ehrlich, Paul R., Gren, Ing-Marie, Kautsky, Nils, Levin, Simon A., Nyborg, Karine, Österblom, Henrik, Polasky, Stephen, Scheffer, Marten, Walker, Brian H., Xepapadeas, Anastasios, & de Zeeuw, Aart. (2017). *Does aquaculture add resilience to the global food system?* PNAS, 114(24), 6217–6222.
- 35) Troell, Max, Jonell, Magnus, & Crona, Beatrice. (2023). *Aquaculture and ecosystem services*. Global Food Security, 36, 100674.
- 36) Veenhof, Ruben J., Van den Burg, Saskia W. K., & Van Duijn, Andrew P. (2024). *Sustainable seaweed aquaculture and climate change*. Frontiers in Marine Science, 11, 1483330.
- 37) Yacout, Doaa M. M., El-Sayed, Ahmed F. M., & Abdel-Latif, Mahmoud. (2025). *Climate change impacts on aquaculture*. Frontiers in Sustainable Food Systems, 9, 1527798.
- 38) Yadav, N. K., Singh, S. K., & Kumar, R. (2024). *Climate change effects on aquaculture production*. Environmental Science and Pollution Research, 31, 31731–31751.

Cichlids Breeding

I. Vasudhevan

Assistant Professor, PG & Research Department of Zoology, Vivekananda College, Agasteeswaram, Kanyakumari, Tamil Nadu, India.

Corresponding Author Email: Email-goldvasu1953@gmail.com

Abstract

Mouth-brooding African cichlid mating occurs something like this the male digs out a cave (nest) for the female and lures her in by doing this silly fin shaking dance. She lays her eggs in the nest and he fertilizes them. Then she picks up the eggs in her mouth and carries them until they hatch. The eggs will hatch between 10-15 days after fertilization. The female then hold the babies in her mouth for up to two more weeks, occasionally letting them out to feed them. Most new mothers will start releasing the babies about 2 weeks after you notice they are carrying them, but will suck them back in when danger threatens. New mothers also have very small batches of eggs around 10 or so. A mature female will have up to 30 eggs. The male fish has no part in the care and raising of young, and may happily eat the babies if he is present when they are released. I suggest a 10-15 gallon tank for the mother and babies. Make sure the new tank is fully cycled and give it lots of tiny hiding places for the babies. You can also try providing small hiding places for the baby fish in your main tank, but in my experience the adult fish find ways to eat the babies. Mouth-brooders like to be in a harem of the same species: one male and three or four females. The male will boss the females around but won't hurt them. Two mature males will shred each other until one dies.

Keywords: Mouth brooder, Reproduction, Filtration, Temperature, PH and Pigmentation

Introduction

Cichlid breeding is easy with some cichlids and difficult with others, but this is made easier if you as a cichlid owner take good care of your cichlids. Keeping the aquarium clean and maintaining the conditions required by that particular cichlid will keep your cichlids healthy. This will induce them to mate more readily. Perhaps the most essential element in cichlid breeding is keeping your cichlids in top shape. By investing in a good filter to keep the water free of pollutants, by purchasing a heater to maintain the temperature, by carrying out regular water changes by maintaining a good water chemistry (water should be

free of ammonia and nitrites) by providing your cichlids with a well-balanced diet. Only healthy fish will want to mate.

Moreover, many female cichlids do not eat during the incubation, which may in some cases last up to 4 weeks. Only a fish that is in good health will be able to endure such abstinence from food. For cichlid breeding, it is helpful to get several females for each male, as this way they male's bullying does not get restricted to just one female. Aggressiveness of tankmates can cause stress among the weaker fish. For cichlid breeding to be accomplished readily, it is essential that your cichlids are not stressed. The speed at which cichlid breeding is accomplished depends also on the diet. A varied diet should be provided. At the time of cichlid breeding it would be advisable to provide food that is rich in proteins.

Cichlids breed in different ways. Open Brooders (e.g. angelfish, discus) spawn on open surfaces like rocks, or on the substrate. Shelter Brooders are of two types – Cavity Brooders (e.g. Apistogramma) who lay their eggs in caves and in depressions, and Mouth Brooders (e.g. Aulonocara) who carry the eggs and the fry in their mouths. Open Brooders tend to lay more eggs (sometimes as much as 10,000) than the Shelter Brooders (about 300). Cichlid breeding becomes easier if these territorial creatures are provided with sheltered areas where they can spawn. Rocks and plants can be used to provide them with hiding places. Mouth brooders will benefit the most from these hideouts because they will need more than usual protection when they are egg-laden. The female releases the fry depending on a number of factors such as her species, her age, and the state of her health.

Water quality may also determine when the fry get released. Remember that young inexperienced females are not too likely to carry the eggs for the full term the first time round. After that first time however, they learn to breed successfully. Sometimes fish do not mate because they are overfed – the hungrier they are, the more their aggressive tendencies are likely to show up, and the more likely they will be to mate. However some species like Aulonocara naturally take time to become sexually mature. Be patient with them. Water temperatures of about 76°C and 78°C and alkaline, hard water will aid in cichlid breeding. The maintenance of a steady temperature will aid in the hatching of the eggs.

Raising and harvesting of cichlids can be done in several ways. The best method is allowing the mother to spit out the fry naturally in a separate tank. Here she does not have to worry about other adults who may eat the fry. The survival of the fry is almost certain in this case unless the mother herself consumes the fry. In such a case it would be good to transfer the mother to another tank once she has released the fry. Most fry start eating flakes and brine shrimp once they have absorbed their egg sacs. The fry will grow rapidly with good water conditions, frequent feeding, and good diets. Cichlid breeding is almost an art form, and you and your cichlids will keep getting better at it with time. Keep trying.

Setting up the Cichlid Aquarium

1. Rinse and clean the tank thoroughly without soap, for soap can be fatal for the cichlids.
2. Place the tank where it will remain permanently. Don't attempt moving the tank around when it has more than two inches of water in it. Not only would the tank be heavy, but this would also be unsafe. Moreover, the stress placed on the silicone on the tank might cause it to leak.
3. If the pH level of the water is too low, it can be combated by placing in the tank coral sand or alternatively, limestone and other rocks can also be used.
4. When adding the sand or gravel ascertain that these have been washed properly and slowly add it into the tank. Don't just drop in a lot of gravel together, as your tank might get damaged.
5. Add water till the tank is three-quarters full.
6. Set up the filter in a back corner, and put the airline tubing in place between the filter and the air-pump. Once you turn on the air-pump, the filter should start working.
7. Set up the heater along the back wall of the tank, and set it to a temperature of about 24 to 26°C. It is advisable to use a heater as this will help avoid fluctuations of temperature in the water. Keep your cichlid aquarium away from sunny areas of your home, as overheating of the tank may kill the cichlids.
8. As far as lighting a cichlid aquarium goes, stick to full spectrum lights that are not too bright. Being able to replicate the correct spectrum of light would go some way in determining how healthy your fish turns out to be. Excessively bright lights would in fact disturb them. Similarly, the background should be made out of neutral colors so that the cichlids feel comfortable and at home. Now fill the aquarium with water, but not right till the brim.

9. Wait for one whole day at the very least so that the temperature gets stabilized, and so that the chlorine present in the water evaporates.
10. Only after 24 hours have elapsed should you go to get your cichlids. When choosing your cichlids get a male and a female.
11. Don't put a whole crowd of fish in a new cichlid aquarium. The natural nitrogen cycle takes a little time to get started in a new aquarium. Hence, it is preferred to start with only one or two fish.
12. Also, try not to mix different species and place them in the same tank. Some like the angelfish feed on other cichlids like the neon tetra. Also, different species would have different needs as regards the conditions within the tank.
13. Don't flood your cichlids with food on the first few days. Start small, and only when your cichlids start exhibiting a desire to eat should you increase the amount of food you give them.
14. When decorating the aquarium, remember that this will be the home of your fish, so try to reproduce the conditions that prevail in their natural habitat. For instance, Mauna cichlids live in rocks, and therefore would be happier in an aquarium filled with rocks than in one filled with plants.
15. You could also place plants in your cichlid aquarium. Find out which plants are found in the natural environment of your particular cichlids. Don't just randomly pick out aquatic plants and place them there.
16. Change no more than 25% of the water every week.
17. At the same time rinse the filter thoroughly.
18. Clean the glass of your cichlid aquarium about once a month without soap.
19. Normally if the conditions in the cichlid tank remain stable, the fish do thrive. However, at times, despite following all the rules to take care of the fish, they may die. Don't let this put you off. With practice and experience you will be able to have a cichlid aquarium that is filled with many of these colorful finned creatures.

Proper Cichlid Care

You need to be able to take care of them properly. Cichlid care comes under a number of heads all of which must be taken care of effectively. Water quality, tank conditions and diet among other things have to be seen to regularly.

Temperature

Cichlid care begins with the aquarium. You need to be able to maintain the aquarium in such a way as to replicate the conditions in the original habitat of the cichlids. The ideal range of temperature would be between 74 F and 82 F, the optimum temperature being 78 F. For a tank of fry you could raise the temperature to between 80 F and 82 F. The higher temperature increases metabolism, leading the fry to eat more and therefore grow more rapidly. Lowering the temperature helps in lowering the aggression levels of aggressive fish, by lowering their metabolism. Care should be taken to see that the cichlid tank is not kept in a sunny area or near an appliance that generates a lot of heat. It would be advisable to invest in a heater to maintain the temperature and bear up against temperature fluctuations.

pH Level

One of the most important aspects of cichlid care is maintaining the pH level. The water in the original habitat of the cichlids is high on the pH scale (alkaline). Do not make any sudden changes to the pH level of the aquarium water as this would adversely affect the health of your cichlids. A 7.5 to 8.5 range on the pH scale would be ideal for the cichlid aquarium water. Acidic water can be tackled by using substances like the common soda which is both inexpensive, and effective.

Filtration

Filtration is essential, as this is what keeps the environment of your cichlids clean. There are several types of filters that are available today. Sponge filters work very well especially for smaller tanks. These are inexpensive, and if cleaned regularly, can work effectively for years. Undergravel filters are not as effective, as they have to be well-covered by the substrate so that they can work effectively, and the cichlids are always trying to dig them out. Outside power filters are also effective. Get one, whose flow rate is suited to your tank size. All filters must be cleaned regularly for best results.

Feeding

A variety of foods must be fed to your cichlids. Cichlid flakes and pellets, frozen, and even live food. Feed them smaller quantities frequently, rather than a large amount at one go. If you are unable to feed them often enough, you could provide them with rocks which have a lot of algal growth on them. Inappropriate kinds of food might make cause bloating, sickness, and may even be fatal. Cichlid care involves being aware of the specific needs of your cichlids, so do some research about them and their diets first.

Water Changes

As I mentioned earlier, cichlid care begins with the aquarium. The aquarium being a closed environment, gets unclean quickly especially in case of overcrowding, excessive feeding, and ineffective filters. Hence you need to change the water regularly, and in small quantities. At least 10% of the aquarium water should be changed every week. Some species, such as the Malawi Cichlids which are prone to aggressiveness may become gentler with water changes of up to 30% every two weeks. Water changes if made more often during the breeding period, and during the time of raising the fry, is beneficial. If water changes are made regularly, then the level of substances that are potentially harmful will be reduced.

Substrates

Substrates are used so that the alkalinity in the water may be maintained at its most optimum level. Substrates like gravel, crushed coral, and limestone help buffer the pH and maintain the alkalinity. Get a gravel siphon to clean the gravel. For tanks with undergravel filters, vacuuming is a good option to remove the dust and debris from the substrate. Tanks with plants however do not necessarily require vacuuming, as the roots themselves play a part in keeping the substrate clean. For African Cichlids, a 2" to 3" substrate works well as they dig a lot. For some cichlids, as little as a quarter inch of gravel works better.

Aggression Control

Cichlids in general are territorial creatures, and hence they tend towards aggression in the lakes. The aquarium with its smaller area would naturally increase their aggression. It may not be possible to completely control cichlid aggression, but there are ways of reducing it. By employing methods such as reducing the temperatures in the aquarium, having a low male population, keeping species that are compatible with each other, isolating pairs that are breeding, a cichlid tank owner can control the aggression among his cichlids.

Lighting

Lighting is also important for the aquarium. You need the appropriate lights to view your aquarium in the best possible way. Fluorescent lighting is the best as it reproduces the natural spectrum of light of the original habitat of the cichlids. Also, keep the lights subtle, as bright lights only disturb the fish, and also make them appear less colorful. Preferably, light your tank only towards evening. Leaving your aquarium lit throughout the day might lead to algae problems.

Cichlid Food

As with humans, cichlids too need to be provided with a well-balanced diet. Only eating meat-based foods will not do. A varied diet that includes vegetable matter as well is necessary. Fry especially need to be well-fed, else malnourishment might cause a stunting of their growth. Cichlids are not picky or finicky fish. They are mostly easy to feed. Plus, they have huge appetites which need to be satisfied. There is a very wide variety of species of cichlids. There is also a wide variety of cichlid foods to suit the differing food habits of this wide variety of cichlids.

The great majority of cichlids are omnivorous in nature. That is, they eat both live and plant foods. In their original habitat, these fish have a diet consisting of insects, crustaceans, worms, and plants. When living in an aquarium, for a cichlid, food should consist of a mix of flakes, live food, and vegetable matter. Cichlasomines, Angelfish, and the Heros species are examples of omnivorous cichlids.

A number of cichlids are carnivorous, that is, they are predators that prey on other fish. For a carnivorous cichlid, food can even mean the smaller fish that share its tank. Hence it is not advisable to keep a carnivorous cichlid with other smaller fish for it might just have them for dinner. Angelfish (of course angelfish are omnivores and not carnivores) for example, are perfectly capable of eating neon tetras if kept in the same aquarium. Carnivorous cichlid food in an aquarium can include worms, live fish, insects, insect larvae, and crustaceans, but they can also be fed pellets, flakes, and tablets. Carnivorous cichlids include Haplochromines and Pike Cichlids.

A cichlid that prefers to eat plant matter is called an herbivorous cichlid. Food for an herbivorous cichlid that is living in an aquarium could include flake and pellet foods that are plant-based. Such cichlids also feed on aquarium plants and can be fed plants and vegetables as well. Some herbivores will also accept live food. Some herbivores even feed on the algae that grow on rocks.

At the time of spawning, it would be beneficial to give the cichlid protein-based foods. Fry can mostly be started out on baby brine shrimp before they graduate to more adult foods. When feeding live foods like worms, one must make sure that these are clean and not

diseased. The same goes for feeder fish. The chances of the cichlids' contracting a disease is lower when the fish are fed manufactured foods such as pellets and flakes. Ideally, aquarium cichlids must be fed a wide variety of foods.

Conclusion

Cichlid breeding is a fascinating hobby that requires careful planning, attention to detail and a deep understanding of these complex fish, by replicating their natural habitats, selecting compatible species and providing proper care aquarists can successfully breed enjoy the beauty of these incredible creatures.

Inducing Spawning: The most effective triggers for breeding include water changes (40–50%) using slightly cooler water, high-quality, protein-rich diets (like frozen brine shrimp), and providing appropriate, private breeding sites (caves, flat rocks, or fine sand).

Separation of Batches: To ensure survival, many breeders "strip" the female of her eggs (removing them from her mouth) after 2–3 days and place them in an egg tumbler, as this provides a higher survival rate than letting the female release them.

Aggression Management: Breeding pairs often become highly aggressive and territorial, requiring separation from other tank mates.

References

- 1) Dawes, John (1991). *Livebearing fishes-Aguide to their Aquarium care, Biology and classification* Blandford. ISBN-0-7137-2152-9
- 2) Sandford,Gina (1995). *An illustrated encyclopedia of Aquarium fish.* Apple Press ISBN-1-85076-597-9
- 3) Dawes, John (1998). *The concise encyclopedia of popular freshwater tropical United Kingdom* ISBN-0-75252-4976
- 4) Santhakumar.R., N.V.Sujathkumar., I.Vasudhevan and K.Veerabhadran 2007. *Manual on freshwater ornamental fish culture, Dep. of Fisheries Extension, Fisheries College and Research Institute, Thoothukudi, TANUVAS.PP.1-47.*

- 5) Vasudhevan.I. et-al. *Live Jewels Breeding and Management Techniques (LJBMT-2012)* Vivekananda College, Agasteeswaram, Kanyakumari on 12th October 2012. PP. 1-181. ISBN -13-978-93-80819-12-9
- 6) David Alderton 2012. *Livebearers: Understanding Guppies, Mollies, Swordtails and Others*. CompanionHouse Books. ISBN 978-1-62008-006-1.
- 7) *Fish Taxonomy - Exotic and Laboratory Animals*. Merck Veterinary Manual. Retrieved 2026-03-25.
- 8) *Livebearers: Raise Guppies, Mollies, Platies & Swordtails in your Aquarium*. www.liveaquaria.com. Retrieved 2022-09-08.
- 9) Blackburn, D.G. 2018. Jégou (ed.). *Viviparity and oviparity: evolution and reproductive strategies*. Michael K. Skinner (2nd ed.). ISBN 978-0-12-815145-7.
- 10) Cole, Kathleen S. 2010. *Reproduction and Sexuality in Marine Fishes: Patterns and Processes*. Berkeley: University of California Press. pp. 129–150. ISBN 978-0-520-26433-5.
- 11) Keenleyside, Miles H.A. 1991. *Cichlid Fishes: Behaviour, Ecology and Evolution*. London: Chapman & Hall. pp. 191–210. ISBN 978-0-412-32200-6.

Live Jewels – Live Bearers

Subhitsha. S^{1*}, Roshini. M¹ and Anisha Devi. B¹

¹UG Student, PG & Research Department of Zoology, Vivekananda College, Agasteeswaram, Kanyakumari, Tamil Nadu, India.

*Corresponding Author Email: subhisubhitsha@gmail.com

Abstract

Livebearers are fish that retain their eggs inside the body and give birth to live, free-swimming young. They are especially prized by aquarium owners. Among aquarium fish, livebearers are nearly all members of the family Poeciliidae and include: guppy, molly, platy, endler's and swordtails. The advantages of livebearing to the aquarist are that the newborn juvenile fish are larger than newly-hatched fry, have a lower chance of mortality and are easier to care for. Unusual livebearers include seahorses and pipefish, where the males care for the young, and certain cichlids that are mouth brooders, with the parent incubating the eggs in the buccal cavity.

Keywords: *Guppy, Reproduction, Molly, Redswordtail, Temperature and Feed*

Introduction

Freshwater ornamental fishes, often called “live jewels” are diverse colourful aquatic species breed for aquarium decoration, aesthetic pleasure and commercial trade. According for roughly 60% of the global aquarium industry, these fishes include popular, hardy, vibrant tropical, coldwater, egg -laying and live bearers species. The industry represents a rapidly growing, high value and space efficient niche in aquaculture. USA is considered to be largest market of ornamental fishes, followed by Europe and Japan. It is reported that export from South East Asian countries accounts for nearly 69 percent of the total world production of ornamental fishes. Singapore accounts for the major share of 35 percent by value, followed by HongKong, Malaysia, Thailand, Philippines, Taiwan and Indonesia. About 80 percent of the market for ornamental fishes is of freshwater origin, next is from marine and brackish waters. India is currently a marginal player in the global ornamental fish trade with a market share of only 0.02 percent.

Live – bearers

Among the live bearers the most popular species are Guppy, Molly, Platy and Sword tail. Live bearing ornamental fishes have great demand in domestic as well as international market. Fishes, which give birth to young ones directly are called live bearers.

Live bearers are found suitable for women to start their own units to produce such fishes due to the following positive characters of the fishes.

- 1) Small sized fish suitable for handling and breeding.
- 2) They are gregarious (living in shoals) in nature sociable and peaceful.
- 3) They do not fight except for the mating disputes of males
- 3) They are handy and thrive in hand and alterative waters
- 4) They are prolific breeders.

Poecilia Sphenops (Molly)

Poecilia sphenops above and *poecilia velifera* right are two of quite a few species of wild available. Although there are quite a few species of wild Molly (including one, *Poecilia formosa*, that consists entirely of females), only three can be regarded as aquarium fish. These are *Poecilia sphenops*, which is, basically, a short-finned variety, and the two Sailfin species, *P.latipinna* and *P.velifera*. I am dealing with all three under one main section because numerous crosses between them mean that many of today's aquarium Mollies are of mixed parentage.

The Green/Black or Sphenops Molly (*P. sphenops*) is the most robust of the three aquarium species. Its extensive geographical distribution has led to the evolution of numerous races and this, in turn, has led to about 45 different scientific synonyms for the species. Although a real expert might be able to distinguish and name many of these, the only two which are of any real significance to us as aquarists are the domesticated form and the Liberty Molly. The Liberty Molly is, fundamentally, very similar to the wild-type in coloration, but has been selected over the years for the beautiful (short) dorsal fin that some of the males possess. It disappeared from circulation some time ago, but I have been very pleased to see it back in recent years, albeit in small numbers.

The domesticated form of the Molly is the one from which all the fancy 'Sphenops-type' varieties have been developed by exploiting the natural tendency of the species towards melanism (black pigmentation) and by selection of other mutations (inherited changes), such

as the one that gives rise to the Lyretail and its numerous permutations. As a result, one can find Lyretails and from black at one extreme, through green, gold, mottled(marbled), orange-tailed, and others, to the pure albino form at the other. Of the two Sailfin species, *P. velifera* (Yucatan Molly) is the larger, growing to 18cm (7in) in length. However, most fish never get to this size, so distinguishing between the two species can be quite difficult without close examination.

For the record, *P. velifera* has a fuller sailfin with 18-19 fin rays (compared with 14 for *P. latipinna*). In addition, the front edge of the dorsal fin originates nearer the head in *P. velifera*. Sailfins are available in a bewildering array of colours, with new ones appearing virtually every year. As well as the above three species, a fourth type of Molly is sometimes available. It has characteristics of both *P. sphenops* and one or other of the Sailfins. Such fish are hybrids and tend to be larger than sphenops, with a dorsal fin which is somewhere between sphenops and the true Sailfins. They are fertile and, in some ways, exhibit the best of both worlds, in that they are substantially larger fish than *P. sphenops* and are tougher all-round than either of the Sailfins.

Wild Mollies, unlike the majority of other popular community fish, often come from coastal regions. They therefore inhabit waters that tend to contain a certain amount of salt. I have, for instance, seen Mollies in Florida estuaries in water with a specific gravity reading of 1.015, which is pretty salty. Indeed, it is quite possible to keep Mollies in perfect health under tropical marine conditions, as long as they are acclimatized properly. In addition, some Molly species, particularly the Sailfins, are never at their best at temperature lower than around 25°C (77°F). This temperature is near the top end of the range as far as many community tanks are concerned. Consequently, a temperature in the low 20s°C (around 72°F), which is fine for many other fish, is below that required by some Mollies. In time, this is likely to have a debilitating effect on the fish, which can, in turn, lead to all sorts of ailments. In view of the above, a little extra should be put into the keeping of Mollies, which, given the conditions they require, are as hardy as most other aquarium fish, despite their reputation to the contrary.

Natural Range:(i) *P. latipinna*: North and South Carolina, Virginia, Florida, Texas and Atlantic coast of Mexico. (ii) *P. sphenops*: Texas to Mexico and down to Colombia, but also introduced into several exotic locations. (iii) *P. velifera*: Yucatan in Mexico.

Size:(i) *P. latipinna*: Around 10cm (4in) for males; around 12cm (4.7in) for females. (ii) *P. sphenops*: Up to 6cm (2.4in) for males; around 8cm (3.2in) for females. (iii) *P. velifera*: Up to 15cm (6in) for males, up to 18cm (7in) for females.

Food: All Mollies will accept a wide range of commercial diets and live foods, but should also receive a regular vegetable supplement.

Tank Conditions: Mollies are shoaling fish which should be kept in groups. Males will display constantly and may become aggressive towards each other, especially if kept in twos. It is, therefore, better to keep either one male with several females, or a group of males with the females. This applies more to the Sailfin Mollies than to Sphenops-type varieties. Ample swimming space bordered by plants should be provided, along with good lighting. Close attention needs to be given to water chemistry, since mollies often fare poorly if kept under 'average' community aquarium conditions. The water should be well filtered, alkaline and slightly to moderately hard with about one teaspoonful of salt added to every 4.5-5liters (1 Imperial gallon).

Temperature: Sphenops-type varieties will be comfortable between 23-28°C (73-82°F), while for Sailfins the ranges should be between 25-28°C (77-82°F), or even a little higher.

Breeding: Sphenops Mollies can produce about 80 fry every 5-7 weeks. Sailfin broods are even larger, with more than 100 fry being produced at intervals of 6-10 weeks, depending on temperature and season.

***Poecilia Reticulata* (Guppy)**

The GUPPY has been known scientifically as *Poecilia reticulata* since 1963 when two ichthyologists, Rosen and Bailey, published a major revision of livebearing fishes. Prior to that, it had been known by a variety of names, the most well-known of them being *Lebistes reticulatus*. With respect to its common name, the Guppy, this is derived from *Girardinus guppii*, the name given to it by Gunther in 1866 in honour of the Reverend Robert John Lechmere Guppy, who is sometimes referred to as the discoverer of this fish in Trinidad. He is, however, more likely to have rediscovered it, since the species was first scientifically described in 1859 by Peters, who based his findings on specimens from Caracas.

After the Mosquito Fish (*Gambusia affinis*), the Guppy (*Poecilia reticulata*) is the most widely distributed of the livebearers. Being so widely distributed and occupying such diverse habitats as ditches, streams and river tributaries, some of which are brackish, it is hardly surprising to find that wild Guppies are found in a variety of size and colour combinations in nature. However different these wild varieties may be, they are nevertheless more similar to each other than to any of the varieties of fancy Guppies found in pet shops. All the wild stocks have short unspectacular fins. Yet they possess a kind of 'pure' beauty that makes them strong favourites among experienced livebearer enthusiasts. A particularly beautiful favourite is the type known as Endler's Livebearer.

Breeders and researchers have been exploiting the inherent genetic variability of the Guppy ever since its introduction to the European hobby in 1908, with the result that we now have endless strains that become progressively more elaborate with each passing year. One of the centres of such guppy development and production is Florida, with Sri Lanka also becoming a progressively stronger Guppy-developing country. Despite this, the far probably still produces more new Guppy varieties than any other region.

Natural Range: Widely distributed north of the Amazon: Dutch Antilles, Trinidad, Windward Islands, Barbados, Grenada, Antigua, Leeward Island, St. Thomas, Venezuela and Guyana. In addition to this 'natural' distribution, this species is now found as established populations in numerous exotic locations from Alberta in Canada to Australia and Singapore.

Size: Up to 3cm (1.2in) for males; around 5cm (2in) for females. These sizes refer to wild-caught fish; cultivated varieties are generally larger.

Food: Wide range of small live, deep-frozen, freeze-dried and dry foods accepted, but the diet should include a vegetable component.

Tank Conditions: Guppies are peaceful shoaling fish which should be kept in a group. Males will display constantly, both towards each other and towards females, but no damage to fins results. Since commercial varieties possess enlarged fins, they should not be kept in the same tank as fin-nipping species such as Tiger Barbs (*Barbus tetrazona*). Open swimming space bordered by vegetation along the sides and back should be provided. Water chemistry is not critical, but the quality must be good, despite the undoubted hardiness of these fish. A small amount of salt may be found beneficial, particularly in the case of wild-caught specimens and for those reared in brackish conditions (one teaspoonful per 4.5-5liters/1Imperial gallon).

Temperature: 21-25°C (70-77°F) for general maintenance and breeding, although a much wider range is tolerated.

Breeding: As many as 193 fry have been recorded in a single breed, but, generally speaking, 30-40 fry are produced every 4-6 weeks.

***Xiphophorus Helleri* (Red Swordtail)**

The red swordtail (*Xiphophorus helleri*) is one of several 'aquarium' species which hardly ever seen in its original wild form in aquaria. This is a shame, because wild Swordtails, with their slim-lined bodies, short fins, magnificent straight swords and brilliant (and variable) coloration are truly beautiful fish. The closest we come to the original article these days are the short-finned Green and Black-spotted Green varieties produced by some commercial breeders. In addition to these relatively basic varieties, today's Swordtails are found in innumerable colour and fin configurations.

Two major factors have contributed to this:

A) the species' inherent tendency towards variation (wild populations can range from green to red, with or without body speckling)

B) the ease with which *X. helleri* can hybridize with its closest relatives, in particular (from the hobby point of view) the southern Platy or Moonfish (*X. maculatus*) and the Variatus or Sunset Platy (*X. variatus*).

Given such excellent 'raw materials', it is little wonder that there are so many commercial varieties of Swordtail in existence today. Broadly speaking, these fall into three main groups.

(i) short-finned fish which are similar in overall body shape to the wild form, but differ from it in coloration.

(ii) high-finned varieties (Hi-fin) of the Simpson type, which have a sail-like dorsal (back) fin, but in which all the other fins are pretty similar to the wild form.

(iii) the so-called Hi-fin Lyretail varieties, in which all the fins are highly developed and in which the caudal (tail) fin possesses ray extensions along the top and bottom edges, thus creating the 'lyre' for which these varieties are famous.

When choosing Swordtails, it is important to take into consideration the other fish with which they will share their aquarium. If these include fin-nipping species, such as Tiger Barbs (*Barbus tetrazona*), it would be sensible to avoid the more delicately finned Swordtail varieties.

A further point to bear in mind is that Swordtail males are generally aggressive towards each other. Therefore, choose only one male per tank or, alternatively, at least four to half a dozen. When kept in higher numbers, males are kept so occupied displaying to one another, that the risks of a single individual becoming the sole target of aggression are minimized. In this species, it is only the males that bear the characteristic sword. In addition, as in all other poeciliids, the anal (belly) fin of the male is modified into the mating organ known as the gonopodium. Females have normal anal fins. The tip of the gonopodium is 'adorned' with tiny structures known as hooks and claws, and it is the precise arrangement of these that make mating possible only between Swordtails, or between them and their very closest relatives.

In the 'true' Lyretail varieties, the gonopodium-just like all the other fins -is much enlarged, making successful mating impossible, particularly since the hook, claw and blade arrangement may be highly reduced or be totally lacking. As a result, these Lyretail males represent an evolutionary dead-end, although they are quite fertile. There are, however, some Lyretails which do not exhibit extended development of the gonopodium and these are capable of mating. The production of fully finned Lyretails, through, is only possible either by crossing a Lyretail female with a short-finned male, or by artificial insemination with sperm packets (spermatozeugmata) obtained from a lyretail male. This latter technique is a highly specialized one and must only be carried out by a suitably experienced person.

There are many anecdotal reports of sex reversal in Swordtails, with females apparently becoming sexually viable males over a period of time. Despite these numerous reports, there is no incontrovertible scientific evidence that female-to-male reversal occurs in this species. Such fish are believed to be late-differentiating males

Natural Range: Atlantic drainage of Central America. This species has become established in numerous locations, including Florida, Nevada, Wyoming, Arizona, California, Canada (Alberta), Hawaii, Puerto Rico, various locations outside the natural range in Mexico, South Africa, Sri Lanka, Australia and other regions.

Size: Up to 14cm (5.5in) for males (excluding sword); females up to 16cm (6.3in), but usually smaller.

Food: All commercial diets and livefoods are accepted.

Tank Conditions: The tank should be provided with a spacious open swimming area bordered by plants, along the sides and back. A tight –fitting cover should be installed (Swordtails are excellent jumpers). Bright illumination and well-filtered water will help show off the brilliant colours to best effect. Swordtails are tolerant of a wide range of water chemistry conditions, but alkaline, slightly hard water is preferred.

Temperature:20-26°C (68-79°C)

Breeding: As many as 150 fry may be produced by large females every 4-6 weeks, although broods are generally smaller than this.

***Xiphophorus Maclatus* (Platies)**

Looking AT today's colourful and highly developed platies, it is often difficult to determine their precise identity. Some look almost as if they are true Southern Platies or Moonfish (*Xiphophorus maculatus*) or Sunset/ Variatus. Platies (*X.variatus*) in every sense other than coloration, but many look as if they possess characteristics of both species. Many even look as if they have some Swordtail (*X. helleri*) genes in them. One variety goes as far as having a respectable sword. The reason why such confusion is possible in the first place is that Swordtails (see previous entry) and Platies are very closely related to one another, both belonging to the genus *Xiphophorus*. Because of their 'biological proximity', they can interbreed with the greatest of ease and produce viable hybrids in the process.

Such a state of affairs lends itself perfectly to commercial development, particularly when the original species themselves are highly variable in any case. The inevitable result of all this is that Platies are now available in a mind-boggling array of fin and colour configurations. However, we must not forget that these coloured, often over-finned, delightful mutants are descended from real species, which can, fortunately, still be found in abundance in the wild.

There are numerous naturally occurring forms of the Southern platy, some populations of which-such as that from Rio Coatzacoalcos-are very beautiful indeed.

X.maculatus can be relatively easily distinguished from *X.variatus*, despite any additions of fancy finnage or colours which may have occurred. *X.maculatus* has a rather stumpy body, while *X.variatus* is more ‘Swordtail-shaped’ (minus the sword, of course). In addition, *X.variatus* usually carries a series of vertical bars on the sides of the body. Obviously, these characteristics apply only to individuals of pure maculatus and variatus parentage. Where they have been hybridized, intermediate characteristics occur and distinguishing criteria become blurred.

X. variatus, despite its name, is not as varied a species as *X.maculatus*. Several natural populations do, however, exist, the most attractive probably being that from the Rio Axtla, which is closest in appearance to more ‘basic’ of the cultivated varieties. Two other fish which were formerly believed to be subspecies of *X.variatus* (namely *X.variatus xiphidium* and *X. variatus evelynae*) are now considered to be valid species in their own right: *X.evelynae*, respectively. Naturally occurring hybrids reportedly exist between *X.variatus* and (*X.kosszanderi* from the Rio La Marina) and between *X.variatus* and *X. couchianus*.

Natural Range: (i) *X.maculatus*: From Veracruz in Mexico to Belize and Guatemala. (ii) *X.variatus*: Atlantic slope of Mexico.

Size: (i) *X.maculatus*: Wild-caught males can be around 3.5cm (1.4in) in length; females, can reach 6cm (2.4in). (ii) *X.variatus*: Wild-caught males, around 5.5cm (2.2in); females around 7cm (2.8in). Cultivated varieties of both species, plus their hybrids, are generally larger.

Food: Wide range of commercial diets and live foods accepted, which should include a regular vegetable component.

Tank Conditions: The aquarium should be well lit and contain open swimming areas bordered by thick vegetation. Platies are peaceful community fish which should be kept as a shoal. Water chemistry is not too critical, but good quality neutral to slightly alkaline, slightly hard water is preferred.

Temperature: 18-25°C(65-77°F) recommended for *X.maculatus* ;16-27°C (61-81°F) Reported for *X.variatus*. Both can be kept and bred at temperature that fall within the *X.maculatus* range.

Breeding: Average broods of 50-60 fry are produced every 4-6 weeks, but larger broods have been reported for both sp.

Conclusion

Livebearers (primarily family Poeciliidae) represent one of the most popular and accessible groups of ornamental fish, making them an ideal entry point for beginners, yet they are also subject to sophisticated breeding and genetic studies. They are characterized by internal fertilization, where males use a modified anal fin called a **gonopodium** to inseminate females, resulting in the birth of free-swimming, fully developed young rather than eggs.

Breeding & Prolificacy: Livebearers are highly prolific, often producing new broods every 4–6 weeks. Common species like Guppies, Mollies, Platies, and Swordtails can produce 20–100+ young at a time, making them excellent for, or potentially overwhelming to, an aquarium hobbyist.

Aquarium Suitability: They are Hardy, adaptable, and generally thrive in community tanks. Because they give birth to advanced, large young, they are much easier to raise than egg-laying species.

Reproductive Biology: Most aquarium livebearers are **ovoviviparous** (eggs hatch inside the mother, with young relying on their own yolk sac), though some exhibit true **viviparity** (receiving nutrition from the mother via a placenta-like structure).

Ecological Impact: While prized in aquaria, some livebearers, such as the two spot livebearer and mosquito fish, are considered "high risk" invasive species due to their high fecundity, environmental tolerance, and negative impacts on native fish through competition and predation.

Selective Breeding: Due to their fast breeding cycles, they are widely used in selective breeding to create numerous color, tail, and pattern variations.

Scientific Value: Livebearers are utilized as models to study the evolution of life history, reproductive strategies, and the evolution of the placenta in fish.

In summary, livebearers are a resilient, colorful, and fascinating group of fish that are excellent for beginners, but they require management to prevent overpopulation and the accidental introduction of potentially invasive species into local ecosystems

References

- 1) Dawes, John (1991). *Livebearing fishes-Aguide to their Aquarium care, Biology and classification* Blandford. ISBN-0-7137-2152-9
- 2) Sandford,Gina (1995). *An illustrated encyclopedia of Aquarium fish.* Apple Press ISBN-1-85076-597-9
- 3) Dawes, John (1998). *The concise encyclopedia of popular freshwater tropical United Kingdom* ISBN-0-75252-4976
- 4) Santhakumar.R., N.V.Sujathkumar., I.Vasudhevan and K.Veerabhadran 2007. *Manual on freshwater ornamental fish culture, Dep. of Fisheries Extension, Fisheries College and Research Institute, Thoothukudi,TANUVAS.PP.1-47.*
- 5) David Alderton 2012. *Livebearers: Understanding Guppies, Mollies, Swordtails and Others.* CompanionHouse Books. ISBN 978-1-62008-006-1.
- 6) *Fish Taxonomy - Exotic and Laboratory Animals.* Merck Veterinary Manual. Retrieved 2026-03-25.
- 7) *Livebearers: Raise Guppies, Mollies, Platies & Swordtails in your Aquarium.* www.liveaquaria.com. Retrieved 2022-09-08.
- 8) Blackburn, D.G. 2018. Jégou (ed.). *Viviparity and oviparity: evolution and reproductive strategies.* Michael K. Skinner (2nd ed.). ISBN 978-0-12-815145-7.
- 9) Cole, Kathleen S. 2010. *Reproduction and Sexuality in Marine Fishes: Patterns and Processes.* Berkeley: University of California Press. pp. 129–150. ISBN 978-0-520-26433-5.
- 10) Keenleyside, Miles H.A. 1991. *Cichlid Fishes: Behaviour, Ecology and Evolution.* London: Chapman & Hall. pp. 191–210. ISBN 978-0-412-32200-6.

Management of Ornamental Fish Diseases

Jesi Nayaki. M^{1*}, Sahaya Nithisha. C¹ and Sheebha. S¹

¹UG Student, PG & Research Department of Zoology, Vivekananda College, Agasteeswaram, Kanyakumari, Tamil Nadu, India.

*Corresponding Author Email: jesinayaki2005@gmail.com

Abstract

Ornamental fisheries represent a dynamic, rapidly expanding sector of the global aquaculture industry, contributing significantly to livelihoods, international trade, recreational aquaculture, and biodiversity conservation. India, with its rich aquatic resources and growing expertise in ornamental fish culture, has emerged as an important player in this sector. However, the sustainability and profitability of ornamental fish farming are severely constrained by disease, which remains a critical challenge for breeders, traders, aquarium operators, and hobbyists. Diseases in ornamental fishes are caused by a wide range of pathogenic organisms, including bacteria, fungi, viruses, parasites, and other opportunistic microbes. Among these, bacterial, fungal, and viral diseases are of major concern due to their high prevalence, rapid transmission in confined systems, and potential to cause heavy mortalities. Ornamental fish are commonly reared in closed or semi-closed environments, such as aquaria, tanks, hatcheries, and recirculating systems, where stressors and suboptimal conditions can quickly lead to disease outbreaks

Keywords: *Bacteria, Fungi, PH, Protozoan and Environment*

Introduction

Major disease outbreaks in aquariums and ponds, are usually due to serious changes in the aquatic environment, the presence of one or more opportunistic pathogens and a susceptible host animal – fish. This explains why several disease problems often occur simultaneously in the same tank and why a tank that has been disease free for months can suddenly experience an epizootic. It also explains why disease outbreaks often coincide with the addition of new fish to an aquarium or pond even if the new animals were quarantined. The change in the biological load and species balance in the aquatic environment stresses the old inhabitants and makes them vulnerable to ubiquitous pathogens. The control of fish diseases in aquarium systems depend on the prevention of fish disease in fish stocks, good husbandry and rapid diagnosis and treatment of any problems as they arise.

A. Prevention

Since removal of the pathogen is an unrealistic option, it is important to know how to minimize the risk of infectious disease. In this sense, the application of hygienic and preventive measures of the environment, such as fish health management practices, sanitation and disease control procedures are critical factors to prevent fish disease. For this, fish should be obtained from apparently disease free stocks. New introduction should be quarantined for a period of at least one month during which careful observation and sampling should be carried out. Maintenance of good water quality is of paramount importance. Disinfection of aquaria and equipment may be routinely carried out. The major preventive measures are;

1. Optimization of environmental conditions

Many fish disease are found to be associated with serious changes in the environmental conditions. Therefore, ensuring good environmental and maintenance condition is one of the primary prophylactic measures in all aspects of fish culture. Make sure that all the water condition. Stocking density should be adequate. Make necessary steps for correcting water quality problems. Many commercial preparation are available for adjusting the PH of aquaria. These consist of carbonate, bicarbonate or phosphate buffers. Frequent routine water changes will prevent the drop in PH and can be used to adjust improper PH also.

Use mechanical aerators to correct dissolved oxygen problems. Ammonia levels can be reduced with frequent water changes. Adding zeolite is a safe and effective way of reducing ammonia quickly. Reducing the PH will reduce the percentage of ammonia that is present as NH_3 . This should be done with much care because a rapid drop in PH can cause other problems. In a new tank, adding a commercial preparation of nitrifying bacteria can speed up the process of establishing an effective filter. In an established tank, ammonia poisoning arises when more fishes are in the tank than the biological filtration can handle. In this case, a few of the fishes must be made by enhancing the rate of flow of water through the filter.

Nitrite toxicity can be eliminated by adding chloride (as sodium chloride). Nitrite is much less toxin when chloride is present, because chloride inhibits nitrite uptake across the gills. The management strategy is to establish chloride: nitrite ratio of at least 6:1. An addition management practices may be to reduce the amount of feed given to the fish. Lime

or other calcium salts are excellent sources of supplemental calcium for pond fish. Salt mixtures are commercially available for increasing hardness in aquaria. If aquarium hardness must be reduced, this can be done by adding distilled water.

2. Preventing the entry of pathogens:

Pathogens can be transmitted through fish and other aquatic animals, and also through water, feed, plants and soil particles. Actually, the introduction of fish suffering from diseases constitutes a special threat of infiltration of pathogens. It should be noted that many parasites undergo only a stage of development in the fish and an actual transmission and entry of the parasites occur only when the intermediate hosts are present at the point of entry and necessary conditions exist for completion of the life cycle. In principle, all aquarium fishes which are to be added to the established stock of other fishes should be subjected to quarantine. If, in the course of this process, microscopic investigations reveal the presence of ecto-or endo parasites, suitable bath treatments or other therapeutic measures must be undertaken.

Aquatic snails are the intermediate hosts of some fish parasites. Their preventive control can be performed by biological (introduction of black carp), mechanical (placing nets in the inflow), physical (drying and freezing of the bottom) and chemical (application of molluscicides) ways. When providing food to aquarium fishes, care must be taken that small animals carrying larvae of parasites of fish are not fed to fish. If live animals, constituting food for fish are taken from bodies of water with some fish living in them and are fed to aquarium fishes, there is always a certain danger of the transmission of parasites.

As far as possible, water should not be taken from sources in which fish have been living and particularly not from those with dense fish populations, since such waters can contain freely floating parasites or their larvae and spores of parasites, fungus and pathogenic bacteria and so on.

B. Treatment

A variety of treatments are available, depending very much on the aquarium system in use and the disease being treated. Primarily it should be determined whether the disease is internal or external. Subsequently, its aetiology, whether viruses, bacteria, parasites, or fungi must be confirmed. When internal infections are indicated, one of the few absorbable

drugs should be used, medications should be mixed with food or the drug should be injection directly into the animal. Determining fish drug dose must be a careful and precise process. Fish drugs should never be used prophylactically. They should only be applied when there are specific indications of disease.

1. An Inaccurate Diagnosis (or guess) is made as to the nature of the problem, which in turn results in the wrong treatment being used

This is often the situation where a guess is made as to the nature of the disease (usually based solely on clinical signs rather than an examination) and a succession of different treatments is applied in the hope that one of them will work.

2. An Accurate Diagnosis is Made but the Selected Treatment is not Successful

Treatments may be unsuccessful for a variety of reasons. In some cases pathogens are resistant to a particular treatment. It is not unusual for a treatment that works in one pond to be less successful against the same parasite in another pond. In such a situation another appropriate treatment should be used.

Another common problem that affects some disease treatments are variations in water chemistry. In these situations, water parameters such as hardness, PH or organic load can interfere with the action of the treatment, rendering it ineffective. There are also situations, particularly with severe parasite infestations, where, because of the host's reactions to their presence, the parasites are protected by excess mucus.

3. A straight Forward Treatment Overdose

This can arise from a simple miscalculation of dosages, or by the toxicity of the treatment being affected by water conditions such as PH and hardness. They are usually toxic to all life forms at relatively low doses. The effective dose is one that is high enough to kill the smaller organisms such as parasites, but not high enough to kill larger animals such as fish. A further consideration is variations in water chemistry. Most chemicals, with the exception of salt, are affected to some degree by variances in water hardness, PH and temperature. Many chemicals also react with dissolved and particulate organic matter such as fish waste, algae and detritus - thus affecting or reducing their efficacy.

Management of Viral Diseases

There are no specific treatments that will eliminate the virus or cause the skin lesions to regress more rapidly. Fish with skin lesions characteristics of pox should be isolated immediately to limit spread of the disease. Make sure that the fish is eating and provide an environment that is as low in stress as possible by maintaining good water quality.

Viral diseases cannot be controlled with medication because they use the host's own cells for reproduction and survival. Provide "good nursing care" for fish a high quality diet, maintaining clean facilities, keeping sick or potentially infected stock separate from all other animals, and disinfection of equipments.

Management of Bacterial Diseases

The susceptibility of fish increases in adverse environmental conditions – extreme temperatures, heavy load of organic matter in water, as well as maintenance of fish in overcrowded conditions. Therefore, maintenance of optimal growth conditions in the pond is indispensable preventive measure against occurrences of both acute and chronic bacteria infections.

Antibiotic baths and dips are probably best used for surface infections such as fin rot, bacteria gill disease and columnaris (cotton wool disease), where short duration baths with high dosages may be useful.

When using oxytetracycline in long-term baths, be aware that it is light sensitive and will start to turn brown as it decomposes. If this happens a 50% water change should be carried out immediately. Degraded tetracycline can be harmful to humans so avoid contact by wearing gloves.

Oral

Antibiotic	Dosage
Amoxicillin	40-80 mg/kg body-weight daily
Choramphenicol	50-100 mg/kg body-weight daily
Enrofloxacin	10 mg/kg body-weight daily
Oxolinic acid	10 mg/kg body-weight daily
Oxytetracycline	75 mg/kg body-weight daily

Management of fungal diseases

Since the superficial injuries are inviting foot holds for fungus, the fish culturist should do all he can to avoid or minimize them; great care must be exercised to avoid damage associated with handling, particularly during stocking, transporting and transplanting, and harvesting.

Several preventive procedures have been recommended. After stocking a pond with new fish, its surface should be sprayed with 0.15 ppm of malachite green, and the treatment repeated three times at intervals of three days.

When infections do break out among fish, therapeutic measures must be used.

Methylene blue: 0.02 ppm term bath; 3 ppm

Acridine: 0.02 ppm up to 3 days

Management of protozoan diseases

Effective treatment of protozoan ectoparasites depends on an understanding of the two major types of life styles: nonencysting protozoans (e.g., *Trichodina*, *Ichthyobodo*) complete their life cycle on the host and are easily treated, usually with a single, short-term application. Encysting protozoans (e.g., *Ichthyophthirius*, *Oodinium*) produce a reproductive cyst off the host. Both the feeding stage and the reproductive cyst are resistant to treatment, so therapy must be directed at the free-swimming, infective stage. This requires that chemicals be present for a long time or that several treatments be applied to ensure that all infective stages are killed.

A mixture of formalin and malachite green can be used effectively for controlling protozoan infection. This mixture has synergistic effect. It is formulated by dissolving 3.3g malachite green in 1L of formalin. It is diluted in 1:40,000 (2.5 ml in 100L designated as 25 ppm) or 1:1,67,000 (1.5 ml in 100L designated as 15 ppm). The higher concentration is used for exposures of up to 1 hour, the lower for prolonged treatment. Change up to 50% of the water on alternate days. Remove all plants before treatment.

The recommended treatment for Hexamita / Spironucleus is metronidazole (Flagyl). Metronidazole can be administered in a bath at a concentration of 5 mg/L every other day for three treatments.

Control of Dropsy

Since the exact cause of the disease is not known, precise treatment is difficult to prescribe. The removal of affected individuals to a separate tank is essential. Start them on a diet of high protein food that is enriched with vitamin.

It is best to up the water temp to about 82-86°F and use Epsom salts (1/8 teaspoon of Epsom salts per 5 gallons) instead of aquarium salts because they will help the fish to lose some of the fluid that has built up. It is also wise to increase aeration since the temperatures will be higher. Water quality is very important at this time. This will have to be tried for about 2 weeks.

As soon as the abdominal swelling is noted, isolate the fish and treat them with a broad-spectrum antibiotic. Early treatment is essential.

Oxytetracycline: 20-100 mg / L; 5 days bath.

Tetracycline hydrochloride: 40-100 mg / L; 5 days bath.

Conclusion

There are many diseases of fish, which can be troublesome to commercial producers as well as the recreational pond owner. There are two broad categories of diseases caused by parasites, bacteria, viruses, or fungi. Non-infectious diseases are broadly categorized as environmental, nutritional, or genetic. Disease is a substantial source of monetary loss to aquaculturists. Production costs are increased by fish disease outbreaks because of the investment, which is lost in dead fish, cost of treatment, and decreased growth during convalescence. A holistic approach should be adopted to the diagnosis, treatment and prevention of disease. This should consider the interaction of the environment, pathogens and host-related factors.

An accurate diagnosis is fundamental to the selection of an appropriate treatment. The economics, possible side effects and environmental impacts of treatments should be considered before application. Preventive management is the most important step in disease control since a systematic and thorough approach to health management can reduce the incidence of disease and associated production losses. Fish, which do not respond to a correctly administered treatment, should be re-evaluated by a fish health professional.

The following precautions can be taken to reduce the possibility of disease and keep it from spreading if it occurs:

1. By only good-quality, compatible fish
2. Quarantine new fish before adding them to aquarium
3. Avoid stressing the fish with rough handling or sudden changes in conditions
4. Don't over feed the fish
5. Remove sick fish to a hospital tank for treatment
6. Disinfect nets used to move sick fish
7. Don't transfer water from the quarantine tank to the main aquarium

References

- 1) Dawes, John 1991. *Live bearing fishes-A guide to their Aquarium care, Biology and classification* Blandford. ISBN-0-7137-2152-9
- 2) Santhakumar.R., N.V.Sujathkumar., I.Vasudhevan and K.Veerabhadran 2007. *Manual on freshwater ornamental fish culture (Tamil)*, Dep. of Fisheries Extension, Fisheries College and Research Institute, Thoothukudi, TANUVAS.PP.1-47.
- 3) Mohan kumar. G 2008.*Hand book on Ornamental fish diseases. The Marine Products Export Development Authority. Cochin, Kerala.*
- 4) Vasudhevan.I. et-al. *Live Jewels Breeding and Management Techniques (LJBMT-2012)* Vivekananda College, Agasteeswaram, Kanyakumari on 12th October 2012. PP. 1-181. ISBN -13-978-93-80819-12-9
- 5) David Alderton 2012. *Livebearers: Understanding Guppies, Mollies, Swordtails and Others. CompanionHouse Books. ISBN 978-1-62008-006-1.*
- 6) *Fish Taxonomy - Exotic and Laboratory Animals. Merck Veterinary Manual. Retrieved 2026-03-25.*
- 7) *Livebearers: Raise Guppies, Mollies, Platies & Swordtails in your Aquarium. www.liveaquaria.com. Retrieved 2022-09-08.*
- 8) Blackburn, D.G. 2018. Jégou (ed.). *Viviparity and oviparity: evolution and reproductive strategies. Michael K. Skinner (2nd ed.). ISBN 978-0-12-815145-7.*
- 9) Keenleyside, Miles H.A. 1991. *Cichlid Fishes: Behaviour, Ecology and Evolution. London: Chapman & Hall. pp. 191–210. ISBN 978-0-412-32200-6.*

Marine Ornamental Fishes in India: Status, Problems and Management Strategies

A. Pushparaj^{1*} and P. Ambika²

¹Assistant Professor, HOD of Zoology, TDMNS College, T. Kallikulam.

²Assistant Professor, St. Joseph College of Education, Nanguneri

*Corresponding Author Email: tdmnspushparaj@gmail.com

Abstract

The global marine ornamental fish trade represents a high-value, biodiversity-dependent sector that links coral reef ecosystems with international aquarium markets. India, endowed with extensive coastlines and ecologically rich reef systems such as the Gulf of Mannar, Lakshadweep Archipelago, and Andaman & Nicobar Islands, possesses significant potential for marine ornamental fisheries. Despite this potential, the Indian marine ornamental sector remains underdeveloped and faces multiple ecological, socio-economic, and regulatory challenges. Overharvesting, habitat degradation, climate change, inadequate monitoring, and limited aquaculture development threaten both reef biodiversity and fisher livelihoods. This review synthesizes available literature on the diversity, trade status, ecological significance, and policy framework governing marine ornamental fishes in India. It critically examines existing constraints and proposes science-based management strategies, including community-based co-management, hatchery-based production, regulatory strengthening, certification mechanisms, and ecosystem-based conservation approaches. The paper highlights the urgent need for integrated management linking biodiversity conservation with sustainable livelihood generation. With proper institutional support, research investment, and stakeholder participation, India can develop a responsible and sustainable marine ornamental fish sector aligned with national biodiversity and blue economy goals.

Keywords: *Marine ornamental fishes, India, Coral reefs, Sustainable fisheries, Aquarium trade, Reef conservation, Aquaculture, Fisheries management*

1. Introduction

Marine ornamental fishes form an integral component of the global aquarium industry, supplying colorful and ecologically unique species to hobbyists, public aquaria, and

commercial exhibits. Unlike food fisheries, the ornamental trade is driven by aesthetic value, rarity, and species-specific demand. Globally, the trade involves more than 2,000 marine fish species and generates hundreds of millions of dollars annually. A substantial proportion of these fishes are harvested from coral reef ecosystems in tropical regions.

India possesses approximately 7,500 km of coastline and diverse marine habitats, including coral reefs, seagrass meadows, mangroves, lagoons, and rocky intertidal zones. The country's reef ecosystems are concentrated in the Gulf of Mannar, Lakshadweep Islands, Andaman & Nicobar Islands, and parts of Gujarat. These regions support a wide array of reef-associated ornamental fishes. Despite this biodiversity richness, India's contribution to the global marine ornamental fish trade remains relatively modest compared to Southeast Asian nations such as Indonesia and the Philippines.

The marine ornamental sector offers a dual opportunity: biodiversity-based livelihood generation and export earnings. However, unregulated exploitation can cause long-term ecological damage. Therefore, understanding the status, challenges, and sustainable management pathways is essential for developing a resilient marine ornamental fish industry in India.

2. Historical Background

The ornamental fish trade in India began gaining momentum post-1990s alongside global commercial interest. Initial focus was largely on freshwater species; marine ornamentals became prominent in the 2000s as international demand surged. Local export hubs emerged in coastal states, particularly Tamil Nadu, Kerala, and Andaman & Nicobar Islands. Despite this growth, systematic assessments of species diversity and harvest levels in India remain limited.

3. Diversity of Marine Ornamental Fishes in India

India's reef ecosystems support more than 500 species of marine ornamental fishes. These species belong to several taxonomic families characterized by bright coloration, manageable size, and adaptability to aquarium conditions.

3.1 Major Families and Species

Important ornamental fish families include:

Family	Common Group	Example Species	Habitat	Trade Demand
Pomacentridae	Clown fishes & Damsel fishes	<i>Amphiprion ocellaris</i>	Coral reefs	Very High
Pomacentridae	Clown fishes & Damsel fishes	<i>Amphiprion ocellaris</i>	Coral reefs	Very High
Chaetodontidae	Butterfly fishes	<i>Chaetodon spp.</i>	Coral reefs	High
Pomacanthidae	Angel fishes	<i>Pomacanthus spp.</i>	Reef slopes	High
Gobiidae	Gobies	<i>Gobiodon spp.</i>	Coral branches	Moderate
Apogonidae	Cardinal fishes	<i>Pterapogon kauderni</i>	Reef lagoons	High

Clown fishes are among the most popular ornamental species due to their symbiotic association with sea anemones and increased demand following media exposure. Damsels and gobies are widely collected due to their abundance and resilience in captivity.

Region	Biodiversity Status	Key Species	Major Issues
Gulf of Mannar	High	Clownfish, Damsel fish	Overharvesting
Lakshadweep	Very High	Butterfly fish	Coral bleaching
Andaman & Nicobar	Very High	Angelfish	Regulatory gaps
Kerala Coast	Moderate	Gobies	Habitat degradation

3.2 Habitat Distribution

Marine ornamental fishes in India inhabit:

- Fringing coral reefs
- Atoll reef systems
- Seagrass beds
- Rocky reef outcrops
- Lagoon systems

The Andaman & Nicobar Islands exhibit the highest diversity, followed by Lakshadweep and the Gulf of Mannar.

4. Ecological Significance

Marine ornamental fishes play crucial ecological roles in coral reef ecosystems.

4.1 Trophic Interactions

Many ornamental fishes are herbivores or planktivores. Surgeonfishes and some damselfishes control algal growth, preventing coral overgrowth. Butterflyfishes often feed on coral polyps and serve as indicators of reef health.

4.2 Reef Resilience

Fish diversity contributes to reef resilience by maintaining functional redundancy. Removal of key species can alter predator-prey relationships and reduce ecosystem stability.

4.3 Bioindicators

Butterflyfish diversity and abundance are commonly used as indicators of coral reef health because of their dependence on live coral cover.

Overharvesting ornamental fishes can disrupt these ecological interactions, potentially leading to cascading effects on reef structure.

5. Economic Importance

Although marine ornamentals account for a smaller volume compared to freshwater ornamentals, they command higher per-unit prices. The global marine ornamental trade is valued at several hundred million US dollars annually.

5.1 Export Markets

Major importing countries include:

- United States
- Germany
- United Kingdom
- Japan

India's marine ornamental exports are relatively small but growing.

5.2 Livelihood Contribution

Coastal fishers, particularly in Tamil Nadu and island territories, engage in ornamental fish collection during lean fishing seasons. This supplementary income can enhance livelihood resilience if properly regulated.

6. Current Status in India

6.1 Trade & Export

Marine ornamental fish exports from India are modest compared to Southeast Asian nations (e.g., Philippines, Indonesia), but increasing. Major centers include:

- Tamil Nadu: Rameswaram, Mandapam
- Kerala: Kochi, Thiruvananthapuram
- Andaman & Nicobar Islands

Export data shows steady growth but is often underreported due to informal trade channels. There is a severe *lack of centralized monitoring*.

6.2 Major Species Traded

Commonly traded marine ornamentals include:

- Clownfishes (e.g., *Amphiprion ocellaris*)
- Blue-green Chromis (*Chromis viridis*)
- Royal Gramma (*Gramma loreto*)
- Yellowtail Damsel (*Chrysiptera parasema*)
- Mandarinfish (*Synchiropus splendidus*)

Quantitative data on catch volumes and population trends are limited owing to inadequate field surveys.

6.3 Harvesting Practices

Common methods include:

- Hand nets
- Barrier nets
- Scoop nets

Destructive practices such as cyanide use are rare in India compared to some Southeast Asian countries but require vigilance.

6.4 Export Volume

Official export data are limited, and underreporting is common due to fragmented supply chains. The lack of a centralized monitoring system hampers accurate assessment.

7. Major Problems and Constraints

7.1 Overexploitation

Selective removal of high-value species can reduce breeding populations and impact local reef dynamics.

7.2 Habitat Degradation

Coral reefs in India face threats from:

- Coastal development
- Pollution
- Sedimentation
- Coral mining (historically)
- Climate-induced bleaching

7.3 Climate Change

Mass bleaching events in Lakshadweep and the Andaman region have reduced live coral cover, affecting habitat-dependent ornamental fishes.

7.4 Post-Harvest Mortality

Improper handling, stress, and inadequate acclimatization lead to mortality during transport.

7.5 Policy Gaps

Marine ornamental fisheries are not comprehensively regulated under existing fisheries laws. Absence of species-specific quotas and weak enforcement remain challenges.

8. Management Strategies for Sustainable Development

To ensure ecological sustainability and enhance economic benefits, an integrated management approach is needed.

8.1 Policy and Institutional Framework

- Establish national marine ornamental fisheries policy: aligned with biodiversity conservation goals.
- Permit and quota systems: science-based harvest limits with traceability.
- Co-management models: involve community cooperatives in decision-making.
- Certification schemes: eco-labeling to promote sustainably sourced ornamentals.

8.2. Regulatory Framework in India

Relevant policies include:

- Wildlife Protection Act (1972) – protects certain marine species.
- Coastal Regulation Zone (CRZ) notifications.
- Marine Fishing Regulation Acts (state-level).
- Biodiversity Act (2002).

However, a dedicated national marine ornamental fishery policy is lacking.

8.3 Ecological and Biological Research

Invest in research on:

- Population assessments.
- Species reproductive biology.
- Habitat mapping using GIS tools.
- Larval rearing and aquaculture techniques.

8.4 Capacity Building & Livelihood Support

- Train fishers in sustainable collection techniques (e.g., hook-and-line, barrier nets).
- Establish aquaculture hatcheries for high-value species to reduce pressure on wild stocks.
- Support value addition and direct marketing to reduce dependence on intermediaries.

8.5 Awareness & Education

- Conduct community workshops on reef conservation.
- Promote consumer awareness on sustainable aquarium fish trade.
- Integrate ornamental fish conservation into marine protected area (MPA) planning.

8.6 Monitoring & Enforcement

- Use mobile reporting tools for real-time catch data.
- Strengthen patrolling and compliance in reef areas.
- Implement traceability systems from collection to export.

8.7 Certification and Eco-Labeling

Eco-certification can promote sustainable sourcing and fetch premium prices.

9. Major Problems and Constraints

Despite potential, multiple problems limit sustainable development:

9.1 Ecological Threats

- **Overharvesting:** Targeted removal of high-value species reduces populations and disrupts ecosystem balance.
- **Destructive collection techniques:** Use of poisons or unsustainable nets harms corals and non-target organisms.
- **Habitat degradation:** Coastal development and pollution threaten reef habitats.
- **Climate change:** Coral bleaching events directly reduce habitat availability.

9.2 Socio-Economic Issues

- **Lack of community rights:** Fishers often lack formal rights and depend on middlemen.
- **Seasonal fluctuations:** Income instability due to seasonal coral accessibility.
- **Low local value capture:** Most economic benefits accrue to exporters rather than collectors

9.3 Policy & Regulatory Gaps

- **Inadequate legal frameworks:** Current fisheries laws do not comprehensively address ornamental collection.
- **Poor enforcement:** Limited monitoring capacity leads to illegal, unreported, and unregulated (IUU) harvesting.

9.4 Knowledge & Research Deficits

- **Taxonomic gaps:** Accurate identification of species is limited.
- **Population dynamics unknown:** No long-term studies on stock status.
- **Post-harvest handling:** High mortality during transit due to inadequate handling procedures.

10. Aquaculture Potential

Marine ornamental aquaculture is a promising alternative.

10.1 Advantages

- Reduces wild collection pressure
- Ensures supply stability
- Enhances export quality

10.2 Challenges

- Broodstock management
- Larval survival rates
- Live feed production (rotifers, Artemia)

India has demonstrated success in captive breeding of clownfish species, indicating strong potential.

11. SWOT Analysis

Strengths

- Rich biodiversity
- Long coastline
- Growing global demand

Weaknesses

- Limited hatchery infrastructure
- Poor data collection

Opportunities

- Blue economy initiatives
- Sustainable certification markets

Threats

- Climate change
- Habitat destruction
- Illegal trade

12. Future Prospects

India's marine ornamental sector can be strengthened by:

- Investing in research and development
- Establishing national monitoring databases
- Integrating ornamental fisheries into blue economy planning
- Encouraging public-private partnerships

13. Conclusion

Marine ornamental fishes represent a valuable yet vulnerable component of India's marine biodiversity and coastal economy. While the country possesses substantial natural resources, the sector remains underdeveloped due to regulatory, ecological, and infrastructural challenges. Sustainable management requires a multi-dimensional strategy integrating ecosystem conservation, scientific assessment, community participation, aquaculture development, and market-based incentives. Strengthening policy frameworks and promoting responsible trade practices can transform the marine ornamental fish sector into a model of biodiversity-based sustainable development. Without proactive intervention, overexploitation and environmental change may compromise both reef ecosystems and dependent livelihoods. Therefore, immediate action guided by science and stakeholder collaboration is essential to secure the future of marine ornamental fisheries in India.

14. References

- 1) Allen, G. R., & Werner, T. B. (2002). *Coral reef fish assessment in the aquarium trade. Marine Policy*, 26(2), 123–129.
- 2) Bruckner, A. W. (2005). *The importance of the marine ornamental reef fish trade. Marine Pollution Bulletin*, 50(5), 478–483.
- 3) FAO. (2020). *The State of World Fisheries and Aquaculture. Food and Agriculture Organization of the United Nations*.
- 4) Rao, G. S. (2010). *Status of marine ornamental fish trade in India. CMFRI Bulletin*, 47, 65–72.
- 5) Rhyne, A. L., Tlusty, M. F., & Kaufman, L. (2014). *Long-term trends of coral reef fish trade. Aquaculture*, 431, 52–59.
- 6) Venkataraman, K., & Rajan, P. T. (2013). *Marine biodiversity of India. Zoological Survey of India*.

A Review on use of Spirulina for Ornamental Fishes

S. Rinna Hamlin

Assistant Professor, Department of Zoology, Nesamony memorial Christian college, Marthandam, Kanyakumari district.

Corresponding Author Email: rinnacharles@gmail.com

Abstract

Spirulina is one of the most concentrated natural source of nutrition for all the animals. Spirulina has 60-70% protein by weight and is the richest source of vitamin B12, β -carotene, essential fatty acids and minerals. Besides, there are eight amino acids such as isoleucine, leucine, lysine, methionine, phenylalanine, threonine, tryptophane and valine in Spirulina. More recently, there has been new interest concerning the therapeutic effects of Spirulina as growth promotor and probiotic or booster for the immune system in animals and fish. In short, Spirulina is a powerful tonic for the immune system. Many authors have studied the effect of Spirulina diet or extract on growth and immune responses in various

Keywords: *Growth, Reproduction, Spirulina and Pigmentation*

Introduction

Ornamental fishes are beautiful pets compared to many other pet animals that are maintained in aquaria (Biswas *et al.*, 2007). Keeping colourful tropical fishes in aquarium and garden ponds is a hobby since centuries by humans. The global ornamental fishes export is also escalating in marine and freshwater fish valued at US\$ 337 million in the year 2017. Colour is one of the major factors, which determine the value of ornamental fish in the global market. Like other animals, fish cannot synthesize carotenoids in their body, and fishes must obtain them via food. Therefore, carotenoid supplementation is needed to enhance the pigmentation in ornamental fishes. Ornamental fishes are mainly characterized by a wide diversity of colours, colour patterns and success of ornamental fish trade is very much dependent on the bouncy colour of the fish. Pigmentation in the skin is responsible for coloration of fish.

The carotenoids act as a vital nutrient for healthy growth, metabolism, and reproduction as well as colour. It is used in aquaculture feed to provide the colour associated

with consumer product, such as the bright vibrant colours of ornamental fish (Mukherjee *et al.*, 2010). Carotenoids are the primary source of the pigmentation on the skin of fishes. In natural environment, the fishes meet their carotenoid requirements by ingesting aquatic plants or through their food chains. More than 600 kinds of carotenoid are found in the nature, but few of them are used in animal food, medications, colour of food, polish chemicals (Bricaud *et al.*, 1998). The colour enhancing diets should contain additional natural pigment to enhance the colour of the ornamental fishes. The freshwater ornamental fish industry has experienced the problem of faded coloration in fish, especially when the fishes are kept under captivity for long duration and also in intensive culture condition.

Enhancement of colouration is not only confined in ornamental fish industry, several scientists are engaged in improvement of colouration of muscle or skin of food fish as it is an important factor which determines the price of that fish in market. In spite of proper size, if the colour of the fish is not up to the mark, then the fish will not get a good market price. If enhancement of coloration can be done by administering pigment enriched feed, it will definitely improve the quality and cost of the fish. Recent efforts have focused on natural compounds as alternative to synthetic carotenoids as because of concerns about the use of synthetic additives and their high cost. To alleviate this problem the present study was made to evaluate pigmentation quality using the natural carotenoids. However, detailed studies on color enrichment through spirulina powder in ornamental fishes are lacking.

Algae, as the earliest life forms on Earth, reproduce independently, providing nutrients for the growth of other producers and the next trophic levels. Besides this, they also generate oxygen as a byproduct of their growth, contributing to around 70% of the Earth's free oxygen (Tietze, 2004). Spirulina, *Arthrospira platensis*, green-blue microalgae, is a symbiotic, multicellular, filamentous organism associated with nitrogen-fixing bacteria. Its unique features include the pigment phycocyanin for photosynthesis, giving it a distinct blue colour. Spirulina's reproduction involves binary fission, and its spiral configuration forms floating mats. It appeared approximately 3.6 billion years ago, utilizing dissolved carbon dioxide present in seawater as a source of nutrients (Vo *et al.*, 2015).

Microalgae play a vital role in the aquatic food chain, widely used in aquaculture for the growth of aquatic animals. For example, microalgae serve as a primary food source for the larval stages of many aquatic species and contribute significantly to the overall health and

productivity of aquatic ecosystems by improving water quality (Ma & Hu, 2023; Cai *et al.*, 2021). The chemical composition of algae provides basic information about their trophic potential. Microalgae are gaining popularity in aquaculture due to their moderate size, high nutritional value, rapid growth, and robust resistance to antioxidants and diseases (Mishra *et al.*, 2022 and Habib *et al.*, 2008).

Spirulina is rich in nutrients that transform this alga into a valuable food source and through the years, extensive studies on biochemical composition, safety consumption, physiological effects of enriched diets on animal organisms, and toxicity testing were carried out and proved that Spirulina is a useful feed ingredient for both human and animal consumption (Anvar & Nowruzi, 2021; Habib *et al.*, 2008)

One ingredient that has been proven to have a high nutritional content and can increase the rate of production and survival rate of various types of fish is *Spirulina platensis* (Adel *et al.*, 2016). One ingredient that has been proven to have a high nutritional content and can increase the rate of production and survival rate of various types of fish is *Spirulina platensis*. Therefore, this article aims to explain the potential of *Spirulina platensis* as an alternative fish feed ingredient and various other ingredients. One ingredient that has been proven to have a high nutritional content and can increase the rate of production and survival rate of various types of fish is *Spirulina platensis* (Joseph. 2013). Therefore, this article aims to explain the potential of *Spirulina platensis* as an alternative fish feed ingredient.

***Spirulina* as an alternative Fish Feed:**

Fishmeal is an essential ingredient as a protein source in the diet of fish (FAO, 2016). Although fishmeal is most nutritious and easily digestible ingredient, use of it in high amounts is not sustainable due to its high price. For this reason, an alternative feed ingredient which contains high protein is needed to replace fishmeal. *Arthrospira platensis* a microalga, is an alternative plant-derived protein source that could replace costly animal-derived protein. Research has shown that consumption of spirulina improves growth performance in several fish species like Coral trout, *Plectropomus leopardus* (Yu *et al.*, 2018), great sturgeon, *Huso huso* (Adel *et al.*, 2016), gibel carp, *Carassius auratus gibelio* (Cao *et al.*, 2018), Nile tilapia, *Oreochromis niloticus* (Elabd *et al.*, 2020; Shalata *et al.*, 2021), blunt snout bream, *Megalobrama amblycephala* (Jiang *et al.*, 2022), Catfish, *Clarias gariepinus* (Rosenau *et al.*, 2021).

The excellent nutritional value of spirulina makes it alternative protein source and growth promoter due to its high protein, diverse amino acid (AA) and fatty acid (FA), vitamins, minerals and several bioactive molecules. In addition, Spirulina has been proven to be an excellent feed attractant in shrimp in very low inclusion in diets (Silva-Neto *et al.*, 2012). There are many research works available about effect of spirulina on growth performances in non-ornamental fish species and these have been used to determine the effect of spirulina on growth and colouration of ornamental fish. That's why there is a lack of information about the nutrient needs of ornamental fish.

Spirulina is not only rich in nutrients but also serves as a source of phytopigments, including chlorophyll, carotenoids, and phycobiliproteins. These pigments play a vital role in capturing light within Spirulina. In 2013, Kuddus *et al.*, found that the phycobiliproteins extracted from Spirulina are phycocyanin (C-PC), allophycocyanin (A-PC), and phycoerythrin (PE). Later, it was determined that phycocyanin represented up to 50% of phycobiliprotein content (Li *et al.*, 2020). High protein content percentage of spirulina and its well-balanced amino acid profile compared with other plant protein sources makes it as potential fish meal replacer in the aquafeed formulation. Different types of synthetic carotenoids, such as astaxanthin, canthaxanthin and lutein (Hanel *et al.* 2007), as well as natural carotenoid sources have been used as dietary supplements to increase the colouration and pigmentation of ornamental fish (Gouveia and Rema, 2005).

James *et al.* (2006) reported that the maximum carotenoid content was obtained in ornamental red swordtail (*Xiphophorus helleri*) fed with 8% spirulina. Skin coloration is one of the most important factors which decide the aesthetic value, therefore the market value of ornamental fishes. Due to this valuable composition of useful nutrients, Spirulina became widely a nutritional supplement suitable for animal feed and humans, too (Kim, 2013). Therefore, from the perspective of the biochemical composition, it has been proved that Spirulina is an excellent source of beneficial nutrients and healthful substances, as well as a good source of energy, making it a suitable addition to feed formulations.

Conclusion

In Conclusion, *Spirulina platensis* contain a lot of nutrition such as amino acid, protein, carbohydrate, minerals, and vitamins. The nutrition content in *Spirulina platensis* can be influenced by medium composition. *Spirulina platensis* can be used as potential

alternative fish feed. However, careful consideration of optimal dosage and avoidance of over-supplementation are imperative to prevent adverse effects on fish health. The advantage of spirulina powder is that, it's easily available in local market and can be smoothly incorporate with feed ingredients. Study revealed that, spirulina powder not only useful in enhancement of body pigmentation in fishes but also supports of growth efficiency in fishes. Consequently, Spirulina emerges as a promising stimulant in aquaculture, for commercial application.

References

- 1) Mukherjee A, Mandal B, Banerjee S. 2010. Turmeric as a carotenoid source on pigmentation and growth of fantail guppy, *Poecilia reticulata*. *Proceedings of the Zoological Society*. 62(2):119-123.
- 2) Tietze, H.W. 2004. *Spirulina Micro Food Macro Blessing*. 4th Edition, Harald W. Tietze Publishing, Australia.
- 3) Vo, T.S., Ngo, D.H. and Kim, S.K., 2015. Nutritional and pharmaceutical properties of microalgal *Spirulina*, in: *Handbook of marine microalgae: Biotechnology advances*. Elsevier Inc., pp. 299–308.
- 4) Ma, M., & Hu, Q. 2023. *Microalgae as feed sources and feed additives for sustainable aquaculture: Prospects and challenges*. *Reviews in Aquaculture*, 1- 18.
- 5) Cai, J., Lovatelli, A., Aguilar-Manjarrez, J., Cornish, L., Dabbadie, L., Desrochers, A., Yuan, X. 2021. *Seaweeds and microalgae: an overview for unlocking their potential in global aquaculture development*. *FAO Fisheries and Aquaculture Circular*, (1229).
- 6) Mishra, B., Tiwari, A., & Mahmoud, A. E. D. 2022. *Microalgal potential for sustainable aquaculture applications: bioremediation, biocontrol, aquafeed*. *Clean Technologies and Environmental Policy*, 1-13.
- 7) Habib, M.A.B.; Parvin, M.; Huntington, T.C.; Hasan, M.R. 2008. *A review on culture, production and use of spirulina as food for humans and feeds for domestic animals and fish*. *FAO Fisheries and Aquaculture Circular*, No. 1034, Rome, 33 p.
- 8) Anvar, A. A., & Nowruzi, B. 2021. *Bioactive Properties of Spirulina: A Review*. *Microbial Bioactives*, 4(1), 134– 142.
- 9) Joseph I. 2013. *Aquaculture feeds and feeding: Major challenges and issues*. *Proceedings of the CMFRI SAARC. International Workshop*, 271-276.

- 10) Adel M, Yeganeh S, Dadar M, Sakai M, Dawood MAO. 2016. Effects of dietary *Spirulina platensis* on growth performance, humoral and mucosal immune responses and disease resistance in juvenile great sturgeon (*Huso huso* Linnaeus, 1754). *Fish and Shellfish Immunology*, 56:436-444.
- 11) FAO, 2016. *The State of World Fisheries and Aquaculture 2016*. www.fao.org/publications.
- 12) Yu.P, Y. Wu, G. Wang, T. Jia, Y. Zhang. 2017. Purification and bioactivities of phycocyanin. *Critical Reviews in Food Science and Nutrition*, 57 (18), pp. 3840-3849,
- 13) Adel, M., Yeganeh, S., Dadar, M., Sakai, M. and Dawood, M.A.O. 2016. Effects of dietary *Spirulina platensis* on growth performance, humoral and mucosal immune responses and disease resistance in juvenile great sturgeon (*Huso huso* Linnaeus, 1754). *Fish and Shellfish Immunology*. 56: 436-444.
- 14) Elabd, H., Wang, H.P., Shaheen, A. and Matter, A. 2020. Nano spirulina dietary supplementation augments growth, antioxidative and immunological reactions, digestion and protection of Nile tilapia, *Oreochromis niloticus*, against *Aeromonas veronii* and some physical stressors. *Fish Physiology and Biochemistry*. 46(6): 2143-2155.
- 15) Cao, S., Zhang, P., Zou, T., Fei, S., Han, D., Jin, J., Liu, H., Yang, Y., Zhu, X. and Xie, S. 2018. Replacement of fishmeal by spirulina *Arthrospira platensis* affects growth, immune related-gene expression in gibel carp (*Carassius auratus gibelio* var. CAS III) and its challenge against *Aeromonas hydrophila* infection. *Fish and Shellfish Immunology*. 79: 265-273.
- 16) Shalata, H.A., Bahattab, O., Zayed, M.M., Farrag, F., Salah, A.S., Al-Awthman, Y.S., Ebied, N.A. and Mohamed, R.A. 2021. Synergistic effects of dietary sodium butyrate and *Spirulina platensis* on growth performance, carcass composition, blood health and intestinal histomorphology of Nile tilapia (*Oreochromis niloticus*). *Aquaculture Reports*. 19: 10063
- 17) Jiang, W., Miao, L., Lin, Y., Ci, L., Liu, B. and Ge, X. 2022. *Spirulina platensis* as a protein source could improve growth, feed utilisation and digestion and physiological status in juvenile blunt snout bream (*Megalobrama amblycephala*). *Aquaculture Reports*. 22: 100932.

- 18) Rosenau, S., Oertel, E., Dietz, C., Wessels, S., Tetens, J., Mörlein, D. and Ciulu, M. 2021. Total replacement of fishmeal by spirulina (*Arthrospira platensis*) and its effect on growth performance and product quality of african catfish (*Clarias gariepinus*). *Sustainability (Switzerland)*. 13 (16).
- 19) Li, Y., Aiello, G., Bollati, C., Bartolomei, M., Arnoldi, A., & Lammi, C. 2020. Phycobiliproteins from *Arthrospira platensis* (*Spirulina*): A new source of peptides with dipeptidyl peptidase-IV inhibitory activity. *Nutrients*, 12(3), 1–11.
- 20) Kim, S., Rahimnejad, S., Kim, K., Lee, K. 2013. Partial replacement of fish meal with *Spirulina pacifica* in diets for parrot fish (*Oplegnathus fasciatus*). *Turkish Journal of Fisheries & Aquatic Science*, 13; 1-11.
- 21) Silva-Neto, J.F., Nunes, A.J.P., Sabry-Neto, H. and Sá, M.V.C. 2012. *Spirulina* meal has acted as a strong feeding attractant for *Litopenaeus vannamei* at a very low dietary inclusion level. *Aquaculture Research*. 43(3): 430-437.
- 22) Kuddus, M., Singh, P., Thomas, G., & Al-Hazimi, A. 2013. Recent developments in production and biotechnological applications of C-phycoyanin. *BioMed research international*. pp.1-9, July 2013.
- 23) Hanel, R., Broekman, D., Graaf, S., Schnack, D. 2007. Partial replacement of fishmeal by lyophilized powder of the microalgae *Arthrospira platensis* in pacific white shrimp diets. *The Open Marine Biology Journal*, 1; 1-5.
- 24) Gouveia, L., Rema, P. 2005. Effect of microalgal biomass concentration and temperature on ornamental goldfish (*Carassius auratus*) skin pigmentation. *Aquaculture Nutrition*, 11; 19-23.
- 25) James, R., Kunchitham, S., Ramasamy, T., Vasudhevan, I. 2006. Effect of Dietary *Spirulina* Level on Growth, Fertility, Coloration and Leucocyte Count in Red Swordtail (*Xiphophorus Helleri*). *The Israel Journal of Aquaculture Bamidgeh*, 58; 97-104

Pigment Studies in Ornamental Fishes

Greeshma Thomas

Research Scholar, Department of Zoology, S.T. Hindu College, Nagercoil, Tamil Nadu, India

Affiliated to Manonmaniam Sundaranar University, Tirunelveli.

Corresponding Author Email: greeshmathomas5757@gmail.com

Abstract

Pigmentation in ornamental fishes represents a complex interplay of biochemical, physiological, genetic, and environmental processes that collectively determine body coloration and pattern formation. The aesthetic appeal of ornamental fishes, which forms the backbone of their commercial value, is primarily governed by the distribution and concentration of pigments within specialized dermal cells known as chromatophores. This chapter provides an in-depth exploration of pigment types, chromatophore biology, molecular pathways, and environmental and nutritional influences on coloration in ornamental fishes. Special emphasis is placed on carotenoid metabolism, melanin biosynthesis, and structural coloration mechanisms mediated by light-reflecting cells. Furthermore, recent advancements in pigment enhancement strategies, including dietary supplementation, selective breeding, and molecular interventions, are critically discussed. Understanding these mechanisms is essential for optimizing ornamental fish quality, improving aquaculture sustainability, and advancing research in fish physiology and biotechnology.

Keywords: *Chromatophores, Pigment metabolism, Ornamental fish coloration, Carotenoid and melanin biosynthesis, Pigmentation enhancement strategies*

Introduction

Ornamental fishes represent one of the most vibrant and economically significant sectors of the global aquaculture industry. Their aesthetic appeal, driven primarily by body coloration and pattern diversity, plays a crucial role in determining their market value and consumer preference. Coloration in ornamental fishes is not merely a superficial trait but a complex biological phenomenon influenced by genetic, physiological, environmental, and nutritional factors.

Fish coloration arises from specialized pigment-containing and light-reflecting cells known as chromatophores, which are located in the dermal layers of the skin. These chromatophores are broadly classified into different types based on the pigments they contain or their optical properties. The major chromatophore types include melanophores (black or brown pigments), xanthophores (yellow pigments), erythrophores (red pigments), iridophores (reflective or iridescent cells), and leucophores (white pigments). The interaction and distribution of these chromatophores contribute to the wide spectrum of colors and patterns observed in ornamental fish species.

In recent years, pigment studies in ornamental fishes have gained considerable attention due to their implications for aquaculture, fish health, and commercial value. Enhancing coloration through dietary manipulation and optimized rearing conditions has become a key focus area for researchers and industry stakeholders. Furthermore, understanding pigment biology contributes to advancements in selective breeding programs aimed at developing strains with superior coloration traits.

Classification and Biochemistry of Pigments Carotenoids: Classification and Biochemistry

Carotenoids are a major class of natural pigments widely responsible for the yellow, orange, and red coloration observed in many ornamental fishes. They are lipid-soluble compounds synthesized by plants, algae, fungi, and some microorganisms, but cannot be synthesized *de novo* by fish.

Classification of Carotenoids

Carotenoids are broadly classified into two main groups based on their chemical structure and functional groups:

a) Carotenes

Carotenes are purely hydrocarbons composed of carbon and hydrogen atoms. They are non-polar molecules and do not contain oxygen.

- Examples: β -carotene, α -carotene, lycopene
- Function: Serve as precursors of vitamin A and contribute to orange coloration
- Characteristics: Highly lipophilic and relatively less reactive compared to oxygenated carotenoids

b) Xanthophylls

Xanthophylls are oxygenated derivatives of carotenes, containing functional groups such as hydroxyl (-OH), keto (=O), or epoxy groups.

- Examples: astaxanthin, lutein, zeaxanthin, canthaxanthin
- Function: Provide yellow, orange, and red pigmentation
- Characteristics: More polar than carotenes and more effective in coloration of fish

Among these, astaxanthin is considered the most important carotenoid in ornamental fish culture due to its strong red pigmentation and high deposition efficiency.

Importance in Ornamental Fish Culture

Carotenoids are crucial for enhancing the visual appeal and market value of ornamental fishes. Since natural pigmentation may fade under captive conditions, dietary supplementation with carotenoid-rich sources such as:

- Microalgae (e.g., spirulina)
- Crustacean meals
- Synthetic carotenoids (astaxanthin, canthaxanthin) is commonly practiced in aquaculture

Melanins: Classification and Biochemistry

Melanins are a major group of natural pigments responsible for black, brown, and sometimes gray coloration in ornamental fishes. They are widely distributed across the animal kingdom and play essential roles not only in coloration but also in protection against environmental stressors such as ultraviolet (UV) radiation and oxidative damage. In fishes, melanins are synthesized internally and stored in specialized pigment cells known as melanophores.

Classification of Melanins

Melanins are broadly classified into three main types based on their chemical composition and color:

a) Eumelanin

- Color: Black to dark brown
- Composition: Derived from the oxidation of tyrosine via intermediates such as DOPA (dihydroxyphenylalanine) and dopaquinone

- Function: Responsible for dark pigmentation and pattern formation in fishes
- Significance: Most abundant type of melanin in fish skin

b) Pheomelanin

- Color: Yellowish-brown to reddish
- Composition: Formed when cysteine reacts with dopaquinone during melanin synthesis
- Function: Contributes to lighter shades and reddish hues
- Significance: Less common in fishes compared to eumelanin

c) Neuromelanin

- Color: Dark pigment found in neural tissues
- Occurrence: Primarily in the brains of vertebrates
- Significance: Limited relevance in fish skin pigmentation but important in neurological functions

Biological Functions of Melanins

Melanins serve multiple important functions in ornamental fishes:

- **Coloration and pattern formation:** Essential for species identification and ornamental value
- **Camouflage:** Helps fish blend with their surroundings
- **Photoprotection:** Absorbs harmful UV radiation
- **Antioxidant activity:** Protects tissues from oxidative stress
- **Thermoregulation:** Dark pigments can aid in heat absorption

Importance in Ornamental Fish Culture

Melanin-based coloration is critical for defining patterns such as stripes, spots, and patches seen in ornamental fishes. Unlike carotenoids, melanins are synthesized internally, so their expression depends more on:

- Genetic selection
- Environmental management
- Stress reduction

Proper husbandry practices—such as maintaining optimal water quality, suitable lighting, and low-stress conditions—help in preserving stable melanin-based coloration.

Pteridines: Classification and Biochemistry

Pteridines are an important group of naturally occurring pigments that contribute to the yellow, orange, red, and sometimes white coloration in ornamental fishes. Unlike carotenoids, which must be obtained from the diet, **pteridines are synthesized endogenously** within the fish body. They are primarily found in specialized pigment cells called xanthophores and erythrophores, where they often interact with other pigments to produce a wide range of color variations.

Classification of Pteridines

Pteridines are classified based on their chemical structure and functional roles. In fish pigmentation, the most relevant groups include:

a) Colored Pteridines

These are directly responsible for visible pigmentation.

- Examples: drosopterins, erythropterins, xanthopterins
- Color: Yellow, orange, red
- Function: Contribute to bright coloration, often enhancing or modifying carotenoid-based colors

b) Colorless or Weakly Colored Pteridines

These compounds may not directly impart strong color but play roles in pigment metabolism and light reflection.

- Examples: biopterin, neopterin
- Function: Act as metabolic intermediates and cofactors in biochemical pathways

Biological Functions of Pteridines

Beyond coloration, pteridines have several physiological roles:

- Color enhancement: Intensify and diversify body coloration
- UV protection: Absorb harmful radiation
- Metabolic roles: Some act as cofactors in enzymatic reactions
- Signaling and communication: Bright colors may play roles in mating and species recognition

Importance in Ornamental Fish Culture

Pteridines are important for maintaining stable and vivid coloration in ornamental fishes. Since they are synthesized within the body:

- Their production depends on genetic factors and metabolic health
- Environmental conditions (light, stress, water quality) can influence their expression
- Nutritional status indirectly affects their synthesis by supporting metabolic pathways

Although dietary supplementation does not directly supply pteridines, providing balanced nutrition helps maintain their biosynthesis and overall pigment expression.

Purines and Structural Pigments: Classification and Biochemistry

In addition to carotenoids, melanins, and pteridines, ornamental fish coloration is also influenced by purine-based pigments and structural (physical) coloration mechanisms. These components are especially important for producing white, silvery, and iridescent appearances that enhance the visual appeal of many ornamental species.

Purine Pigments

a) Overview

Purine pigments are nitrogen-containing compounds derived from purine metabolism. In fishes, the most important purine involved in coloration is guanine, which contributes to silvery, white, and reflective coloration rather than vivid hues.

b) Classification of Purine Pigments

Purine compounds in fish pigmentation mainly include:

- Guanine → principal pigment responsible for reflectivity
- Hypoxanthine and xanthine → minor roles in pigmentation

Among these, guanine is the most significant due to its optical properties.

Biological Functions

- Camouflage: Reflective surfaces help fish blend into aquatic environments
- Light reflection: Reduces visibility to predators
- Visual signaling: Enhances brightness and attractiveness

Structural Pigments (Structural Coloration)

a) Overview

Structural coloration arises not from pigments themselves but from physical interactions between light and microscopic structures in fish tissues. These structures manipulate light through reflection, refraction, scattering, and interference.

b) Types of Structural Coloration

i) Iridescence

- Produced by multilayered reflecting structures (often guanine crystals)
- Color changes with viewing angle
- Common in species like tetras and zebrafish

ii) Scattering-Based Coloration

- Produced by random structures that scatter light
- Results in white or bluish appearance

iii) Interference Coloration

- Occurs when light waves overlap and interfere
- Produces shimmering colors

c) Cellular Basis

Structural coloration is mainly associated with:

- **Iridophores** → contain guanine crystals
- **Leucophores** → scatter light to produce white coloration

These cells do not rely on chemical pigments but on microstructural organization.

d) Biophysical Mechanism

Structural colors are produced through:

- **Reflection:** Light bouncing off crystal layers
- **Refraction:** Bending of light as it passes from one medium to another due to a change in its speed
- **Interference:** Interaction of light waves
- **Diffraction:** Bending and spreading of light waves as they pass around an obstacle or through a small opening.

The precise arrangement and spacing of structures determine the resulting colour.

e) Interaction with Pigments

Structural coloration often works in combination with chemical pigments:

- With melanins → enhances contrast and depth
- With carotenoids and pteridines → intensifies brightness
- With purines → produces metallic and iridescent effects

f) Importance in Ornamental Fish

Structural pigments are crucial for:

- Metallic shine in goldfish and koi
- Iridescent colours in guppies and bettas
- Pearl and glossy effects in many aquarium species

Their presence significantly increases the aesthetic and commercial value of ornamental fishes.

Chromatophore Biology and Organization

Chromatophores are specialized pigment-containing and light-reflecting cells located in the dermal layers of fish skin. Their arrangement and density determine the visible coloration.

Types	Pigment	Colour	Function
Melanophores	Melanin	Black/ Brown	Protection, Camouflage
Xanthophores	Carotenoids	Yellow	Visual Signaling
Erythrophores	Carotenoids	Red	Mate Attraction
Iridophores	Guanine	Iridescent	Light Reflection
Leucophores	Purines	White	Background Colouration

Chromatophores can rapidly change color intensity through pigment dispersion and aggregation, a process controlled by neural and hormonal signals.

Mechanisms of Color Production

Pigmentary Mechanisms

Colour production in ornamental fishes is primarily achieved through pigmentary mechanisms, which involve the presence, synthesis, and distribution of chemical pigments within specialized skin cells called chromatophores. These pigments absorb specific wavelengths of light and reflect others, resulting in the wide range of colours observed in fish.

Chromatophores and their Role

Pigmentary coloration is controlled by different types of chromatophores located in the dermal layers of the skin. Each type contains specific pigments:

- **Melanophores** → contain melanin (black/brown coloration)
- **Xanthophores** → contain carotenoids and pteridines (yellow coloration)
- **Erythrophores** → contain carotenoids and pteridines (orange/red coloration)
- **Leucophores** → contain purine crystals (white appearance)

The type, density, and arrangement of these chromatophores determine the overall coloration and patterns in ornamental fishes.

Nature of Pigments and Light Absorption

Pigments produce colour by selective absorption of light:

- They absorb certain wavelengths of visible light
- Reflect or transmit the remaining wavelengths

For example:

- Carotenoids absorb blue and green light → reflect yellow, orange, and red
- Melanin absorbs most wavelengths → appears black or dark

Thus, the observed colour depends on which wavelengths are reflected back to the observer.

Pigment Synthesis and Source

Pigments in fishes originate from two main sources:

a) Endogenous Pigments

- Synthesized within the fish body
- Examples: melanin, pteridines, purines
- Controlled by genetic and enzymatic processes

b) Exogenous Pigments

- Obtained from diet
- Example: carotenoids
- Require proper nutrition for deposition and maintenance

Pigment Distribution and Movement

Chromatophores can change colour intensity through pigment movement:

- **Dispersion of pigment granules** → darker or more intense coloration
- **Aggregation of pigment granules** → lighter appearance

This process is known as physiological colour change and is regulated by:

- Nervous system
- Hormones (e.g., melanocyte-stimulating hormone)
- Environmental stimuli (light, background, stress)

Layering and Interaction of Pigments

Fish coloration is often the result of multiple pigment layers interacting:

- Upper layers: xanthophores and erythrophores
- Middle layers: iridophores (reflective)
- Lower layers: melanophores

This layered arrangement allows:

- Mixing of colours
- Enhancement of brightness
- Formation of complex patterns (spots, stripes, bands)

Factors Affecting Pigmentary Coloration

Several factors influence pigment-based coloration:

- **Genetics** → determines baseline colour and pattern
- **Nutrition** → affects carotenoid availability
- **Light intensity and background** → influence pigment expression
- **Water quality and temperature** → affect physiological processes
- **Stress and health status** → can reduce colour intensity

Biological Significance

Pigmentary coloration serves multiple functions:

- **Camouflage** → protection from predators

- **Communication** → mating and social signalling
- **Species recognition**
- **Commercial value** → key factor in ornamental fish trade

Factors Influencing Pigmentation

Genetic Regulation

Pigmentation is under strict genetic control involving multiple genes that regulate chromatophore differentiation, pigment synthesis, and pattern formation. Mutations in these genes can lead to color variations and morphs.

Nutritional Factors

Diet plays a pivotal role in pigmentation, particularly for carotenoids. Deficiency results in faded coloration, while supplementation enhances vibrancy.

Environmental Factors

Environmental conditions significantly influence pigmentation:

- Light intensity affects pigment synthesis
- Tank background influences perceived coloration
- Water quality impacts pigment deposition

Hormonal Control

Hormones such as melanocyte-stimulating hormone (MSH) regulate pigment dispersion within chromatophores. Stress hormones like cortisol may cause color fading.

Nutritional Strategies for Color Enhancement

Modern aquaculture employs various strategies to enhance pigmentation:

- Inclusion of natural pigments (spirulina, marigold extract)
- Use of synthetic carotenoids
- Formulation of balanced feeds for optimal absorption

Molecular Insights into Pigmentation

Recent advances in molecular biology have revealed the genetic and biochemical basis of pigmentation:

- MITF gene regulates melanophore development
- Tyrosinase gene (TYR) controls melanin synthesis
- Carotenoid-binding proteins facilitate pigment transport

Gene expression studies and transcriptomic analyses are increasingly used to understand pigmentation pathways.

Industrial Applications

Pigment studies have direct implications in the ornamental fish industry:

- Enhancement of fish market value
- Development of designer fish varieties
- Standardization of feed formulations
- Export quality improvement

Challenges and Future Directions

Challenges

- High cost of carotenoid supplements
- Uneven pigment deposition
- Environmental stress-induced color loss

Future Directions

- Genetic engineering for stable coloration
- Sustainable natural pigment production
- Use of probiotics for improved pigment absorption
- Nanotechnology-based pigment delivery

Conclusion

Pigment studies in ornamental fishes reveal that body coloration is a complex and multifactorial phenomenon, governed by the interaction of biochemical, genetic, physiological, and environmental processes. The remarkable diversity of colors and patterns observed in ornamental species arises from the coordinated function of different pigment types—primarily carotenoids, melanins, pteridines, and purine-based compounds—along with structural mechanisms that influence light reflection and perception.

In addition to their economic relevance, pigment studies contribute to broader biological understanding, including cellular biology, metabolic pathways, and adaptive

functions such as camouflage, communication, and protection. These insights also support advancements in selective breeding programs aimed at producing new and improved ornamental varieties with superior coloration patterns.

In conclusion, pigmentation in ornamental fishes is a dynamic and integrative process that combines biochemical pathways, cellular mechanisms, and environmental interactions. A comprehensive understanding of these factors is essential for enhancing ornamental fish quality, supporting sustainable aquaculture practices, and advancing scientific knowledge in fish biology.

References

- 1) Bagnara, J.T., Matsumoto, J., Ferris, W., Frost, S.K., Turner, W.A., & Tchen, T.T. (1979). *Common origin of pigment cells*. *Science*, 203(4385), 410–415.
- 2) Fox, D.L. (1976). *Animal Biochromes and Structural Colours*. University of California Press.
- 3) Goodwin, T.W. (1984). *The Biochemistry of Carotenoids*. Chapman and Hall, London.
- 4) Torrissen, O.J., Hardy, R.W., & Shearer, K.D. (1989). *Pigmentation of salmonids: carotenoid deposition and metabolism*. *Aquaculture*, 79, 363–374
- 5) Gupta, S.K., & Jha, A.K. (2017). *Role of nutrition in fish pigmentation*. *Journal of Aquaculture Research & Development*, 8(4).
- 6) Hasegawa, T., & Yamada, S. (1992). *Effects of dietary carotenoids on pigmentation in fish*. *Nippon Suisan Gakkaishi*, 58(2), 293–298.
- 7) Storebakken, T., Sørensen, M., & Bjerkeng, B. (2004). *Impact of dietary astaxanthin on fish pigmentation*. *Aquaculture Nutrition*, 10(3), 175–182.
- 8) Matsuno, T. (2001). *Aquatic animal carotenoids*. *Fisheries Science*, 67(5), 771–783.
- 9) Fujii, R. (2000). *The regulation of motile activity in fish chromatophores*. *Pigment Cell Research*, 13(5), 300–319.

A Path to Smart Farming: Exploring Opportunities and Challenges of Artificial Intelligence (AI) in Aquaculture

V. Pattukumar

Assistant Professor, PG & Research Department of Zoology, S.T. Hindu College, Nagercoil

Corresponding Author Email: pattubiotech@gmail.com

Abstract

Over the past few decades, the demand for aquatic products has significantly increased. Additionally, both producers and customers are increasingly taking quality assurance into account. As a result, fish producers are investigating every avenue to increase profitability and productivity. In addition to increasing producers' profitability, keeping an eye on the health and behaviour of fish during cultivation may lessen the risk of serious losses due to illness and stressful situations. By greatly increasing productivity and efficiency for sustainable agriculture, artificial intelligence (AI) is transforming aquaculture. Precision AI technology enable aqua farmers successful culture, optimize crop schedules, choose the right healthy seeds depending on the weather, forecast the weather, control the pathogen, monitor water quality, soil, and analyse data. AI-powered solutions accelerate market delivery, boost yields with less resource, and enhance crop quality. Furthermore, using state-of-the-art equipment like drones, sensors, and artificial intelligence, monitoring water quality, aquatic animal health, and animal behaviour on farms has become incredibly efficient. Effective technology adoption in aquaculture is facilitated by summarizing research and development in aquatic animal health and monitoring. The aquaculture sector is expanding and becoming more efficient as a result of the ongoing development and use of this cutting-edge technology in industrialized nations, which benefits local farmers and consumers. However, there is a shortage of knowledge about these technologies among educators and farmers in developing nations. The creation of standardized diagnostic datasets, lightweight and farmer accessible tools, and integrative frameworks that connect AI with aquaculture knowledge and sustainable farming practices must be given top priority in future research due to the advantages for fish health. This chapter emphasizes that AI is a critical breakthrough for improving fish health and welfare in contemporary aquaculture, not only an auxiliary tool. It offers an aquaculture-focused viewpoint with the goal of directing scholars, professionals, and researchers toward the ethical and successful integration of AI in aquatic animal health management. This chapter takes a narrative approach to summarize

recent developments and future paths in AI-based fish illness diagnosis and management. It also looks at AI's position in aquaculture, describing its uses and advantages, and emphasizes the difficulties and promise of AI in farming.

Keywords: *Internet of Things (IoT); Water quality monitoring; optical sensors; Machine learning (ML); algorithms; Support vector machines (SVM)*

Introduction

Aquaculture is becoming a "smart farming" ecosystem thanks to technologies like the Internet of Things (IoT), artificial intelligence (AI), computer vision, robots, and cloud computing. The combination of machine learning (ML) with the Internet of Things (IoT) presents a paradigm shift in the monitoring and evaluation of water quality. Both farmers and customers are paying more attention to the aquaculture industry's quality assurance these days. The value, cost, and best used before date of fish and fishery products are determined by their quality, freshness, and authenticity (Menesatti *et al.*, 2010). According to Mathiassen *et al.* (2011), fish farmers are therefore investigating every avenue to increase the output and profitability of their products. As of 2020, aquaculture accounted for over 53% of all fish and invertebrate output and 97% of all seaweed manufacturing worldwide, making it one of the fastest-growing food industries. In 2019, the estimated worldwide production of farmed salmon increased by 7% to slightly more than 2.6 million tons of market share. In commercial farms, artificial intelligence (AI) and smart sensing technologies are quickly changing aquatic animal health and water quality management from reactive treatment to proactive, real-time disease prevention. Experience from throughout the world demonstrates that well-designed AI systems may increase feed and resource efficiency at scale, minimize disease-related economic losses by up to half, and achieve diagnostic accuracy above 90%. The main point is evident to an audience that deals with the industry: AI is now a useful productivity tool that can be used in shrimp ponds, fish cages, and aquaculture. Fish health status has a hazardous effect on fish both in the natural world and in aquaculture. Diseases are widely acknowledged as one of the biggest threats to aquaculture's financial success. A wide variety of infectious organisms, including bacteria, viruses, protozoa, and metazoan parasites, can cause diseases in fish. Aquaculture is a cutting-edge farming method that enhances fish and seafood output by utilizing automation, digital technologies, and data-driven decision-making. It focuses on real-time monitoring, measurement, and control of farming activities to boost profitability, sustainability, and production, much like precision

agriculture. Networks of in situ sensors are used by IoT systems to continually record many data, including turbidity, pH, conductivity, dissolved oxygen, and surrogate measurements for pathogens (Essamlali *et al.*, 2024). When certain image processing is used in conjunction with disease detection, good features can be extracted. For several study areas, including computer vision and artificial intelligence, image segmentation becomes essential. Segmentation is a common image processing technique. Support vector machines (SVM), one of the most widely used supervised machine learning approaches, have provided practical answers for numerous classification problems in a variety of domains. It is an effective classification technique that produces high-quality predictions for unlabeled data. In order to distinguish between healthy and dengue-infected human blood sera, the authors of LeCun *et al.* (2015) developed an SVM model based on three kernel functions. Another SVM architecture for image classification was developed in (A. F. Agarap 2017), wherein neural network (CNN) and SVM are combined to replicate the architecture. In many situations, SVM offers exceptional accuracy.

Data Analysis and Technical Complexity

Due to Researchers' inexperience, a number of conventional techniques, such as the manual identification of animal diseases and carriers and statistical calculations to estimate quantity and forecast production and loss of production, were typically cumbersome and led to human error. The capacity of technology to learn from experiences is known as machine learning. It makes the hidden patterns and correlations between the horticultural parameters pH, salinity, DO, etc. visible. Artificial Neural Networks (ANN), SVM Regression and Logistic Regression, neural network recognition technology, Support Vector Machine (SVM) (Singh and Gupta, 2016), fuzzy technology for recognition, etc. are the most popular and related machine learning techniques in forecasting diseases and water quality parameters in which weather data are analyzed. The best approaches focused on the association of image processing and machine learning. A diagnostic method for a particular fish ailment known as Epizootic Ulcerative Syndrome (EUS) was proposed by Malik *et al.*, 2017. This illness is brought on by the fungus *Aphanomyces invadans*. In this case, they used combination methods that classify using a machine learning algorithm (neural network) after combining Principal Component Analysis (PCA) and Histogram of Oriented Gradients (HOG) with features from the Accelerated Segment Test (FAST) feature detector.

The FAST-PCA-NN sequence yields an accuracy of 86% through the classifier, while the HOG-PCA-NN sequence yields a lower accuracy of 65.8%. Kidney stone detection is a delicate subject that (Verma *et al.*, 2017) suggested. The authors of this work use segmentation and morphological procedures to identify the ROI (region of interest) for the SVM classification method. Following the use of this technique, they examined the kidney stone photos with some challenges, including low image quality and kidney stone similarities. A device-free present identification and localization method with SVM assistance was presented by Zhou *et al.* 2018. In this case, the SVM classifier employing the CSI (channel state information) fingerprint allows the detection method to identify human presence. Trojan detection in hardware relies on SVM-based methods. Analyze how to identify trojans using the software they have built. Their net lists include many hardware Trojan types that have been examined for both normal and aberrant behaviour in SVM analysis. Uttarkhand's Himachal Pradesh state is where the system was created. The authors of Shinde and Kulkarni (2017) demonstrated that the current systems are inexpensive and unreliable. They provided information on the use of IoT and machine learning in precision agriculture to forecast crop diseases. They suggested an IoT and machine learning-based system concept.

They collected data using environmental sensors, such as temperature and humidity. Following the creation of the final product, an SMS was sent to the nearby farms. The writers of Huang *et al.*, (2018) created intelligent communications using machine learning techniques. Intelligent machine learning techniques have been applied to enhance the QoS (quality of service) of scarce wireless resources. Sensor devices placed in the agricultural fields were used to record environmental factors including temperature and humidity, which were then transmitted to the central gateway. The information gathered aids in determining the degree and danger of blight. The significance of smart mobile phones for gathering agronomy facts about various parameters, such as soil moisture, humidity, temperature, etc., The advantages of smartphones in the agriculture sector are emphasized in the same piece. Through surveys and interviews, the authors surveyed a variety of farmers to learn about their preferences. They conclude that farmers are interested in using smartphones to obtain information on current agricultural data and analysis after completing the process. The best way to accomplish goals is to find appropriate ways to use continuously recorded data. It explained the value of machine learning, neural networks, and regression analysis in making decisions.

Computer Vision Technology

In situations where the inspection duty is similar in these industries or where an image processing algorithm or method could potentially be helpful in vision-based technologies, the postharvest aquaculture industry is also described. Commercial imaging technologies are also discussed to complete the picture, along with a few non-imaging technologies that offer different approaches. The goal of this section is to highlight areas of computer vision research and development that have made some progress but have not yet developed into practical tools for aquaculture. Examples of these studies include those that have used computer vision in the lab, such as to investigate the effects of diets or chemicals on fish behavior or quality.

The technology has a wide range of possible uses in aquaculture that could enhance production efficiency or product quality. MVSs with their basic concept and principles in fishery. Recent functions of image analysis in fish industry presented in the current review can be used to consider the fundamental methods and technologies associated with computer vision (Saberioon et al., 2017). Computer vision systems track fish behavior, calculate biomass, and identify anomalies using underwater cameras and image analysis. Fish size can now be precisely measured and feeding needs can be estimated thanks to technologies like YOLOv8-based image recognition. Additionally, a research or development procedure is required to advance the technology.

Smart Aquaculture in Monitoring Water Quality Parameter:

The way aquatic ecosystems are observed and managed has significantly changed in recent years due to the incorporation of Internet of Things and machine learning technology into water quality monitoring. Aquaculture is one of the many industries where IoT has found use. With intelligent equipment and real-time connected water quality monitoring capabilities, the use of IoT in aquaculture has created a new trend for the sustainable development of this business and improved working conditions Rahman et al., 2020. The primary element that influences aquaculture's success and effective management is water quality. Numerous water quality characteristics, including temperature, turbidity, carbon dioxide, pH, alkalinity, ammonia, nitrite, and nitrate, are important for the survival and growth of cultured species, either directly or indirectly. The most reliable of these are pH, temperature, and dissolved oxygen. An IoT aquaculture system consists of different layers (Table 1): the Physical Layer, Monitoring Layer, and Virtual Layer. The smart farm industry and intelligent aquaculture have emerged as an inevitable trend to reduce labour costs,

improve operational efficiency, and boost productivity. Future IoT-based solutions may potentially seek to avoid production loss and identify fish infections. It is important to note that while aquaculture has made significant progress in developing intelligent and high-precision fish farming through the use of artificial intelligence devices, fully automated systems still face numerous obstacles. Due to the unique nature of aquaculture and its products, labour management is risky since fish farming still requires a certain level of observation, information analysis, and decision-making. Nonetheless, a variety of intelligent devices, including the technologies listed above, are in charge of keeping an eye on fish farming environments. Production robotics, data and information sorting, and energy-efficient processing equipment will all continue to significantly automate various phases of fish farming operations. These can be used in aquaculture for behavior control, mass estimate, and fish identification. Furthermore, novel sensors that combine many functions into a sensor or multifunctional sensor with high reliability and a wide variety of applications must be developed. Numerous research have been conducted to date regarding the use of intelligent information, such as AI and IoT, in aquaculture in various settings, including ponds, hatcheries, and cages. The Raspberry Pi, which is used to examine real-time monitoring systems in aquaculture with temperature, dissolved oxygen, pH, and ammonia sensors, was employed by Chavan *et al.*, 2018. The gathered information is transmitted to the aqua farmer mobile device after being stored in a cloud computing system. (Kim *et al.*, 2018) installed a recirculating aquaculture system (RAS) with a MICOM controller, MQTT protocol, and water quality monitoring sensors in a smart fish farm. In order to solve the current issues, Al-Hussaini *et al.*, 2018 additionally concentrate on automated data collection and monitoring systems for RAS employing fog computing technologies and inexpensive systems with Raspberry Pi. The Raspberry Pi 3 is connected to a temperature sensor (LM35DT) via an ADC MCP3002, a water level sensor (HC-SR04), and a Rpi camera module to create the data collection system for RAS (RaspDAQ). A smart fish farm with two tanks—a balancing tank and a recirculating aquaculture system (RAS)—was proposed by Shin *et al.* [69]. The proportional integral derivative (PID) controller, which is linked to water temperature, dissolved oxygen (DO), potential hydrogen (pH), and water level controlling sensors, regulates the water effluence in aquaculture systems. The Message Queue Telemetry Transport (MQTT) protocol is used for both remote control and real-time monitoring, and the measured huge data is kept on lab servers. A smart, real-time water monitoring system utilizing IoT devices was proposed by Monirul *et al.*, 2019. Sensors for temperature, dissolved oxygen, turbidity, pH, water level, and CO₃ gas were used in the construction of

the system, along with an Arduino and an Internet of Things platform. Nocheski *et al.*, 2018 provided an update on a working Internet of Things (IoT) system to automatically monitor the water quality in cultured systems in order to deploy IoT systems in ponds. Temperature, light intensity, and water level are just a few of the critical water quality metrics that the Internet of Things system can measure. Additionally, it has a tiny computer board that processes and analyzes the data gathered from those sensors. An Internet of Things system to automatically monitor these environmental characteristics was proposed by Darmalim *et al.*, 2019. The user can obtain information from the Internet of Things device, and the water quality metrics are gathered in real time from the aquaculture system and instantly updated on the online. A Python framework is used in its development. By using the web application to obtain data that might display environmental conditions, users can directly learn what transpired in the pond and promptly take appropriate action. NodeMcuESP8266, ADC Ads1115, a logic converter, a temperature and dissolved oxygen sensor, a pH sensor, a turbidity sensor, a TDS sensor, a DC-DC converter, a power supply, and a web application for a Python web development framework that is linked to a MySQL database are the electrical components.

Table1 : Technology Building Blocks for Smart Health Management in aquaculture

Sources	Model Technologies	Parameter
Biosensors	Biosensors use biological elements, enzymes and antibodies to detect target substances.	Rapidly detect small signal changes to detect the organism. changes in parameters like current, resistance or heat.
Apps based	Farm management apps system combining health, feeding, and animal monitoring	Detection of animal health status, and water quality
Data capture	DO Probe based system, Underwater and surface cameras, fixed CCTV, wearable sensors, IoT temperature, pH, Nitrites, Nitrate, ammonia	Collect continuous visual and environmental data from ponds, cages, RAS units, and shrimp/Fish hatcheries.

Connectivity	Wi-Fi, cellular networks, LoRa, edge gateways	Transmit data securely from remote farms to local servers or cloud platforms in near real time.
AI/ML models	ML (SVM, Random Ecosystem) Convolutional neural networks (CNNs), YOLO detectors, recurrent networks (RNN/LSTM), transformers	Video streams, Analyze the images, and time-series, sensor data for disease signatures and risk patterns.
Applications	Dashboards, mobile advisory apps, automated alerts, decision-support systems	Present results to farm staff , Technicians, veterinarians in simple, actionable formats.
Drones	Aerial Imaging using unmanned aerial vehicles (UAVs) is a relatively new concept.	Farm monitoring is crucial in in-land and off-shore aquaculture management practices. Larger size aquaculture farm monitoring

NGS technology and fish features analysis

Generally speaking, the primary method for determining fish size is computer vision or machine vision. Next-generation sequencing (NGS) technology is a useful diagnostic tool used to sequence viruses to better understand their evolution and identification. Images are captured using a camera coupled to a fish measurement system. A method that uses image processing algorithms to detect and quantify the size of various fish species was proposed by White *et al.*, 2006. A variety of fish species, including *Hippoglossoides platessoides*, *Solea vulgaris*, *Microstomus kitt*, and *Pleuronectes platessa*, were photographed. It has been effectively used to diagnose diseases in humans, plants, and animals. However, this method has a number of drawbacks, including the need for complex machinery, increased expenses, and lengthier turnaround times.

In order to tackle the difficulties of NGS, Oxford Nanopore Technologies unveiled a new third-generation sequencing technology in 2014. This technology uses a compact,

portable device that weighs only 90 g to provide long and accurate sequencing reads in real time. This approach adds a sequencing library using a flow cell sequencing chip. Reliable, onsite findings are obtained when the DNA/RNA passes through the nanopore, causing a variable current that is subsequently decoded into base pairs. Disease diagnosis is one of the many areas in which nanopore sequencing has been used recently. Costa and associates carried out an experiment with the goal of developing methodology tools to directly measure size, identify sex, and diagnose abnormal performance of seabass (*Dicentrarchus labrax* L.) in online conditions at an aquaculture farm.

In order to monitor growth status and improve stock management, aqua farmers must have information on individual fish characteristics such as length, weight, sex, maturity, and skin colour at various phases of growth. Because selection is the basis for breeding and producing high-quality fish through repetitive sorting and grading of numerous fish based on gender and quality throughout grow-out, these individual fish characteristics are more important for fish quality assessment (Gomelsky *et al.*, 1995; Zion *et al.* 2008). The most popular conventional methods for researching and tracking individual fish characteristics are extractive methods (Trenkel & Cotter 2009). These methods provide precise information about individual fish, but they are more frequently employed for inventories and small-scale observations than for monitoring. Additionally, extractive methods like net casting are intrusive, which is unpopular when it comes to monitoring. Aquatic creatures' skin colour has a significant role in revealing their physiological, behavioral, and ecological state (Pavlidis *et al.* 2006). Fish skin pigmentation patterns show the welfare of the fish as well as growth rates. Fish that are under stress experience a variety of biochemical changes, all of which manifest externally as variations in skin colour.

Additionally, skin colour offers helpful information for effective nutrition planning. Ornamental fish's skin colour even affects their market value. Fish skin color has been measured and determined by researchers using various optical sensors. (Pavlidis *et al.*, 2006) measured the three-dimensional aspects of colour appearance using a portable spectrophotometer. The Entire Colour Index (ECI) is a recently developed index for displaying fish skin color patterns. (Wallat *et al.*, 2005) showed how an MVS might be used to measure the skin color of live goldfish (*Carassius auratus*) objectively and to improve the feeding plan to get the best skin color. (Zat'ková, *et al.*, (2011) estimated changes in catfish (*Silurus glans*) skin colour using an MVS.

They demonstrated the viability of using MVS to track changes in skin tone brought on by dietary modifications. (Colihueque *et al.* 2011) created a technique for classifying cultured rainbow trout (*Oncorhynchus mykiss*) that uses computer-based picture analysis to estimate skin color, spottiness, and blackness. An algorithm for determining fish skin color in Vis bands was developed. (Urban *et al.*,2012). Additionally, their method can determine the fish skin's dominant wavelength. Fish size is another crucial characteristic that farmers must be aware of. Farmers can make better decisions about grading, harvesting, and when to harvest the stock based on the size of the fish (Beddow *et al.*, 1996). Furthermore, stereo vision systems have been used to forecast reef fish with 95% accuracy (Harvey *et al.*, 2001) and salmon biomass with less than 4% inaccuracy (Beddow *et al.* 1996). Several picture properties have been identified by researchers as viable indicators for fish mass measurement. For instance, Zion *et al.* (1999) employed fish area to estimate the fish mass of St. Peter (*Oreochromis*), carp (*Cyprinus carpio*), and grey mullet (*Mugil cephalus*) from images with 95%, 99%, and 99% accuracy, respectively. By manually (Tillett *et al.* 2000) or automatically (Lines *et al.* 2001) fitting Point-Distribution Mode (PDM) for fish picture segmentation, researchers were able to estimate fish length, which was then used to determine size and mass.

To monitor growth rate and schedule precise feeding to prevent overfeeding, which is one of the main causes of waste and water pollution, farmers require information on fish mass and size at various stages of growth. To track growth rate and make plans, farmers want information on fish mass and size at various phases of development. Catching and sedating fish is the most common way to measure size. Although this approach is accurate, it is time-consuming, labor-intensive, intrusive, and only represents a small percentage of the population (Lines *et al.* 2001). Over the past three decades, researchers have been drawn to image processing in aquaculture as a non-invasive technique for measuring fish mass and size (Zion 2012). Fish size and length can be measured remotely using MVS without stressing the fish. By measuring fin-to-fin length, height, and length dimension, (Dios *et al.*, 2003) reported using a stereovision system to predict mass with 96% accuracy in nurseries and 95% accuracy in marine cages. However, because of poor image quality and high fish density, the accumulation of these techniques would be reduced when live fish photographs were obtained from commercial fish cages. (Torisawa *et al.*, 2011) estimated the length of Pacific bluefin tuna (*Thunnus orientalis*) using a stereovision system placed at various depths in a commercial marine cage. For tuna allocated up to 5.5 meters from the cameras, they reported

a 5% fish length estimation inaccuracy. Fish mass and size have also been estimated using a dual-camera optical ranging system. Costa et al. (2006) created a system that uses an artificial neural network (ANN) and two underwater cameras to quantify mass and size. They measured fish size with remarkable precision. However, measuring little fish significantly reduces accuracy. Identification of fish species is crucial for fishery management. For improved selling in a polyculture fish farming system, farmers must sort caught fish by species and size. Additionally, fish species identification can be used by farmers for stock assessment (Hoggarth 2006) and feeding techniques (Gerking 1994; Alcaraz *et al.* 2015).

Automated Feeding System

Automated feeding avoids water pollution from overfeeding and cuts down on feed waste. Feed conversion ratios and overall profitability can be enhanced by intelligent feeding systems. By automatically identifying and learning samples from data, algorithmic models are trained to perform particular tasks quickly and accurately. This technique can be divided into three categories: supervised, semi-supervised/unsupervised, and reinforcement learning; supervised is the most widely used approach (Erickson *et al.*, 2017). Utilize an algorithm system to gain experience by using a labeled dataset for training. Data analysis from these encounters differs from that of the trained ones.

These algorithms will examine new data resources and images to determine whether or not they are the target images after completing trained phases (Moore *et al.*, 2019). Feeding is one of the biggest issues in aquaculture. Depending on the eating habits of the cultivated species, farmers will either distribute food around the pond or at a specific spot. As a result, IoT must be used in the feeding system to regulate the feeding quantity and feed automatically. This has several advantages, including minimizing water quality contamination in aquaculture, managing the number of leftovers, and saving labour. A few studies have been carried out. An effective semi-automatic method that promotes the healthy growth of aquatic organisms in aquaculture, Using sensors like a pH and temperature sensor, this study is carried out not only for a feeding system but also for water quality factors in a cultured system. Every time the quality metrics deviate from the typical range, the cultivator is notified by the GSM module. The fish is automatically fed according to its needs by the feeding system. An Arduino Uno, a DS18B20 Temperature Sensor, a pH sensor, and a SIM900A-GSM module make up the majority of the system. An IoT-based water quality system in a smart aquarium was proposed (Daud *et al.* 2020). Maintaining freshwater levels

in aquarium tanks that are appropriate for fish habitats and feeding conditions is the aim of this study. MEGA and NodeMCU controllers were used in the creation of this system, which is operated via a smartphone. The system is meant to use Arduino. The smartphone and the controller are connected via Wi-Fi communication on the NodeMCU to regulate the operation. Every piece of water quality data that has been gathered is shown on an LCD. Classification, regression, clustering, and DR (dimensionality reduction) are the four primary tasks in machine learning. Regression and classification are two of these features of supervised learning models. While DR falls under both supervised and unsupervised learning, clustering falls under unsupervised learning.

Conclusion

An innovative approach to sustainable and intelligent fish farming is the smart farming aquaculture system. By enhancing monitoring, automation, and decision-making procedures, technologies including IoT, AI, robots, computer vision, and cloud computing are transforming aquaculture. Various uses of MVSs and optical sensors in fisheries, along with their fundamental ideas and concepts. The current chapter's discussion of recent applications of image analysis in the fish industry can be used to examine the basic techniques and technologies related to computer vision, which would offer a practical method for automated, impartial, quick, and hygienic inspection of fish and fish products. The prospects for sustainable fish production and increased profitability are significant, despite obstacles such as high implementation costs, technical complexity, sensor dependability, and cyber security issues. The employment of optical and imaging technology to maximize fish production costs and enhance their quality in numerous ways appears to be the general trend. These systems could play a key role in automated behavioural research, feature analysis, aquatic food control, and processing activities. Smart aquaculture is anticipated to be crucial to maintaining global food security, environmental sustainability, and the modernization of aquaculture businesses globally as technology become more widely available and reasonably priced.

References

- 1) F. Agarap (2017) *An architecture combining convolutional neural net-work (cnn) and support vector machine (svm) for image classification. arXiv preprint arXiv:1712.03541,*

- 2) Alcaraz, C., Gholami, Z., Esmaili, H. R., & García-Berthou, E. (2015). *Herbivory and seasonal changes in diet of a highly endemic cyprinodontid fish (Aphanius farsicus)*. *Environmental Biology of Fishes*, 98(6), 1541-1554.
- 3) Al-Hussaini, K.; Zainol, S.M.; Ahmed, R.B.; Daud, S. *IoT Monitoring and Automation Data Acquisition for Recirculating Aquaculture System Using Fog Computing*. *J. Comput. Hardw. Eng.* 2018, 1.
- 4) Beddow TA, Ross LG, Marchant JA (1996) *Predicting salmon biomass remotely using a digital stereo-imaging technique*. *Aquaculture* 146(3): 189–203.
- 5) Chavan, M.S.; Patil, V.P.; Chavan, S.; Sana, S.; Shinde, C. (2018) *Design and Implementation of IoT Based Real Time Monitoring System for Aquaculture using Raspberry Pi*. *Int. J. Recent Innov. Trends Comput. Commun.* 6, 159–161.
- 6) Colihueque N, Parraguez M, Estay FJ, Diaz NF (2011) *Skin color characterization in rainbow trout by use of computer-based image analysis*. *North American Journal of Aquaculture* 73(3): 249–258.
- 7) Costa C, Loy A, Cataudella S, Davis D, Scardi M (2006) *Extracting fish size using dual underwater cameras*. *Aquaculture Engineering* 35: 218–227.
- 8) Darmalim, U.; Darmalim, F.; Darmalim, S.; Ahmad, A.; Budiarto, A.; Mahesworo, B.; Pardamean, B. *IoT solution for intelligent pond monitoring*. In *Proceedings of the 3rd International Conference on Eco Engineering Development, Solo, Indonesia, 13–14 November 2019; Volume 426, pp. 1–10*.
- 9) Daud, A.K.P.M.; Sulaiman, N.A.; Yusof, Y.W.M.; Kassim, M. (2020) *An IoT-Based Smart Aquarium Monitoring System*. In *Proceedings of the 2020 IEEE 10th Symposium on Computer Applications & Industrial Electronics (ISCAIE), Penang, Malaysia, 18–19 April; pp. 277–282*.
- 10) Dios JRM-D, Serna C, Ollero A (2003) *Computer vision and robotics techniques in fish farms*. *Robotica* 21(3): 233–243
- 11) Erickson, B.J.; Korfiatis, P.; Akkus, Z.; Kline, T.L. (2017) *Machine learning for medical imaging*. *Radiographics*, 37, 505–515.
- 12) Essamlali, I., Nhaila, H., & Khaili, M. E. (2024). *Advances in machine learning and IoT for water quality monitoring: A comprehensive review*. *Heliyon*, 10(6), e27920–e27920.
- 13) Gerking SD (1994) *Feeding Ecology of Fish*. Academic Press, SanDiego.

- 14) Gomelsky B, Cherfas NB, Hulata G, Ben-Dom N (1995) Color variability in normal and gynogenetic progenies of ornamental (Koi) common carp. *Aquaculture* 137: 99–102.
- 15) Harvey E, Fletcher D, Shortis M (2001) A comparison of the precision and accuracy of estimates of reef-fish lengths determined visually by divers with estimates produced by a stereovideo system. *Fishery Bulletin* 99(1): 63–71.
- 16) Hoggarth DD (2006) *Stock Assessment for Fishery Management*. FAO, Rome, Italy
- 17) Huang R, Lai Y, Tsao C et al (2018) Applying convolutional networks to underwater tracking without training. *IEEE Int Conf Appl Syst Invent* 2018:342–345.
- 18) Kim, B.J.; Shin, K.J. (2018) Development of Smart Fish Farming System Based on IoT Using Wasted Warm Water Energy. *J. Inst. Control Robot. Syst.*, 24, 155–163.
- 19) Lines JA, Tillett RD, Ross LG, Chan D, Hockaday S, McFarlane NJB (2001) An automatic image-based system for estimating the mass of free-swimming fish. *Computers and Electronics in Agriculture* 31(2): 151–168
- 20) Malik, S.; Kumar, T.; Sahoo, A.K. (2017) A novel approach to fish disease diagnostic system based on machine learning. *Adv. Image Video Process.* 5, 49–57.
- 21) Mathiassen JR, Misimi E, Bondø M, Veliyulin E, Østvik SO (2011) Trends in application of imaging technologies to inspection of fish and fish products. *Trends in Food Science and Technology* 22(6): 257–275.
- 22) Menesatti P, Costa C, Aguzzi J (2010) Quality evaluation of fish by hyperspectral imaging. *Hyperspectral Imaging for Food Quality Analysis and Control*, 1st edn, pp. 273–294. Academic Press, San Diego.
- 23) Monirul, I.; Mohammad, A.K.; Farzana, I.J. (2019,) Aqua Fishing Monitoring System Using IoT Devices. *IJISSET Int. J. Innov. Sci. Eng. Technol.* 6, 108–114.
- 24) Moore, M.M.; Slonimsky, E.; Long, A.D.; Sze, R.W.; Iyer, R.S. (2019) Machine learning concepts, concerns and opportunities for a pediatric radiologist. *Pediatr. Radiol.* 49, 509–516
- 25) Nocheski, S.; Naumoski (2018)A. Water monitoring iot system for fish farming ponds. *Industry 4.0* 3, 77–79.
- 26) Pavlidis M, Papandroulakis N, Divanach P (2006) A method for the comparison of chromaticity parameters in fish skin: preliminary results for coloration pattern of red skin Sparidae. *Aquaculture* 258(1–4): 211–219.

- 27) Rahman, M.; Bapery, C.; Hossain, M.J.; Hassan, Z.; Hossain, G.M.J.; Islam, M. (2020) *Internet of Things (IoT) based water quality monitoring system*. *Int. J. Multidiscip. Curr. Educ. Res. (IJMCER)* 2(4), 168–180.
- 28) Saberioon, M., Gholizadeh, A., Cisar, P., Pautsina, A., & Urban, J. (2017). *Application of machine vision systems in aquaculture with emphasis on fish: state-of-the-art and key issues*. *Reviews in Aquaculture*, 9(4), 369-387.
- 29) Shinde, S.S. and Kulkarni, M., 2017, December. *Review paper on prediction of crop disease using IoT and machine learning*. In *2017 International conference on transforming engineering education (ICTEE)* (pp. 1-4). IEEE.
- 30) Singh, J. and Gupta, V., 2016. *Text stemming: Approaches, applications, and challenges*. *ACM Computing Surveys (CSUR)*, 49(3), pp.1-46.
- 31) Tillett R, McFarlane N, Lines J (2000) *Estimating dimensions of free-swimming fish using 3D point distribution models*. *Computer Vision and Image Understanding* 79(1): 123–141.
- 32) Trenkel VM, Cotter J (2009) *Choosing survey time series for populations as part of an ecosystem approach to fishery management*. *Aquatic Living Resources* 22(02): 121–126.
- 33) Urban, J., Štys, D., Sergejevová, M., & Masojídek, J. (2013). *Expertomica Fishgui: comparison of fish skin colour*. *Journal of Applied Ichthyology*, 29(1), 172-180.
- 34) Verma, J., Nath, M., Tripathi, P., & Saini, K. K. (2017). *Analysis and identification of kidney stone using Kth nearest neighbour (KNN) and support vector machine (SVM) classification techniques*. *Pattern Recognition and Image Analysis*, 27(3), 574-580.
- 35) Wallat G, Lazur A, Chapman F (2005) *Carotenoids of different types and concentrations in commercial formulated fish diets affect color and its development in the skin of the red oranda variety of goldfish*. *North American Journal of Aquaculture* 67 (1): 42–51
- 36) White DJ, Svellingen C, Strachan NJC (2006) *Automated measurement of species and length of fish by computer vision*. *Fisheries Research* 80(2–3): 203–210
- 37) Začková, I., Sergejevová, M., Urban, J., Vachta, R., Štys, D., & Masojídek, J. (2011). *Carotenoid-enriched microalgal biomass as feed supplement for freshwater ornamentals: albinic form of wels catfish (Silurus glanis)*. *Aquaculture Nutrition*, 17(3), 278-286.
- 38) Zion B (2012) *The use of computer vision technologies in aquaculture – a review*. *Computers and Electronics in Agriculture* 88(C): 125–132
- 39) Zion B, Shklyar A, Karplus I (1999) *Sorting fish by computer vision*. *Computers and Electronics in Agriculture* 23(3): 175–187.

ABOUT THE EDITORS

Dr. M. Sini Margret

Dr. M. Sini Margret is an Assistant Professor in the Department of Advanced Zoology and Biotechnology, with specialization in Malacology and Aquaculture. She obtained her Ph.D. from Manonmaniam Sundaranar University and has nearly a decade of teaching and research experience, including long service at Nanjil Catholic College of Arts and Science, where she also served as Head of the Department of Zoology. She has published several research papers, book chapters, and conference proceedings in national and international journals, and actively participates in seminars, conferences, and faculty development programmes.

Dr. I. Vasudhevan

Dr. I. Vasudhevan is an Assistant Professor and Research Supervisor in the PG & Research Department of Zoology, with specialization in Nutrition, Biochemistry and Aquaculture. He obtained her Ph.D. from V.O.Chidambaram College, Thoothukudi, Manonmaniam Sundaranar University. In 4 years Research experience in Fisheries College and Research Institution, Thoothukudi and 16 years teaching and research Experience in Vivekananda College, Agasteeswaram, Kanyakumari. He has published 80 research papers, book chapters and conference proceedings in national and international journals and actively presented a research article and participates in seminars, conferences, resource person, chair person, faculty development programmes and doctoral committee member.

Dr. M. Santhiya

Dr. M. Santhiya is an accomplished academician and researcher currently serving as an Assistant Professor in the Department of Biotechnology at S.T. Hindu College, Nagercoil. With a strong foundation in life sciences, she specializes in Molecular Biology and Plant Biology, demonstrating a deep commitment to advancing knowledge in these dynamic fields. She obtained her Doctor of Philosophy (Ph.D.) degree from Manonmaniam Sundaranar University, where she developed a solid research background and analytical expertise.

Dr. S. Rinna Hamlin

Dr. S. Rinna Hamlin is an Assistant Professor in the Department of Zoology, Nesamony Memorial Christian College (NMCC), Marthandam, Kanyakumari District, Tamil Nadu. Dr. S. Rinna Hamlin received his Ph.D. from Manonmaniam Sundaranar University, Thirunelveli, Tamil Nadu. She has published several research papers and conference proceedings in national and international journals and actively participates in seminars, conferences, and faculty development programmes.

ISBN 978-81-69360-86-9



9 788169 360869



Dr.BGR
Publications